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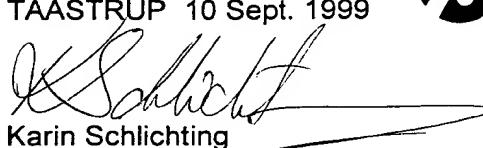
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Phytases (*myo*-inositol hexakisphosphate phosphohydrolases; EC 3.1.3.8) are 5 enzymes that hydrolyze phytate (*myo*-inositol hexakisphosphate) to *myo*-inositol and inorganic phosphate and are known to be valuable feed additives.

A phytase was first described in rice bran in 1907 [Suzuki et al., Bull. Coll. Agr. Tokio Imp. Univ. 7, 495 (1907)] and phytases from *Aspergillus* species in 1911 [Dox and Golden, J. Biol. Chem. 10, 183-186 (1911)]. Phytases have also been found in wheat bran, 10 plant seeds, animal intestines and in microorganisms [Howsen and Davis, Enzyme Microb. Technol. 5, 377-382 (1983), Lambrechts et al., Biotech. Lett. 14, 61-66 (1992), Shieh and Ware, Appl. Microbiol. 16, 1348-1351 (1968)].

The cloning and expression of the phytase from *Aspergillus niger* (ficum) has been described by Van Hartingsveldt et al., in Gene, 127, 87-94 (1993) and in European Patent 15 Application, Publication No. (EP) 420 358 and from *Aspergillus niger* var. awamori by Piddington et al., in Gene 133, 55-62 (1993).

Cloning, expression and purification of phytases with improved properties have been disclosed in EP 684 313. However, since there is a still ongoing need for further improved phytases, especially with respect to their thermostability, it is an object of the present 20 invention to provide the following process which is, however, not only applicable to phytases.

A process for the preparation of a consensus protein, whereby such process is characterized by the following steps:

- a) at least three, preferably four amino acid sequences of a defined protein family 25 are aligned by any standard alignment program known in the art;
- b) amino acids at the same position according to such alignment are compared regarding their evolutionary similarity by any standard program known in the art, whereas the degree of similarity provided by such a program which defines the least similarity of the amino acids that is used for the determination of an 30 amino acid of corresponding positions is set to a less stringent number and the parameters are set in such a way that it is possible for the program to determine

from only 2 identical amino acids at a corresponding position an amino acid for the consensus protein; however, if among the compared amino acid sequences are sequences that show a much higher degree of similarity to each other than to the residual sequences, these sequences are represented by their consensus sequence determined as defined in the same way as in the present process for the consensus sequence of the consensus protein or a vote weight of 1 divided by the number of such sequences is assigned to every of those sequences;

10 c) in case no common amino acid at a defined position can be identified by the program, any of the amino acids of all sequences used for the comparison, preferably the most frequent amino acid of all such sequences is selected or an amino acid is selected on the basis of the consideration given in Example 2;

d) once the consensus sequence has been defined, such sequence is back-translated into a DNA sequence, preferably using a codon frequency table of the organism in which expression should take place;

15 e) the DNA sequence is synthesized by methods known in the art and used either integrated into a suitable expression vector or by itself to transform an appropriate host cell;

f) the transformed host cell is grown under suitable culture conditions and the consensus protein is isolated from the host cell or its culture medium by methods known in the art.

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In a preferred embodiment of this process step b) can also be defined as follows:

b) amino acids at the same position according to such an alignment are compared regarding their evolutionary similarity by any standard program known in the art, whereas the degree of similarity provided by such program is set at the lowest possible value and the amino acid which is the most similar for at least half of the sequences used for the comparison is selected for the corresponding position in the amino acid sequence of the consensus protein.

In another preferred embodiment the consensus sequence is used in order to improve 30 a specific protein. In this process first a consensus sequence is determined from a number of highly homologous sequences according to steps a), b) and c) as described above. In a second step the amino acid sequence of another protein which is homologous to the consensus sequence is compared with the consensus sequence and in a third step only those amino acid residues are replaced in the amino acid sequence of the other protein

which clearly differ from the consensus sequence of this protein family calculated under moderately stringent conditions whereas at all positions of the alignment where no preferred single amino acid can be determined under moderately stringent conditions the amino acids of the other protein remain unchanged.

- 5 By using this preferred embodiment the consensus sequence derived from a number of highly homologous sequences is used in order to replace only certain amino acid residues in the protein in such a manner that only those amino acid residues are replaced which clearly and unambiguously differ from the corresponding consensus sequence of this protein family which has been calculated on moderately stringent conditions. At all other 10 positions of the alignment, however, where the method of the present invention is not able to determine clearly a preferred amino acid residue under moderately stringent conditions the amino acid residues of the other protein are maintained unchanged.

- 15 A further preferred embodiment is a process wherein at first a consensus sequence is determined from homologous sequences as described above. In a second step the active center of the protein comprising all amino acid residues that are involved in forming the active center is determined in the consensus sequence and in the sequence of a single homologous protein as well. The single homologous protein may have preferred properties like high specific activity or different pH dependency of enzymatic activity. In a third step some or all amino acid residues that are involved in forming the active centre of the 20 homologous protein are inserted into the backbone of the consensus sequence. The result thereof is a chimeric protein having the active centre derived from a single protein and the backbone of the consensus sequence.

- 25 The active centre of the protein can be determined e.g. by using any analysis of the three-dimensional structure of the protein, e.g. by homology modelling on the basis of a known 3D-structure of a known protein. Frequently the single homologous protein is an enzyme.

- 30 It is furthermore an object of the present invention to provide such a process, wherein the program used for the comparison of amino acids at a defined position regarding their evolutionary similarity is the program "PRETTY". It is more specifically an object of the present invention to provide such a process, wherein the defined protein family is the family of phytases, especially wherein the phytases are of fungal origin.

It is furthermore an object of the present invention to provide such processes, wherein the host cell is of eukaryotic, especially fungal, preferably *Aspergillus* or yeast, preferably *Saccharomyces* or *Hansenula* origin.

It is also an object of the present invention to provide a consensus protein obtainable preferably obtained, by such processes and specifically the consensus protein, which has the amino acid *sequences shown in Figures 2, 4 and 6* or a variant thereof. A "variant" refers in the context of the present invention to a consensus protein with amino acid sequence shown in Figure 2, 5, 7, and 8 wherein at one or more positions amino acids have been deleted, added or replaced by one or more other amino acids with the proviso that the resulting sequence provides for a protein whose basic properties like enzymatic activity (type of and specific activity), thermostability, activity in a certain pH-range (pH-stability) have not significantly been changed. "Significantly" means in this context that a man skilled in the art would say that the properties of the variant may still be different but would not be unobvious over the ones of the consensus protein with the amino acid sequence of Figure 2 itself.

A "mutein" refers in the context of the present invention to replacements of the amino acid in the amino acid sequences of the consensus proteins shown in Figure 2 which lead to consensus proteins with further improved properties e.g. activity. Such muteins can be defined and prepared on the basis of the teachings given in European Patent Application number 97810175.6, e. g. Q50L, Q50T, Q50G, Q50L-Y51N, or Q50T-Y51N. "Q50L" means in this context that at position 50 of the amino acid sequence (Figure 2) the amino acid Q has been replaced by amino acid L.

In addition, a food, feed or pharmaceutical composition comprising a consensus protein as defined above is also an object of the present invention.

In this context "at least three preferably four amino acid sequences of such defined protein family" means that three, four, five, six to 12, 20, 50 or even more sequences can be used for the alignment and the comparison to create the amino acid sequence of the consensus protein. "Sequences of a defined protein family" means that such sequences fold into a three dimensional structure, wherein the alpha-helices, the beta-sheets and beta-turns are at the same position so that such structures are, as called by the man skilled in the art, largely superimposable. Furthermore these sequences characterize proteins which show the same type of biological activity, e.g. a defined enzyme class, e.g. the phytases. As known in the art, the three dimensional structure of one of such sequences is sufficient to allow the modelling of the structure of the other sequences of such a family. An example, how this can be effected, is given in the Reference Example of the present case. "Evolutionary similarity" in the context of the present invention refers to a scheme which classifies amino acids regarding their structural similarity which allows that one amino acid can be replaced by another amino acid with a minimal influence on the overall structure, as this is done e.g.

by programs, like "PRETTY", known in the art. The phrase "the degree of similarity provided by such a program...is set to less stringent number" means in the context of the present invention that values for the parameters which determine the degree of similarity in the program used in the practice of the present invention are chosen in a way to allow the 5 program to define a common amino acid for a maximum of positions of the whole amino acid sequence, e. g. in case of the program PRETTY a value of 2 or 3 for the THRESHOLD and a value of 2 for the PLURALITY can be chosen. Furthermore, "a vote weight of one divided by the number of such sequences" means in the context of the present invention that the sequences which define a group of sequences with a higher 10 degree of similarity as the other sequences used for the determination of the consensus sequence only contribute to such determination with a factor which is equal to one divided by a number of all sequences of this group.

As mentioned before should the program not allow to select the most similar amino acid, the most frequent amino acid is selected, should the latter be impossible the man 15 skilled in the art will select an amino acid from all the sequences used for the comparison which is known in the art for its property to improve the thermostability in proteins as discussed e.g. by

Janecek, S. (1993), *Process Biochem.* 28, 435-445 or
Fersht, A. R. & Serrano, L. (1993), *Curr. Opin. Struct. Biol.* 3, 75-83.
20 Alber, T. (1989), *Annu. Rev. Biochem.* 58, 765-798 or
Matthews, B. W. (1987), *Biochemistry* 26, 6885-6888.
Matthews, B. W. (1991), *Curr. Opin. Struct. Biol.* 1, 17-21.

The stability of an enzyme is a critical factor for many industrial applications. Therefore, a lot of attempts, more or less successful, have been made to improve the 25 stability, preferably the thermostability of enzymes by rational (van den Burg *et al.*, 1998) or irrational approaches (Akanuma *et al.*, 1998). The forces influencing the thermostability of a protein are the same as those that are responsible for the proper folding of a peptide strand (hydrophobic interactions, van der Waals interactions, H-bonds, salt bridges, conformational strain (Matthews, 1993). Furthermore, as shown by Matthews *et al.* (1987), 30 the free energy of the unfolded state has also an influence on the stability of a protein. Enhancing of protein stability means to increase the number and strength of favorable interactions and to decrease the number and strength of unfavorable interactions. It has been possible to introduce disulfide linkages (Sauer *et al.*, 1986) to replace glycine with

alanine residues or to increase the proline content in order to reduce the free energy of the unfolded state (Margarit et al, 1992; Matthews, 1987a). Other groups concentrated on the importance of additional H-bonds or salt bridges for the stability of a protein (Blaber et al, 1993) or tried to fill cavities in the protein interior to increase the buried hydrophobic 5 surface area and the van der Waals interactions (Karpusas et al, 19898). Furthermore, the stabilization of secondary structure elements, especially α -helices, for example, by improved helix capping, was also investigated (Munoz & Serrano, 1995).

However, there is no fast and promising strategy to identify amino acid replacements which will increase the stability, preferably the thermal stability of a protein. Commonly, 10 the 3D structure of a protein is required to find locations in the molecule where an amino acid replacement possibly will stabilize the protein's folded state. Alternative ways to circumvent this problem are either to search for a homologous protein in a thermo- or hyperthermophile organism or to detect stability-increasing amino acid replacements by a random mutagenesis approach. This latter possibility succeeds in only 10^3 to 10^4 mutations 15 and is restricted to enzymes for which a fast screening procedure is available (Arase et al, 1993; Risse et al, 1992). For all these approaches, success was variable and unpredictable and, if successful, the thermostability enhancements nearly always were rather small.

Here we present an alternative way to improve the thermostability of a protein. Imanaka et al (1986) were among the first to use the comparisons of homologous proteins 20 to enhance the stability of a protein. They used a comparison of proteases from thermophilic with homologous ones of mesophilic organisms to enhance the stability of a mesophilic protease. Serrano et al (1993) used the comparison of the amino acid sequences 25 of two homologous mesophilic RNases to construct a more thermostable Rnase. They mutated individually all of the residues that differ between the two and combined the mutations that increase the stability in a multiple mutant. Pantoliano et al (1989) and, in particular, Steipe et al (1994) suggested that the most frequent amino acid at every position 30 of an alignment of homologous proteins contribute to the largest amount to the stability of a protein. Steipe et al (1994) proved this for a variable domain of an immunoglobulin, whereas Pantoliano et al (1989) looked for positions in the primary sequence of subtilisin in which the sequence of the enzyme chosen to be improved for higher stability was 35 singularly divergent. Their approach resulted in the replacement M50F which increased the T_m of subtilisin by 1.8 °C.

Steipe et al. (1994) proved on a variable domain of immunoglobulin that it is possible to predict a stabilizing mutation with better than 60% success rate just by using a 35 statistical method which determines the most frequent amino acid residue at a certain position of this domain. It was also suggested that this method would provide useful results

not only for stabilization of variable domains of antibodies but also for domains of other proteins. However, it was never mentioned that this method could be extended to the entire protein. Furthermore, nothing is said about the program which was used to calculate the frequency of amino acid residues at a distinct position or whether scoring matrices were used as in the present case.

5 It is therefore an object of the present invention to provide a process for the preparation of a consensus protein comprising a process to calculate an amino acid residue for nearly all positions of a so-called consensus protein and to synthesize a complete gene from this sequence that could be expressed in a pro- or eukaryotic expression system.

10 DNA sequences of the present invention can be constructed starting from genomic or cDNA sequences coding for proteins, e.g. phytases known in the art [for sequence information see references mentioned above, e.g. EP 684 313 or sequence data bases, for example like Genbank (Intelligenetics, California, USA), European Bioinformatics Institute (Hinstone Hall, Cambridge, GB), NBRF 15 (Georgetown University, Medical Centre, Washington DC, USA) and Vecbase (University of Wisconsin, Biotechnology Centre, Madison, Wisconsin, USA) or disclosed in the figures by methods of in vitro mutagenesis [see e.g. Sambrook et al., Molecular Cloning, Cold Spring Harbor Laboratory Press, New York]. A widely used strategy for "site directed mutagenesis", as originally outlined by Hurchinson and Edgell [J. Virol. 8, 181 (1971)], 20 involves the annealing of a synthetic oligonucleotide carrying the desired nucleotide substitution to a target region of a single-stranded DNA sequence wherein the mutation should be introduced [for review see Smith, Annu. Rev. Genet. 19, 423 (1985) and for improved methods see references 2-6 in Stanssen et al., Nucl. Acid Res., 17, 4441-4454 (1989)]. Another possibility of mutating a given DNA sequence which is also preferred for 25 the practice of the present invention is the mutagenesis by using the polymerase chain reaction (PCR). DNA as starting material can be isolated by methods known in the art and described e.g. in Sambrook et al. (Molecular Cloning) from the respective strains. For strain information see, e.g. EP 684 313 or any depositary authority indicated below. Aspergillus niger [ATCC 9142], Myceliophthora thermophila [ATCC 48102], 30 Talaromyces thermophilus [ATCC 20186] and Aspergillus fumigatus [ATCC 34625] have been redeposited according to the conditions of the Budapest Treaty at the American Type Culture Cell Collection under the following accession numbers: ATCC 74337, ATCC 74340, ATCC 74338 and ATCC 74339, respectively. It is however, understood that DNA encoding a consensus protein in accordance with the present invention can also be 35 prepared in a synthetic manner as described, e.g. in EP 747 483 or the examples by methods known in the art.

The process of the present invention can preferably be used in order to improve the thermostability of the enzyme phytase. After having constructed different consensus phytase sequences it was possible to decide whether single amino acid replacements had a positive or a negative effect on the protein stability. It is therefore another subject of the 5 present invention to improve the thermostability of a phytase.

In this embodiment single amino acids are changed in the sequence of the phytase by the introduction of at least one mutation selected from the group consisting of

E58A	F54Y
D69K	I73V
D197N	K94A
T214L	R101A
E222T	N153K
E267D	V158I
R291I	A203G
R329H	S205G
S364T	V217A
A379K	A227V
G404A	V234L
	P238A
	Q277E
	A287H
	A292Q
	V366I
	A396S
	E415Q
	G437A
	E451R

In the above-given mutations the number represents the position in the consensus phytase-1-sequence as given in Figure 2 and the letter before the number represents the amino acid in the phytase which is replaced by the respective amino acid behind the number. The numbers given correspond to the consensus phytase sequence or relate to a corresponding residue calculated by an alignment as shown in Figure 1 when 26 amino acids (signal sequence) are added to the sequences shown in Fig. 1. Those mutations can be introduced into consensus sequences or into sequences of specific enzymes which have been improved by a process of the present invention. The above-mentioned amino acid replacements have a positive effect on the protein stability.

Once complete DNA sequences of the present invention have been obtained they can be integrated into vectors by methods known in the art and described e.g. in Sambrook et al. (s.a.) to overexpress the encoded polypeptide in appropriate host systems. However, a man skilled in the art knows that also the DNA sequences themselves can be used to transform the suitable host systems of the invention to get overexpression of the encoded polypeptide. Appropriate host systems are for example fungi, like Aspergilli, e.g. Aspergillus niger [ATCC 9142] or Aspergillus ficuum [NRRL 3135] or like Trichoderma, e.g. Trichoderma reesei or yeasts, like Saccharomyces, e.g. Saccharomyces cerevisiae or Pichia, like Pichia pastoris, or Hansenula polymorpha, e.g. H. polymorpha (DSM5215) or plants, as described, e.g. by Pen et al., Bio/Technology 11, 811-814 (1994). A man skilled in the art knows that such microorganisms are available from depository authorities, e.g. the American Type Culture Collection (ATCC), the Centraalbureau voor Schimmelcultures (CBS) or the Deutsche Sammlung für Mikroorganismen und Zellkulturen GmbH (DSM) or any other depository authority as listed in the Journal "Industrial Property" [(1991) 1, pages 29-40]. Bacteria which can be used are e.g. E. coli, Bacilli as, e.g. Bacillus subtilis or Streptomyces, e.g. Streptomyces lividans (see e.g. Anné and Mallaert in FEMS Microbiol. Letters 114, 121 (1993). E. coli, which could be used are E. coli K12 strains e.g. M15 [described as DZ 291 by Villarejo et al. in J. Bacteriol. 120, 466-474 (1974)], HB 101 [ATCC No. 33694] or E. coli SG13009 [Gottesman et al., J. Bacteriol. 148, 265-273 (1981)].

Vectors which can be used for expression in fungi are known in the art and described e.g. in EP 420 358, or by Cullen et al. [Bio/Technology 5, 369-376 (1987)] or Ward in Molecular Industrial Mycology, Systems and Applications for Filamentous Fungi, Marcel Dekker, New York (1991), Upshall et al. [Bio/Technology 5, 1301-1304 (1987)] Gwynne et al. [Bio/Technology 5, 71-79 (1987)], Punt et al. [J. Biotechnol. 17, 19-34 (1991)] and for yeast by Sreekrishna et al. [J. Basic Microbiol. 28, 265-278 (1988), Biochemistry 28,

4117-4125 (1989)], Hitzemann et al. [Nature 293, 717-722 (1981)] or in EP 183 070, EP 183 071, EP 248 227, EP 263 311. Suitable vectors which can be used for expression in *E. coli* are mentioned, e.g. by Sambrook et al. [s.a.] or by Fiers et al. in Procd. 8th Int. Biotechnology Symposium" [Soc. Franc. de Microbiol., Paris (Durand et al., 5 eds.), pp. 680-697 (1988)] or by Bujard et al. in Methods in Enzymology, eds. Wu and Grossmann, Academic Press, Inc. Vol. 155, 416-433 (1987) and Stüber et al. in Immunological Methods, eds. Lefkovits and Pernis, Academic Press, Inc., Vol. IV, 121-152 (1990). Vectors which could be used for expression in *Bacilli* are known in the art and described, e.g. in EP 405 370, Procd. Natl. Acad. Sci. USA 81, 439 (1984) by Yansura and 10 Henner, Meth. Enzymol. 185, 199-228 (1990) or EP 207 459. Vectors which can be used for the expression in *H. Polymorpha* are known in the art and described, e.g. in Gellissen et al., Biotechnology 9, 291-295 (1991).

Either such vectors already carry regulatory elements, e.g. promotors, or the DNA sequences of the present invention can be engineered to contain such elements. Suitable 15 promotors which can be used are known in the art and are, e.g. for *Trichoderma reesei* the *cbh1*- [Haarki et al., Biotechnology 7, 596-600 (1989)] or the *pki1*-promotor [Schindler et al., Gene 130, 271-275 (1993)], for *Aspergillus oryzae* the *amy*-promotor [Christensen et al., Abstr. 19th Lunteren Lectures on Molecular Genetics F23 (1987), Christensen et al., Biotechnology 6, 1419-1422 (1988), Tada et al., Mol. Gen. Genet. 229, 20 301 (1991)], for *Aspergillus niger* the *glaA*- [Cullen et al., Bio/Technology 5, 369-376 (1987), Gwynne et al., Bio/Technology 5, 713-719 (1987), Ward in Molecular Industrial Mycology, Systems and Applications for Filamentous Fungi, Marcel Dekker, New York, 83-106 (1991)], *alcA*- [Gwynne et al., Bio/Technology 5, 718-719 (1987)], *suc1*- [Boddy et 25 al., Curr. Genet. 24, 60-66 (1993)], *aphA*- [MacRae et al., Gene 71, 339-348 (1988), MacRae et al., Gene 132, 193-198 (1993)], *tpiA*- [McKnight et al., Cell 46, 143-147 (1986), Upshall et al., Bio/Technology 5, 1301-1304 (1987)], *gpdA*- [Punt et al., Gene 69, 49-57 (1988), Punt et al., J. Biotechnol. 17, 19-37 (1991)] and the *pkiA*-promotor [de Graaff et al., Curr. Genet. 22, 21-27 (1992)]. Suitable promotors which could be 30 used for expression in yeast are known in the art and are, e.g. the *pho5*-promotor [Vogel et al., Mol. Cell. Biol., 2050-2057 (1989); Rudolf and Hinnen, Proc. Natl. Acad. Sci. 84, 1340-1344 (1987)] or the *gap*-promotor for expression in *Saccharomyces cerevisiae* and for 35 *Pichia pastoris*, e.g. the *aox1*-promotor [Koutz et al., Yeast 5, 167-177 (1989); Sreekrishna et al., J. Basic Microbiol. 28, 265-278 (1988)], or the FMD promoter [Hollenberg et al., EPA No. 0299108] or MOX-promotor [Ledeboer et al., Nucleic Acids Res. 13, 3063-3082 (1985)] for *H. polymorpha*.

Accordingly vectors comprising DNA sequences of the present invention, preferably for the expression of said DNA sequences in bacteria or a fungal or a yeast host and such transformed bacteria or fungal or yeast hosts are also an object of the present invention.

It is also an object of the present invention to provide a system which allows for high expression of proteins, preferably phytases like the consensus phytase of the present invention in *Hansenula* characterized therein that the codons of the encoding DNA sequence of such a protein have been selected on the basis of a codon frequency table of the organism used for expression, e.g. yeast as in the present case (see e.g. in Example 3) and optionally the codons for the signal sequence have been selected in a manner as described for the specific case in Example 3. That means that a codon frequency table is prepared on the basis of the codons used in the DNA sequences which encode the amino acid sequences of the defined protein family. Then the codons for the design of the DNA sequence of the signal sequence are selected from a codon frequency table of the host cell used for expression whereby always codons of comparable frequency in both tables are used.

Once such DNA sequences have been expressed in an appropriate host cell in a suitable medium the encoded protein can be isolated either from the medium in the case the protein is secreted into the medium or from the host organism in case such protein is present intracellularly by methods known in the art of protein purification or described in case of a phytase, e.g. in EP 420 358. Accordingly a process for the preparation of a polypeptide of the present invention characterized in that transformed bacteria or a host cell as described above is cultured under suitable culture conditions and the polypeptide is recovered therefrom and a polypeptide when produced by such a process or a polypeptide encoded by a DNA sequence of the present invention are also an object of the present invention.

Once obtained the polypeptides of the present invention can be characterized regarding their properties which make them useful in agriculture any assay known in the art and described e.g. by Simons et al. [Br. J. Nutr. 64, 525-540 (1990)], Schöner et al. [J. Anim. Physiol. a. Anim. Nutr. 66, 248-255 (1991)], Vogt [Arch. Geflügelk. 56, 93-98 (1992)], Jongbloed et al. [J. Anim. Sci., 70, 1159-1168 (1992)], Perney et al. [Poultry Sci. 72, 2106-2114 (1993)], Farrell et al., [J. Anim. Physiol. a. Anim. Nutr. 62, 278-283 (1993)], Broz et al., [Br. Poultry Sci. 35, 273-280 (1994)] and Düngelhoef et al. [Animal Feed Sci. Technol. 49, 1-10 (1994)] can be used.

In general the polypeptides of the present invention can be used without being limited to a specific field of application, e.g. in case of phytases for the conversion of inositol polyphosphates, like phytate to inositol and inorganic phosphate.

Furthermore the polypeptides of the present invention can be used in a process for the preparation of a pharmaceutical composition or compound food or feeds wherein the components of such a composition are mixed with one or more polypeptides of the present invention. Accordingly compound food or feeds or pharmaceutical compositions comprising one or more polypeptides of the present invention are also an object of the present invention. A man skilled in the art is familiar with their process of preparation. Such pharmaceutical compositions or compound foods or feeds can further comprise additives or components generally used for such purpose and known in the state of the art.

It is furthermore an object of the present invention to provide a process for the reduction of levels of phytate in animal manure characterized in that an animal is fed such a feed composition in an amount effective in converting phytate contained in the feedstuff to inositol and inorganic phosphate.

Before describing the present invention in more detail a short explanation of the Figures enclosed is given below.

15 Figure 1: Design of the consensus phytase sequence. The letters represent the amino acid residues in the one-letter code. The following sequences were used for the alignment: *phyA* from *Aspergillus terreus* 9A-1 (Mitchell et al, 1997; from amino acid (aa) 27), *phyA* from *A. terreus* cbs116.46; (van Loon et al., 1998; from aa 27), *phyA* from *Aspergillus niger* var. *awamori* (Piddington et al, 1993; from aa 27), *phyA* from *A. niger* T213; from aa 27), *phyA* from *A. niger* strain NRRL3135 (van Hartingsveldt et al, 1993; from aa 27), *phyA* from *Aspergillus fumigatus* ATCC 13073 (Pasamontes et al, 1993; from aa 25), *phyA* from *A. fumigatus* ATCC 32722 (van Loon et al, 1998; from aa 27), *phyA* from *A. fumigatus* ATCC 58128 (van Loon et al, 1998; from aa 27), *phyA* from *A. fumigatus* ATCC 26906 (van Loon et al, 1998; from aa 27), *phyA* from *A. fumigatus* ATCC 32239 (van Loon et al, 1998; from aa 30), *phyA* from *Emericella nidulans* (Pasamontes et al, 1997a; from aa 25), *phyA* from *Talaromyces thermophilus* (Pasamontes et al, 1997a; from aa 24), and *phyA* from *Myceliophthora thermophila* (Mitchell et al, 1997; from aa 19). The alignment was calculated using the program PILEUP. The location of the gaps was refined by hand. Capitalized amino acid residues in the alignment at a given position belong to the amino acid coalition that establish the consensus residue. In bold, beneath the calculated consensus sequence, the amino acid sequence of the finally constructed consensus phytase (Fcp) is shown. The gaps in the calculated consensus sequence were filled by hand according to principals stated in Example 1.

5 **Figure 2:** DNA sequence of the consensus phytase-1 gene (*scp*) and of the primers used for the gene construction. The calculated amino acid sequence (Figure 1) was converted into a DNA sequence using the program BACKTRANSLATE (Devereux *et al.*, 1984) and the codon frequency table of highly expressed yeast genes (GCG program package, 9.0). The signal peptide of the phytase from *A. terreus* cbs.116.46 was fused to the N-terminus. The bold bases represent the sequences of the oligonucleotides used to generate the gene. The names of the respective oligonucleotides are alternately noted above or below the sequence. The underlined bases represent the start and stop codon of the gene. The bases written in italics show the two introduced *Eco* RI sites.

10 10 **Figure 3:** Alignment and consensus sequence of five *Basidiomycetes* phytases. The letters represent the amino acid residues in the one-letter code. The amino acid sequences of the phytases from *Paxillus involutus*, phyA1 (aa 21) and phyA2 (aa 21, WO 98/28409), *Trametes pubescens* (aa 24, WO 98/28409), *Agrocybe pediades* (aa 19, WO 98/28409), and *Peniophora lycii* (aa 21, WO 98/28409) starting with the amino acid residues 15 mentioned in parentheses, were used for the alignment and the calculation of the corresponding consensus sequence called "Basidio" (Example 2). The alignment was performed by the program PILEPUP. The location of the gaps was refined by hand. The consensus sequence was calculated by the program PRETTY. While a vote weight of 0.5 was assigned to the two *P. involutus* phytases, all other genes were used with a vote weight 20 of 1.0 for the consensus sequence calculation. At positions, where the program was not able to determine a consensus residues, the Basidio sequence contains a dash. Capitalized amino acid residues in the alignment at a given position belong to the amino acid coalition that establish the consensus residue.

25 **Figure 4:** Design of consensus phytase-10 amino acid sequence. Adding the phytase sequence of *Thermomyces lanuginosa* (Berka *et al.*, 1998) and the consensus sequence of the phytases from five *Basidiomycetes* to the alignment of Figure 1, an improved consensus sequence was calculated by the program PRETTY. Additionally, the amino acid sequence of *A. niger* T213 was omitted, therefore, using a vote weight of 0.5 for the remaining *A. niger* phytase sequences. For further information see Example 2.

30 **Figure 5:** DNA and amino acid sequence of consensus phytase-10. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The sequence of the oligonucleotides which were used to assemble the gene are in bold letters. The label of oligonucleotides and the amino acids, which were changed compared to those for consensus phytase -1, are underlined and their corresponding triplets are highlighted in

small cases. The *fcp10* gene was assembled from the following oligonucleotides: CP-1, CP-2, CP-3.10, CP-4.10, CP-5.10, CP-6, CP-7.10, CP-8.10, CP-9.10, CP-10.10, CP-11.10, CP-12.10, CP-13.10, CP-14.10, CP-15.10, CP-16.10, CP-17.10, CP18.10, CP-19.10, CP-20.10, CP-21.10, CP-22.10. The newly synthesized oligonucleotides are additionally 5 marked by number 10. The phytase contains the following 32 exchanges: Y54F, **E58A**, D69K, D70G, A94K, N134Q, I158V, S187A, Q188N, **D197N**, S204A, T214L, D220E, L234V, A238P, D246H, T251N, Y259N, **E267D**, E277Q, A283D, **R291I**, A320V, **R329H**, **S364T**, I366V, **A379K**, S396A, **G404A**, Q415E, A437G, A463E. The mutations 10 accentuated in bold letters revealed a stabilizing effect on consensus phytase-1 as tested as single mutation in consensus phytase-1.

Figure 6: Alignment for the design of consensus phytase-11. In contrast to the design of consensus phytase-10, for the design of the amino acid sequence of consensus phytase-11, all *Basidiomycetes* phytases were used as independent sequences using an assigned vote weight of 0.2 for each *Basidiomycetes* sequence. Additionally, the amino acid sequence of 15 *A. niger* T213 was used in that alignment, again.

Figure 7: DNA and amino acid sequence of consensus phytase-1-thermo[8]-Q50T-K91A. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (*). 20 Figure 8: DNA and amino acid sequence of consensus phytase-10-thermo[3]-Q50T-K91A. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (*).

Figure 9: DNA and amino acid sequence of *A. fumigatus* ATCC 13073 phytase a-mutant. 25 The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (*).

Figure 10: DNA and amino acid sequence of consensus phytase-7. The amino acids are written above the corresponding DNA sequence using the one-letter code. The sequence of 30 the oligonucleotides used to assemble the gene are in bold letters. Oligonucleotides and amino acids that were exchanged are underlined and their corresponding triplets are highlighted in small cases. The *fcp7* gene was assembled from the following

oligonucleotides: CP-1, CP-2, CP-3, CP-4.7, CP-5.7, CP-6, CP-7, CP-8.7, CP-9, CP-10.7, CP-11.7, CP-12.7, CP-13.7, CP-14.7, CP-15.7, CP-16, CP-17.7, CP-18.7, CP-19.7, CP-20, CP-21, CP-22. The newly synthesized oligonucleotides are additionally marked by number 7. The phytase contains the following 24 exchanges in comparison to the original 5 consensus phytase: S89D, S92G, A94K, D164S, P201S, G203A, G205S, H212P, G224A, D226T, E255T, D256E, V258T, P265S, Q292H, G300K, Y305H, A314T, S364G, M365I, A397S, S398A, G404A, and A405S.

10 **Figure 11:** Differential scanning calorimetry (DSC) of consensus phytase-1 and consensus phytase-10. The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-10 (upper graph) yielded a melting temperature of 85.4 °C, which is 7.3 °C higher than the melting point of consensus phytase-1 (78.1 °C, lower graph).

15 **Figure 12:** Differential scanning calorimetry (DSC) of consensus phytase-10-thermo-Q50T and consensus phytase-10-thermo-Q50T-K91A. The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-10-thermo-Q50T (upper graph) yielded a melting temperature of 88.6 °C, while the melting point of consensus phytase-10-thermo-Q50T-K91A was found at 89.3 °C.

20 **Figure 13:** Comparison of the temperature optimum between consensus phytase-1, consensus phytase-10 and consensus phytase-10-thermo-Q50T. For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 86 °C. The diluted supernatant of transformed *S. cerevisiae* strains was used for the determination. The other components of the supernatant showed 25 no influence on the determination of the temperature optimum: ^, consensus phytase-1; ◇, consensus phytase-10; ■, consensus phytase 10-thermo-Q50T.

30 **Figure 14:** pH-dependent activity profile and substrate specificity of consensus phytase-10 and its variants thermo-Q50T and thermo-Q50T-K91A. The phytase activity was determined using the standard assay in appropriate buffers (see Example 9) at different pH-values. Graph a) shows the pH-dependent activity profile of consensus phytase-10 (□), consensus phytase-10-thermo-Q50T (●), and consensus phytase-10-thermo-Q50T-K91A (^). Graph b) shows the corresponding substrate specificity tested by replacement of phytate by the indicated compounds in the standard assay; open bars, consensus phytase-10

(grey bars, consensus phytase-10-thermo-Q50T; dark bars, consensus phytase-10-thermo-Q50T-K91A). The numbers correspond to the following compounds: 1, phytate; 2, *p*-nitrophenyl phosphate; 3, phenyl phosphate; 4, fructose-1,6-bisphosphate; 5, fructose-6-phosphate; 6, glucose-6-phosphate; 7, ribose-5-phosphate; 8, DL-glycerol-3-phosphate; 9, glycerol-2-phosphate; 10, 3-phosphoglycerate; 11, phosphoenolpyruvate; 12, AMP; 13, ADP; 14, ATP.

5

10 Figure 15: pH-dependent activity profile and substrate specificity of consensus phytase-1-thermo[8]-Q50T and of consensus phytase-1-thermo[8]-Q50T-K91A. The phytase activity was determined using the standard assay in appropriate buffers (see Example 9) at different pH-values. Graph a) shows the pH-dependent activity profile of the Q50T- (■) and the Q50T-K91A-variant (○). Graph b) shows the corresponding substrate specificities tested by replacement of phytate by the indicated compounds in the standard assay (open bars, consensus phytase-1-thermo[8]-Q50T; filled bars, consensus phytase-1-thermo[8]-Q50T-K91A.). The substrates are listed in the legend of Figure 14.

15 Figure 16: Differential scanning calorimetry (DSC) of consensus phytase-1-thermo[8]-Q50T and consensus phytase-1-thermo[8]-Q50T-K91A. The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-1-thermo[8]-Q50T (upper graph) showed a melting temperature of 84.7 °C, while 20 the melting point of consensus phytase-1-thermo[8]-Q50T-K91A was found at 85.7 °C.

15 Figure 17: Comparison of the temperature optimum between consensus phytase-1, consensus phytase-1-thermo[3] and consensus phytase-1-thermo[8]. For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 86 °C. Purified protein from the supernatant of transformed 25 *S. cerevisiae* strains was used for the determination. O, consensus phytase-1; □, consensus phytase-1-thermo[3]; ▲, consensus phytase 1-thermo[8].

20 Figure 18: Comparison of the pH-dependent activity profile and substrate specificity of consensus phytase-1, consensus phytase-7, and of the phytase from *A. niger* NRRL 3135. The phytase activity was determined using the standard assay in appropriate buffers (see Example 9) at different pH-values. Graph a) shows the pH-dependent activity profile of consensus phytase-1 (■), the phytase from *A. niger* NRRL 3135 (○), and of consensus 30 phytase-7 (▲). Graph b) shows the corresponding substrate specificity tested by replacement of phytate by the indicated compounds in the standard assay (black bars, *A.*

niger NRRL 3135 phytase; grey bars, consensus phytase-1, dashed bars, consensus phytase-7). The substrates are listed in the legend of Figure 14.

Figure 19: Differential scanning calorimetry (DSC) of the phytase from *A. fumigatus* ATCC 13073 and of its stabilized α -mutant, which contains the following amino acid exchanges F55Y, V100I, F114Y, A243L, S265P, N294D.

The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus *A. fumigatus* 13073 phytase (upper graph) revealed a melting temperature of 62.5 °C, while the melting point of the α -mutant was found at 67.0

10 °C

Figure 20: Comparison of the temperature optimum of *A. fumigatus* 13073 wild-type, its *A. fumigatus* α -mutant, and a further stabilized α -mutant (E59A-S126N-R329H-S364T-G404A). For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 75 °C. The diluted supernatant

15 of transformed *S. cerevisiae* strains was used for the determination. The other components of the supernatant showed no influence on the determination of the temperature optimum.

○, *A. fumigatus* ATCC 13073 phytase; ▲, *A. fumigatus* ATCC 13073 α -mutant; □, *A. fumigatus* ATCC 13073 alpha-mutant-(E59A-S126N-R329H-S364T-G404A)-Q27T; ■, *A. fumigatus* ATCC 13073 α -mutant-(E59A-S126N-R329H-S364T-G404A)-Q27T-K68A.

20 Q27T and K68A corresponds to consensus phytase-1 Q50T and K91A, respectively.

Figure 21: Amino acid sequence of consensus phytase 12 (consphy12) which contains a number of active site residues transferred from the "basidio" consensus sequence to consensus phytase-10-thermo-Q50T-K91A.

Example 1:

Design of the amino acid sequence of consensus phytase-1

Alignment of the amino acid sequences

The alignment was calculated using the program PILEUP from the Sequence Analysis Package Release 9.0 (Devereux *et al.*, 1984) with the standard parameter (gap creation penalty 12, gap extension penalty 4). The location of the gaps was refined using a text editor. Table 1 shows the sequences (see Figure 1) without the signal sequence that were used for the performance of the alignment starting with the amino acid (aa) as mentioned in Table 1.

10 Table 1: Origin and vote weight of the phytase amino acid sequences used for the design of consensus phytase-1

- *phyA* from *Aspergillus terreus* 9A-1, aa 27, vote weight 0.5 (Mitchell *et al.*, 1997)

- *phyA* from *Aspergillus terreus* cbs116.46, aa 27, vote weight 0.5 (van Loon *et al.*, 1998)

- *phyA* from *Aspergillus niger* var. *awamori*, aa 27, vote weight 0.33 (Piddington *et al.*, 15 1993)

- *phyA* from *Aspergillus niger* T213, aa 27, vote weight 0.33

- *phyA* from *Aspergillus niger* strain NRRL3135, aa 27, vote weight 0.33 (van Hartingsveldt *et al.*, 1993)

- *phyA* from *Aspergillus fumigatus* ATCC 13073, aa 26, vote weight 0.2 (Pasamontes *et al.*, 20 1997)

- *phyA* from *Aspergillus fumigatus* ATCC 32722, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)

- *phyA* from *Aspergillus fumigatus* ATCC 58128, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)

25 - *phyA* from *Aspergillus fumigatus* ATCC 26906, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)

- *phyA* from *Aspergillus fumigatus* ATCC 32239, aa 30, vote weight 0.2 (van Loon *et al.*, 1998)

- *phyA* from *Emericella nidulans*, aa 25, vote weight 1.0 (Roche Nr. R1288, Pasamontes *et al.*, 30 1997a)

- *phyA* from *Talaromyces thermophilus* ATCC 20186, aa 24, vote weight 1.0 (Pasamontes *et al.*, 1997a)

- *phyA* from *Myceliophthora thermophila*, aa 19, vote weight 1.0 (Mitchell *et al.*, 1997)

Calculation of the amino acid sequence of consensus phytase-1

Using the refined alignment as input, the consensus sequence was calculated by the program PRETTY from the Sequence Analysis Package Release 9.0 (Devereux *et al.*, 1984). PRETTY prints sequences with their columns aligned and can display a consensus sequence for an alignment. A vote weight that pays regard to the similarity between the amino acid sequences of the phytases aligned was assigned to all sequences. The vote weight was set such as the combined impact of all phytases from one sequence subgroup (same species, but from different strains), e. g. the amino acid sequences of all phytases from *A. fumigatus*, on the election was set one, that means that each sequence contributes with a value of 1 divided by the number of strain sequences (see Table 1). By this means, it was possible to prevent that very similar amino acid sequences, e. g. of the phytases from different *A. fumigatus* strains, dominate the calculated consensus sequence.

The program PRETTY was started with the following parameters: The plurality defining the number of votes below which there is no consensus was set on 2.0. The threshold, which determines the scoring matrix value below which an amino acid residue may not vote for a coalition of residues, was set on 2. PRETTY used the PrettyPep.Cmp consensus scoring matrix for peptides.

Ten positions of the alignment (position 46, 66, 82, 138, 162, 236, 276, 279, 280, 308; Figure 1), for which the program was not able to determine a consensus residue, were filled by hand according to the following rules: if a most frequent residue existed, this residue was chosen (138, 236, 280); if a prevalent group of similar or phylogenetically equivalent residues occurred, the most frequent or, if not available, one residues of this group was selected (46, 66, 82, 162, 276, 308). If there was either a prevalent residue nor a prevalent group, one of the occurring residues was chosen according to common assumption on their influence on the protein stability (279). Eight other positions (132, 170, 204, 211, 275, 317, 384, 447; Figure 1) were not filled with the amino acid residue selected by the program but normally with amino acids that occur with the same frequency as the residues that were chosen by the program. In most cases, the slight underrating of the three *A. niger* sequences (sum of the vote weights: 0.99) was eliminated by this corrections.

Conversion of the consensus phytase-1 amino acid sequence to a DNA sequence

The first 26 amino acid residues of *A. terreus* cbs116.46 phytase were used as signal peptide and, therefore, fused to the N-terminus of all consensus phytases. For this stretch,

we used a special method to calculate the corresponding DNA sequence. Purvis et al (1987) proposed that the incorporation of rare codons in a gene has an influence on the folding efficiency of the protein. Therefore, at least the distribution of rare codons in the signal sequence of *A. terreus* CBS 116.46, which was used for the consensus phytase and 5 which is very important for secretion of the protein, but converted into the *S. cerevisiae* codon usage, was transferred into the new signal sequence generated for expression in *S. cerevisiae*. For the remaining parts of the protein, we used the codon frequency table of highly expressed *S. cerevisiae* genes, obtained from the GCG program package, to translate the calculated amino acid sequence into a DNA sequence.

10 The resulting sequence of the *fcp* gene is shown in Figure 2.

Construction and cloning of the consensus phytase-1 gene

The calculated DNA sequence of consensus phytase-1 (*fcp*) was divided into oligonucleotides of 85 bp, alternately using the sequence of the sense and the anti-sense strand. Every oligonucleotide overlaps 20 bp with its previous and its following 15 oligonucleotide of the opposite strand. The location of all primers, purchased by Microsynth, Balgach (Switzerland) and obtained in a PAGE-purified form, is indicated in Figure 2.

PCR-Reactions

In three PCR reactions, the synthesized oligonucleotides were composed to the entire 20 gene. For the PCR, the High Fidelity Kit from Boehringer Mannheim (Boehringer Mannheim, Mannheim, Germany) and the thermo cycler The ProtokolTM from AMS Biotechnology (Europe) Ltd. (Lugano, Switzerland) was used.

Oligonucleotide CP-1 to CP-10 (Mix 1, Figure 2) were mixed to a concentration of 0.2 pMol/μl of each oligonucleotide. A second oligonucleotide mixture (Mix 2) was 25 prepared with CP-9 to CP-22 (0.2 pMol/μl of each oligonucleotide). Additionally, four short primers were used in the PCR reactions:

CP-a: *Eco RI*

5'-TATATGAATTCATGGCGTGTTCGTC-3'

CP-b:

30 5'-TGAAAAGTTCATTGAAGGTTTC-3'

CP-c:

5'-TCTTCGAAAGCAGTACAAGTAC-3'

CP-e:

Eco RI

5'-TATATGAATTCTTAAGCGAAC-3'

PCR reaction *a*:

5 10 μ l Mix 1 (2.0 pmol of each oligonucleotide)
2 μ l nucleotides (10 mM each nucleotide)
2 μ l primer CP-a (10 pmol/ μ l)
2 μ l primer CP-c (10 pmol/ μ l)
10,0 μ l PCR buffer
0.75 μ l polymerase mixture
10 73.25 μ l H₂O

PCR reaction *b*:

15 10 μ l Mix 2 (2.0 pmol of each oligonucleotide)
2 μ l nucleotides (10 mM each nucleotide)
2 μ l primer CP-b (10 pmol/ μ l)
2 μ l primer CP-e (10 pmol/ μ l)
10,0 μ l PCR buffer
0.75 μ l polymerase mixture (2.6 U)
73.25 μ l H₂O

Reaction conditions for PCR reaction *a* and *b*:

20 step 1 2 min - 45°C
step 2 30 sec - 72°C
step 3 30 sec - 94°C
step 4 30 sec - 52°C
step 5 1 min - 72°C

Step 3 to 5 were repeated 40-times.

25 The PCR products (670 and 905 bp) were purified by an agarose gel electrophoresis (0.9% agarose) and a following gel extraction (QIAEX II Gel Extraction Kit, Qiagen, Hilden, Germany). The purified DNA fragments were used for the PCR reaction *c*.

PCR reaction *c*:

30 6 μ l PCR product of reaction *a* (\approx 50 ng)
6 μ l PCR product of reaction *b* (\approx 50 ng)
2 μ l primer CP-a (10 pmol/ μ l)
2 μ l primer CP-e (10 pmol/ μ l)
10,0 μ l PCR buffer
0.75 μ l polymerase mixture (2.6 U)
73.25 μ l H₂O

35 Reaction conditions for PCR reaction *c*:

step 1 2 min - 94°C
step 2 30 sec - 94°C
step 3 30 sec - 55°C
step 4 1 min - 72°C

5 Step 2 to 4 were repeated 31-times.

The resulting PCR product (1.4 kb) was purified as mentioned above, digested with *Eco* RI, and ligated in an *Eco* RI-digested and dephosphorylated pBsk(-)-vector (Stratagene, La Jolla, CA, USA). 1 µl of the ligation mixture was used to transform *E. coli* XL-1 competent cells (Stratagene, La Jolla, CA, USA). All standard procedures were 10 carried out as described by Sambrook *et al.* (1987). The DNA sequence of the constructed consensus phytase gene (*fcp*, Figure 2) was controlled by sequencing as known in the art.

Example 2

Design of an improved consensus phytase (consensus phytase-10) amino acid sequence

15 The alignments used for the design of consensus phytase-10 were calculated using the program PILEUP from the Sequence Analysis Package Release 9.0 (Devereux *et al.*, 1984) with the standard parameter (gap creation penalty 12, gap extension penalty 4). The location of the gaps was refined using a text editor.

20 The following sequences were used for the alignment of the *Basidiomycetes* phytases starting with the amino acid (aa) mentioned in Table 2:

Table 2: Origin and vote weight of five *Basidiomycetes* phytases used for the calculation of the corresponding amino acid consensus sequence (basidio)

- *phyA1* from *Paxillus involutus* NN005693, aa 21, vote weight 0.5 (WO 98/28409)
- *phyA2* from *Paxillus involutus* NN005693, aa 21, vote weight 0.5 (WO 98/28409)
- 25 - *phyA* from *Trametes pubescens* NN9343, aa 24, vote weight 1.0 (WO 98/28409)
- *phyA* from *Agrocybe pediades* NN009289, aa 19, vote weight 1.0 (WO 98/28409)
- *phyA* from *Peniophora lycii* NN006113, aa 21, vote weight 1.0 (WO 98/28409)

The alignment is shown in Figure 3.

30 In Table 3 the genes, which were used for the performance of the final alignment, are arranged. The first amino acid (aa) of the sequence which is used in the alignment is mentioned behind the organism designation.

Table 3: Origin and vote weight of the phytase sequences used for the design of consensus phytase 10

- *phyA* from *Aspergillus terreus* 9A-1, aa 27, vote weight 0.5 (Mitchell *et al.*, 1997)
- *phyA* from *Aspergillus terreus* cbs116.46, aa 27, vote weight 0.5 (van Loon *et al.*, 1998)
- 5 - *phyA* from *Aspergillus niger* var. *awamori*, aa 27, vote weight 0.5 (Piddington *et al.*, 1993)
- *phyA* from *Aspergillus niger* strain NRRL3135, aa 27, vote weight 0.5 (van Hartingsveldt *et al.*, 1993)
- *phyA* from *Aspergillus fumigatus* ATCC 13073, aa 26, vote weight 0.2 (Pasamontes *et al.*, 1997)
- 10 - *phyA* from *Aspergillus fumigatus* ATCC 32722, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)
- *phyA* from *Aspergillus fumigatus* ATCC 58128, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)
- 15 - *phyA* from *Aspergillus fumigatus* ATCC 26906, aa 26, vote weight 0.2 (van Loon *et al.*, 1998)
- *phyA* from *Aspergillus fumigatus* ATCC 32239, aa 30, vote weight 0.2 (van Loon *et al.*, 1998)
- *phyA* from *Emericella nidulans*, aa 25, vote weight 1.0 (Roche Nr. R1288, Pasamontes *et al.*, 1997a)
- 20 - *phyA* from *Talaromyces thermophilus* ATCC 20186, aa 24, vote weight 1.0 (Pasamontes *et al.*, 1997a)
- *phyA* from *Myceliophthora thermophila*, aa 19, vote weight 1.0 (Mitchell *et al.*, 1997)
- *phyA* from *Thermomyces lanuginosa*, aa 36, vote weight 1.0 (Berka *et al.*, 1998)
- 25 - Consensus sequence of five *Basidiomycetes* phytases, vote weight 1.0 (Basidio, Figure 3)

The corresponding alignment is shown in Figure 4.

Calculation of the amino acid sequence of consensus-10

To improve the alignment, we added the original consensus sequence of five phytases from four different *Basidiomycetes*, called Basidio, still containing the undefined sequence positions (see Figure 3), nearly all phytase sequences used for calculation of the original consensus phytase and one new phytase sequence from the *Ascomycete* *Thermomyces lanuginosa* to a larger alignment. Using the consensus sequence of the basidiomycetal phytase sequences, does not pay regard to the diversity among the five amino acid sequences, but pays regard to the common and different amino acid residues between the phytases from the *Ascomycetes* and the *Basidiomycetes*.

We set plurality on 2.0 and threshold on 3. The used vote weight are listed in Table 3. The alignment and the corresponding consensus sequence is presented in Figure 4. The new consensus phytase sequence has 32 different amino acids in comparison to the original consensus phytase. Positions for which the program PRETTY was not able to calculate a consensus amino acid residue were filled according to rules mentioned in Example 1. None of the residues suggested by the program was replaced.

Furthermore, we included all *Basidiomycetes* phytases as single amino acid sequences but assigning a vote weight of 0.2 in the alignment. The corresponding alignment is shown in Figure 6. The calculated consensus amino acid sequence (consensus phytase-11) has the following differences to the sequence of consensus phytase-10. Letter X means that the program was not able to calculate a consensus amino acid; the amino acid in parenthesis corresponds to the amino acid finally included into the consensus phytase-10.

D35X, X(K)69K, X(E)100E, A101R, Q134N, X(K)153N, X(H)190H, X(A)204S, X(E)220D, E222T, V227A, X(R)271R, H287A, X(D)288D, X(K)379K, X(I)389I, E390X, X(E)415E, X(A)416A, X(R)446L, E463A, whereas the numbering is as in Fig. 5.

We also checked single amino acid replacements suggested by the improved consensus sequences 10 and 11 on their influence on the stability of the original consensus phytase. The approach is described in example 3.

20 Conversion of consensus phytase-10 amino acid sequence to a DNA sequence

The first 26 amino acid residues of *A. terreus* cbs116.46 phytase were used as signal peptide and, therefore, fused to the *N*-terminus of consensus phytase-10. The used procedure is further described in Example 1.

25 The resulting sequence of the *fcp10* gene is shown in Figure 5.

25

Construction and cloning of the consensus phytase-10 gene (*fcp10*)

The calculated DNA sequence of *fcp10* was divided into oligonucleotides of 85 bp, alternately using the sequence of the sense and the anti-sense strand. Every oligonucleotide overlaps 20 bp with its previous and its following oligonucleotide of the opposite strand.

30 The location of all primers, purchased by Microsynth, Balgach (Switzerland) and obtained in a PAGE-purified form, is indicated in Figure 5.

PCR-Reactions

In three PCR reactions, the synthesized oligonucleotides were composed to the entire gene. For the PCR, the High Fidelity Kit from Boehringer Mannheim (Boehringer Mannheim, Mannheim, Germany) and the thermo cycler The ProtokolTM from AMS Biotechnology (Europe) Ltd. (Lugano, Switzerland) was used. The following oligonucleotides were used in a concentration of 0.2 pMol/ml.

5

Mix 1.10: CP-1, CP-2, CP-3.10, CP-4.10, CP-5.10, CP-6, CP-7.10, CP-8.10, CP-9.10, CP-10.10

10 Mix 2.10: CP-9.10, CP-11.10, CP-12.10, CP-13.10, CP-14.10, CP-15.10, CP-16.10, CP-17.10, CP-18.10, CP-19.10, CP-20.10, CP-21.10, CP-22.10

The newly synthesized oligonucleotides are marked by number 10. The phytase contains the following 32 exchanges, which are underlined in Figure 5, in comparison to the original consensus phytase: Y54F, E58A, D69K, D70G, A94K, N134Q, I158V, S187A, Q188N, D197N, S204A, T214L, D220E, L234V, A238P, D246H, T251N, Y259N, 15 E267D, E277Q, A283D, R291I, A320V, R329H, S364T, I366V, A379K, S396A, G404A, Q415E, A437G, A463E.

Four short PCR primer were used for the assembling of the oligonucleotides:

CP-a: *Eco RI*
5'-TATATGAATTCTATGGCGTGTTCGTC-3'

20 CP-b:
5'-TGAAAAGTTCATTAAGGTTTC-3'

CP-c.10:
5'-TCTTCGAAAGCAGTACACAAAC-3'

25 CP-e: *Eco RI*
5'-TATATGAATTCTTAAGCGAAAC-3'

PCR reaction a: 10 µl Mix 1.10 (2.0 pmol of each oligonucleotide)
2 µl nucleotides (10 mM each nucleotide)
2 µl primer CP-a (10 pmol/ml)
2 µl primer CP-c.10 (10 pmol/ml)
30 10,0 µl PCR buffer
0,75 µl polymerase mixture
73,25 µl H₂O

PCR reaction *b*: 10 µl Mix 2.10 (2.0 pmol of each oligonucleotide)
2 µl nucleotides (10 mM each nucleotide)
2 µl primer CP-b (10 pmol/ml)
2 µl primer CP-e (10 pmol/ml)
10,0 µl PCR buffer
0.75 µl polymerase mixture (2.6 U)
73.25 µl H₂O

5

Reaction conditions for PCR reaction *a* and *b*:

10 step 1 2 min - 45 °C
step 2 30 sec - 72 °C
step 3 30 sec - 94 °C
step 4 30 sec - 52 °C
step 5 1 min - 72 °C

Step 3 to 5 were repeated 40-times.

15 The PCR products (670 and 905 bp) were purified by an agarose gel electrophoresis (0.9% agarose) and a following gel extraction (QIAEX II Gel Extraction Kit, Qiagen, Hilden, Germany). The purified DNA fragments were used for the PCR reaction *c*.

PCR reaction *c*: 6 µl PCR product of reaction *a* ≈50 ng
6 µl PCR product of reaction *b* ≈50 ng
20 2 µl primer CP-a (10 pmol/ml)
2 µl primer CP-e (10 pmol/ml)
10,0 µl PCR buffer
0.75 µl polymerase mixture (2.6 U)
73.25 µl H₂O

25 Reaction conditions for PCR reaction *c*:

step 1 2 min - 94 °C
step 2 30 sec - 94 °C
step 3 30 sec - 55 °C
step 4 1 min - 72 °C

30 Step 2 to 4 were repeated 31-times.

The resulting PCR product (1.4 kb) was purified as mentioned above, digested with *Eco* RI, and ligated in an *Eco* RI-digested and dephosphorylated pBsk(-)-vector (Stratagene, La Jolla, CA, USA). 1 µl of the ligation mixture was used to transform *E. coli* XL-1 competent cells (Stratagene, La Jolla, CA, USA). All standard procedures were 35 carried out as described by Sambrook *et al.* (1987). The DNA sequence of the constructed gene (*fcp10*) was checked by sequencing as known in the art.

Example 3

Increasing the thermostability of consensus phytase-1 by introduction of single mutations suggested by the amino acid sequence of consensus phytase-10 and consensus phytase-11

5 In order to increase the thermostability of homologous genes, it is also possible to test the stability effect of each differing amino acid residue between the protein of interest and the calculated consensus sequence and to combine all stabilizing mutations into the protein of interest. We used the consensus phytase as protein of interest and tested the effect on the protein stability of 34 amino acid residues, differing to consensus phytase 10
10 and/or 11 as single mutations.

To construct muteins for expression in *A. niger*, *S. cerevisiae*, or *H. polymorpha*, the corresponding expression plasmid containing the consensus phytase gene was used as template for site-directed mutagenesis (see Example 6-8). Mutations were introduced using the "quick exchangeTM site-directed mutagenesis kit" from Stratagene (La Jolla, CA, 15 USA) following the manufacturer's protocol and using the corresponding primers. All mutations made and their corresponding primers are summarized in Table 4. Plasmids harboring the desired mutation were identified by DNA sequence analysis as known in the art.

Table 4: Primers used for site-directed mutagenesis of consensus phytase

20 (Exchanged bases are highlighted in bold. The introduction of a restriction site is marked above the sequence. When a restriction site is written in parenthesis, the mentioned site was destroyed by introduction of the mutation.)

mutation	Primer set
25 Q50T	<i>Kpn</i> I 5'-CACTTGTGGGT A CC T ACTCTCCAT A CTTCTC-3' 5'-GAGAAGTATGGAGAG T AG G TACCCACAAGTG-3'
30 Y54F	5'-GGTCAATA T CTCCATTCT T CT T GG A AG-3' 5'-CTTCCAAAGAGAAGAATGGAGAGTATTGACC-3'
E58A	5'-CATA T CT T GGCAGACGAAT T GC-3' 5'-GCAGATT CG T C CAAAGAGAAGTATG-3'

		<i>Aat</i> II
	D69K	5'-CTCCAGACGTCCCAAAGGACTGTAGAGTTAC-3' 5'-GTAACCTACAGTCCTTGGGACGTCTGGAG-3'
		<i>Aat</i> II
5	D70G	5'-CTCCAGACGTCCCAGACGGCTGTAGAGTTAC-3' 5'-GTAACCTACAGCCGTCTGGGACGTCTGGAG-3'
		<i>Sca</i> I
10	A94K	5'-CTTCTAAGTCTAAGAAGTACTCTGCTTG-3' 5'-CAAAGCAGAGTACTTCTTAGACTAGAACG-3'
		<i>Nru</i> I
15	A101R	5'-GCTTACTCTGCTTGATTGAACGGATTCAAAAGAACGCTAC-3' 5'-GTAGCGTTCTTGAAATCCGTTCAATCAAAGCAGAGTAAGC-3'
		<i>Bss</i> HI
	N134Q	5'-CCATTGGTGAACAGCAAATGGTTAACTC-3' 5'-GAGTTAACCATTTGCTGTTCACCGAATGG-3'
20	K153N	5'-GATACAAGGCTCTCGCGAGAACATTGTT-3' 5'-GGAACAAATGTTCTCGCGAGAGCCTGTATC-3'
		<i>Bcl</i> I
25	D197N	5'-CTCCAGTTATTAACGTGATCATTCCAGAACGG-3' 5'-CCTCTGGAAATGATCACGTTAAACTGGAG-3'
		<i>Apa</i> I
30	S187A	5'-GGCTGACCCAGGGGCCAACACACCAAGC-3' 5'-GCTTGGTGTGGTGGGCCCTGGTCAGCC-3'
		<i>Nco</i> I
	T214L	5'-CACTTGGAACCATGGTCTTGTACTGCTTCG-3' 5'-CGAAAGCAGTACAAAGACCATGGTCCAAAGTG-3'
		<i>Avr</i> II
35	E222T	5'-GCTTCGAAGACTCTACCCCTAGGTGACGACGTTG-3' 5'-CAACGTCGTACCTAGGGTAGAGTCTCGAAAGC-3'

	V227A	5'-GGTGACGACGCTGAAGCTAACCCAC-3' 5'-GTGAAGTTAGCTTCAGCGTCGTACCC-3'
5	L234V	<i>Sac</i> II 5'-CTAACTTCACCGCGGTGTTGCTCCAG-3' 5'-CTGGAGCGAACACCGCGGTGAAGTTAG-3'
	A238P	5'-GCTTTGTTGCTCCACCTATTAGAGCTAGATTGG-3' 5'-CCAATCTAGCTCTAATAGGTGGAGCGAACAAAGC-3'
10	T251N	<i>Hpa</i> I 5'-GCCAGGTGTTAACCTGACTGACGAAG-3' 5'-TTCGTCAGTCAGTTAACACACCTGGC-3'
	Y259N	<i>Aat</i> II 5'-GACGAAGACGTCGTTAACCTGATGGAC-3' 5'-GTCCATCAAGTTAACGACCGTCTTCGTC-3'
15	E267D	<i>Asp</i> I 5'-GTCCATTGACACTGTCGCTAGAACTT C-3' 5'-GAAGTTCTAGCGACAGTGTGCAATGGAC-3'
20	E277Q	5'-CTGACGCTACTCAGCTGTCTCCATTTC-3' 5'-GAATGGAGACAGCTGAGTAGCGTCAG-3'
	A283D	5'-GTCTCCATTCTGTGATTGTTCACTCAC-3' 5'-GTGAGTGAACAAATCACAGAATGGAGAC-3'
25	H287A	<i>Ksp</i> I 5'-GCTTTGTTCACCGCGGACGAATGGAG-3' 5'-CTCCATTGTCGCGGTGAACAAAGC-3'
	R291I	<i>Bam</i> HI 5'-CACGACGAATGGATCCAATACGACTAC-3' 5'-GTAGTCGTATTGGATCCATTGTCGTG-3'
30	Q292A	<i>Bsi</i> WI 5'-GACGAATGGAGAGCGTACGACTACTTG-3' 5'-CAAGTAGTCGTACGCTCTCCATTGTC-3'
35	A320V	<i>Hpa</i> I 5'-GGTGGTTGGTTCTGTTAACGAATTGATTGC-3' 5'-GCAATCAATTGTTAACGAAACCAACACC-3'
	R329H	(<i>Bgl</i> II) 5'-GCTAGATTGACTCACTCTCCAGTTCAAG-3' 5'-CTTGAACGGAGAGTGTGAGTCAATCTAGC-3'

		<i>Eco</i> RV
S364T	5'-CTCACGACAACACTATGATATCTATTTCTTC-3' 5'-GAAGAAAATAGATATCATAGTGTGTCGTGAG-3'	
5 I366V		<i>Nco</i> I
	5'-CGACAACTCCATGGTTCTATTTCTTCGC-3' 5'-GCGAAGAAAATAGAAACCATGGAGTTGTCG-3'	
		<i>Kpn</i> I
A379K	5'-GTACAACGGTACCAAGCCATTGTCTAC-3' 5'-GTAGACAATGGCTTGGTACCGTTGTAC-3'	
10 S396A	5'-CTGACGGTTACGCTGCTTCTGGAC-3' 5'-GTCCAAGAACGAGCGTAACCGTCAG-3'	
15 G404A	5'-CTGTTCCATTGCGCTGCTAGAGCTTAC-3' 5'-GTAAGCTCTAGCAGCGAATGGAACAG-3'	
Q415E	5'-GATGCAATGTGAAGCTGAAAAGGAACC-3' 5'-GGTCCTTTCAGCTCACATTGCATC-3'	
20 A437G	5'-CACGGTTGTGGTGTGACAAGTTGGG-3' 5'-CCCAACTTGTGACACCACAAACCGTG-3'	<i>Sal</i> I
		<i>Mun</i> I
A463E	5'-GATCTGGTGGCAATTGGGAGGAATGTTCG-3' 5'-CGAAACATTCCTCCCAATTGCCACCAGATC-3'	

25 and accordingly for other mutations.

The temperature optimum of the purified phytases, expressed in *Saccharomyces cerevisiae* (Example 7), was determined as outlined in Example 9. Table 5 shows the effect on the stability of consensus phytase for each mutation introduced.

30 Table 5: Stability effect of the individual amino acid replacements in consensus phytase-1
(+ or - means a positive, respectively, negative effect on the protein stability up to 1 °C, ++
and -- means a positive, respectively, negative effect on the protein stability between 1 and
3 °C; the number 10 or 11 corresponds to the consensus phytase sequence that suggests the
35 amino acid replacement.)

stabilizing		neutral		destabilizing	
mutation	effect	mutation	effect	mutation	effect
E58A (10)	+	D69A	±	Y54F (10)	-
D69K (11)	+	D70G (10)	±	V73I	-
D197N (10)	+	N134Q (10)	±	A94K (10)	-
T214L (10)	++	G186H	±	A101R (11)	-
E222T (11)	++	S187A (10)	±	K153N (11)	-
E267D (10)	+	T214V	±	I158V (10)	--
R291I*	+	T251N (10)	±	G203A	--
R329H (10)	+	Y259N (10)	±	G205S	-
S364T (10)	++	A283D (10)	±	A217V	-
A379K (11)	+	A320V (10)	±	V227A (11)	--
G404A (10)	++	K445T	±	L234V (10)	-
		A463E (10)	±	A238P (10)	--
				E277Q (10)	-
				H287A (11)	-
				Q292A (10)	-
				I366V (10)	-
				S396A (10)	--
				Q415E (11)	-
				A437G (10)	--
				E451R	--

*: This amino acid replacement was found in another round of mutations.

We combined eight positive mutations (E58A, D197N, E267D, R291I, R329H, S364T, A379K, G404A) in the consensus phytase using the primers and the technique mentioned above in this example. Furthermore, the mutations Q50T and K91A were introduced which mainly influence the catalytical characteristics of the phytase (see patent application EP 5 97810175.6 and EP 97112688 as well as Example 9). The DNA and amino acid sequence of the resulting phytase gene (consensus phytase-thermo[8]-Q50T-K91A) is shown in Figure 7. In this way, the temperature optimum and the melting point of the consensus phytase was increased by 7 °C (Figure 15, 16, 17).

Using the results of Table 5, we further improved the thermostability of consensus phytase 10 by the following back mutations K94A, V158I, and A396S that revealed a strong negative influence on the stability of consensus phytase. The resulting protein is phytase-10-thermo [3]. Furthermore, we introduced the mutations Q50T and K91A which mainly influence the catalytical characteristics of consensus phytase (see patent application EP 97810175.6 and EP 97112688 as well as Example 9 and Figure 14 and 15). The resulting 15 DNA and amino acid sequence is shown in Figure 8. The optimized phytase showed a 4 °C higher temperature optimum and melting point than consensus phytase 10 (Figure 12 and 13). Furthermore, the phytase has also a strongly increased specific activity with phytate as substrate of 250 U/mg at pH 5.5 (Figure 14).

Example 4

20 Stabilization of the phytase of *A. fumigatus* ATCC 13073 by replacement of amino acid residues with the corresponding consensus phytase-1 and consensus phytase-10 residues

At six typical positions where the *A. fumigatus* 13073 is the only or nearly the only phytase in the alignment of Figure 1 that does not contain the corresponding consensus 25 phytase amino acid residue, the non-consensus amino acid residue was replaced by the consensus one. In a first round, the following amino acids were substituted in *A. fumigatus* 13073 phytase, containing the Q27T substitution and the signal sequence of *A. terreus* cbs.116.46 phytase (see European Patent Application No. 97810175.6 and Figure 9):

F55(28)Y, V100(73)I, F114(87)Y, A243(220)L, S265(242)P, N294(282)D.

30 The numbers in parentheses confer to the numbering of Figure 1.

In a second round, four of the seven stabilizing amino acid exchanges (E59A, R329H, S364T, G404A) found in the consensus phytase-10 sequence and, tested as single mutation in consensus phytase-1 (Table 5), were additionally introduced into the *A. fumigatus* a-

mutant. Furthermore, the amino acid replacement S126N, shown to reduce the protease susceptibility of the phytase, was introduced.

The mutations were introduced as described in example 3 (see Table 6) and expressed as described in example 6 to 8. The resulting *A. fumigatus* 13073 phytase variants were called 5 α-mutant and β-mutant-E59A-S126N-R329H-S364T-G404A.

The temperature optimum (60 °C, Figure 20) and the melting point (67.0 °C, Figure 19) of the *A. fumigatus* 13073 phytase β-mutant was increased by 5 °C in comparison to the values of the wild-type (temperature optimum: 55 °C, T_m : 60 °C). The five additional amino acid replacements further increased the temperature optimum by 3 °C (Figure 20).

10 Table 6: Mutagenesis primers for stabilization of *A. fumigatus* phytase ATCC 13073

Mutation	Primer
F55Y	5'-CACGTACTGCCATA CTTTCGCTCGAG-3' 5'-CTCGAGCGAAAAGT ATGGCGAGTACGTG-3'
15 E58A	5'-CCATACTTTCGCTCGCGGACGAGCTGTCCGTG-3' 5'-CACGGACAGCTCGCCG AGCGAAAAGTAGG-3'
	<i>(Xba I)</i>
V100I	5'-GTATAAGAAGCTTATTACGGCGATCCAGGCC-3' 5'-GGCCTGGATCGCCGTAATAAG CTTCTTATAC-3'
20 F114Y	5'-CTTCAAGGGCAAGT ACGCC TTTG GAAGACG-3' 5'-CGTCTTCAAAAAGGCGTACTTG CCCT GAAG-3'
A243L	5'-CATCCGAGCTGCC TCGAGAAGCA TCTTC-3' 5'-GAAGATG CTTCTCGAGGCGAGCTCGGATG-3'
25 S265P	5'-CTAATGGA TGTGTCCGTTGATA ACGGTAG-3' 5'-CTACCGTATCAA ACGGACACATGTCCATTAG-3'

	N294D	5'-GTGGAAGAAGTACGACTACCTTCAGTC-3' 5'-GACTGAAGGTAGTCGTACTTCTTCCAC-3'
		<i>(Mlu I)</i>
5	R329H	5'-GCCCGGTTGACGCATTGCCAGTGCAGG-3' 5'-CCTGCACTGGCGAATGCGTCAACCGGGC-3'
		<i>Nco I</i>
	S364T	5'-CACACGACAACACC <u>ATGGTTCCATCTTC</u> -3' 5'-GAAGATGGAA <u>ACCATGGT</u> TTGTCGTGTG-3'
10		<i>(Bss HI)</i>
	G404A	5'-GTGGTGCCTTCGCGCGCGAGCCTACTTC-3' 5'-GAAGTAGGCTCGCGCGAAAGGCACCAC-3'

Example 5

Introduction of the active site amino acid residues of the *A. niger* NRRL 3135 phytase into the consensus phytase-1

We used the crystal structure of the *Aspergillus niger* NRRL 3135 phytase to define all active site amino acid residues (see Reference Example and EP 97810175.6). Using the alignment of Figure 1, we replaced the following active site residues and additionally the not identical adjacent ones of the consensus phytase by that of the *A. niger* phytase:

20 S89D, S92G, A94K, D164S, P201S, G203A, G205S, H212P, G224A, D226T, E255T, D256E, V258T, P265S, Q292H, G300K, Y305H, A314T, S364G, M365I, A397S, S398A, G404A, and A405S

25 The new protein sequence consensus phytase -7 was backtranslated into a DNA sequence (Figure 10) as described in Example 1. The corresponding gene (*fcp7*) was generated as described in Example 1 using the following oligonucleotide mixes:

Mix 1.7: CP-1, CP-2, CP-3, CP-4.7, CP-5.7, CP-6, CP-7, CP-8.7, CP-9, CP-10.7

Mix 2.7: CP-9, CP-10.7, CP-11.7, CP-12.7, CP-13.7, CP-14.7, CP-15.7, CP-16, CP-17.7, CP-18.7, CP-19.7, CP-20, CP-21, CP-22.

30 The DNA sequences of the oligonucleotides are indicated in Figure 3. The newly synthesized oligonucleotides are additionally marked by number 7. After assembling of the

oligonucleotides using the same PCR primers as mentioned in Example 1, the gene was cloned into an expression vector as described in Examples 6-8.

The pH-profile determined after expression in *H. polymorpha* and purification was shifted into the acidic range of the pH-spectrum showing an optimum at pH 4.5-5.0 (see Figure 5 18). The enzyme had a broad pH-optimum reaching at least 60% of its maximum activity from pH 2.5 to pH 6.0. Up to pH 5.0, the profile resembled the profile of the *A. niger* NRRL 3135 phytase. However, below pH 5.0 it lacked the typical low at pH 4.0 of the profile of *A. niger* phytase.

Example 6

10 Expression of the consensus phytase genes in *Hansenula polymorpha*

The phytase expression vectors, used to transform *H. polymorpha* RB11 (Gellissen *et al.*, 1994), was constructed by inserting the *Eco* RI fragment of pBsk-*fcp* or variants thereof into the multiple cloning site of the *H. polymorpha* expression vector pFPMT121, which is based on an *ura3* selection marker from *S. cerevisiae*, a formate dehydrogenase 15 (*FMD*) promoter element and a methanol oxidase (*MO*) terminator element from *H. polymorpha*. The 5' end of the *fcp* gene is fused to the *FMD* promoter, the 3' end to the *MOX* terminator (Gellissen *et al.*, 1996; EP 0299 108 B). The resulting expression vector are designated pFPMT-*fcp*, pFPMT-*fcp10*, pFPMT-*fcp7*.

The constructed plasmids were propagated in *E. coli*. Plasmid DNA was purified 20 using standard state of the art procedures. The expression plasmids were transformed into the *H. polymorpha* strain RP11 deficient in orotidine-5'-phosphate decarboxylase (*ura3*) using the procedure for preparation of competent cells and for transformation of yeast as described in Gellissen *et al.* (1996). Each transformation mixture was plated on YNB (0.14% w/v Difco YNB and 0.5% ammonium sulfate) containing 2% glucose and 1.8% 25 agar and incubated at 37 °C. After 4 to 5 days individual transformant colonies were picked and grown in the liquid medium described above for 2 days at 37 °C. Subsequently, an aliquot of this culture was used to inoculate fresh vials with YNB-medium containing 2% glucose. After seven further passages in selective medium, the expression vector integrates 30 into the yeast genome in multimeric form. Subsequently, mitotically stable transformants were obtained by two additional cultivation steps in 3 ml non-selective liquid medium (YPD, 2% glucose, 10 g yeast extract, and 20 g peptone). In order to obtain genetically 35 homogeneous recombinant strains an aliquot from the last stabilization culture was plated on a selective plate. Single colonies were isolated for analysis of phytase expression in YNB containing 2% glycerol instead of glucose to derepress the *fmd* promoter. Purification of the consensus phytases was done as described in Example 7.

Example 7

Expression of the consensus phytase genes in *Saccharomyces cerevisiae* and purification of the phytases from culture supernatant

The consensus phytase genes were isolated from the corresponding Bluescript-
5 plasmid (pBsk⁺scp, pBSK⁺scp10, pBsk⁺scp7) and ligated into the *Eco* RI sites of the expression cassette of the *Saccharomyces cerevisiae* expression vector pYES2 (Invitrogen, San Diego, CA, USA) or subcloned between the shortened GAPFL (glyceraldehyde-3-phosphate dehydrogenase) promoter and the *pho5* terminator as described by Janes *et al.* (1990). The correct orientation of the gene was checked by PCR. Transformation of *S.*
10 *cerevisiae* strains. e. g. INVSc1 (Invitrogen, San Diego, CA, USA) was done according to Hinnen *et al.* (1978). Single colonies harboring the phytase gene under the control of the GAPFL promoter were picked and cultivated in 5 ml selection medium (SD-uracil, Sherman *et al.*, 1986) at 30°C under vigorous shaking (250 rpm) for one day. The preculture was then added to 500 ml YPD medium (Sherman *et al.*, 1986) and grown
15 under the same conditions. Induction of the *gall* promoter was done according to manufacturer's instruction. After four days of incubation cell broth was centrifuged (7000 rpm, GS3 rotor, 15 min, 5°C) to remove the cells and the supernatant was concentrated by way of ultrafiltration in Amicon 8400 cells (PM30 membranes) and ultrafree-15 centrifugal filter devices (Biomax-30K, Millipore, Bedford, MA, USA). The concentrate
20 (10 ml) was desalting on a 40 ml Sephadex G25 Superfine column (Pharmacia Biotech, Freiburg, Germany), with 10 mM sodium acetate, pH 5.0, serving as elution buffer. The desalting sample was brought to 2 M (NH₄)₂SO₄ and directly loaded onto a 1 ml Butyl Sepharose 4 Fast Flow hydrophobic interaction chromatography column (Pharmacia Biotech, Feiburg, Germany) which was eluted with a linear gradient from 2 M to 0 M
25 (NH₄)₂SO₄ in 10 mM sodium acetate, pH 5.0. Phytase was eluted in the break-through, concentrated and loaded on a 120 ml Sephadryl S-300 gel permeation chromatography column (Pharmacia Biotech, Freiburg, Germany). Consensus phytase and consensus phytase -7 eluted as a homogeneous symmetrical peak and was shown by SDS-PAGE to be approx. 95% pure.

30 Example 8

Expression of the consensus phytase genes in *Aspergillus niger*

The Bluescript-plasmids pBsk⁺scp, pBSK⁺scp10, and pBsk⁺scp7 were used as template for the introduction of a *Bsp* HI-site upstream of the start codon of the genes and an *Eco* RV-site downstream of the stop codon. The ExpandTM High Fidelity PCR Kit (Boehringer
35 Mannheim, Mannheim, Germany) was used with the following primers:

Primer Asp-1:

Bsp HI

5'-TATATCATGAGCGTGGTCGTCGTGCTACTGTTC-3'

Primer Asp-2 used for cloning of *fcp* and *fcp7*:

5 *Eco* RV

3'-ACCCGACTTACAAAGCGAATTCTATAGATATAT-5'

Primer Asp-3 used for cloning of *fcp10*:

Eco RV

3'-ACCCTTCTTACAAAGCGAATTCTATAGATATAT-5'

10 The reaction was performed as described by the supplier. The PCR-amplified *fcp*-genes had a new *Bsp* HI site at the start codon, introduced by primer Asp-1, which resulted in a replacement of the second amino acid residue glycine by serine. Subsequently, the DNA-fragment was digested with *Bsp* HI and *Eco* RV and ligated into the *Nco* I site downstream of the glucoamylase promoter of *Aspergillus niger* (*glaA*) and the *Eco* RV site upstream of the *Aspergillus nidulans* tryptophan C terminator (*trpC*) (Mullaney *et al.*, 1985). After this cloning step, the genes were sequenced to detect possible failures introduced by PCR. The resulting expression plasmids which basically corresponds to the pGLAC vector as described in Example 9 of EP 684 313, contained the orotidine-5'-phosphate decarboxylase gene (*pyr4*) of *Neurospora crassa* as a selection marker.

20 Transformation of *Aspergillus niger* and expression of the consensus phytase genes was done as described in EP 684 313. The consensus phytases were purified as described in Example 7.

Example 9

Determination of phytase activity and of temperature optimum

25 Phytase activity was determined basically as described by Mitchell *et al* (1997). The activity was measured in an assay mixture containing 0.5% phytic acid (\approx 5 mM) in 200 mM sodium acetate, pH 5.0. After 15 min of incubation at 37 °C, the reaction was stopped by addition of an equal volume of 15% trichloroacetic acid. The liberated phosphate was quantified by mixing 100 μ l of the assay mixture with 900 μ l H₂O and 1 ml of 0.6 M H₂SO₄, 2% ascorbic acid and 0.5% ammonium molybdate. Standard solutions of potassium phosphate were used as reference. One unit of enzyme activity was defined as the amount of enzyme that releases 1 μ mol phosphate per minute at 37 °C. The protein

concentration was determined using the enzyme extinction coefficient at 280 nm calculated according to Pace et al (1995): consensus phytase, 1.101; consensus phytase 7, 1.068; consensus phytase 10, 1.039.

In case of pH-optimum curves, purified enzymes were diluted in 10 mM sodium acetate,
5 pH 5.0. Incubations were started by mixing aliquots of the diluted protein with an equal volume of 1% phytic acid (\approx 10 mM) in a series of different buffers: 0.4 M glycine/HCl, pH 2.5; 0.4 M acetate/NaOH, pH 3.0, 3.5, 4.0, 4.5, 5.0, 5.5; 0.4 M imidazole/HCl, pH 6.0, 6.5; 0.4 M Tris/HCl pH 7.0, 7.5, 8.0, 8.5, 9.0. Control experiments showed that pH was only slightly affected by the mixing step. Incubations were performed for 15 min at 37 °C as
10 described above.

For determinations of the substrate specificities of the phytases, phytic acid in the assay mixture was replaced by 5 mM concentrations of the respective phosphate compounds. The activity tests were performed as described above.

For determination of the temperature optimum, enzyme (100 μ l) and substrate
15 solution (100 μ l) were pre-incubated for 5 min at the given temperature. The reaction was started by addition of the substrate solution to the enzyme. After 15 min incubation, the reaction was stopped with trichloroacetic acid and the amount of phosphate released was determined.

The pH-optimum of the original consensus phytase was around pH 6.0-6.5 (70
20 U/mg). By introduction of the Q50T mutation, the pH-optimum shifted to pH 6.0 (130 U/mg). After introduction of K91A, the pH optimum shifted one pH-unit into the acidic pH-range showing a higher specific activity between pH 2.5 and pH 6.0. That was shown for the stabilized mutants and for consensus phytase-10, too (Figure 14 and 15).

Consensus phytase-7, which was constructed to transfer the catalytic characteristics of the
25 *A. niger* phytase NRRL 3135 into the consensus phytase, had a pH-profile which is shifted into the acidic range of the pH-spectrum showing an optimum between pH 4.5 and 5.0 (see Figure 19). The enzyme had a broad pH-optimum reaching at least 60% of its increased maximum activity from pH 2.5 to pH 6.0. The substrate spectrum, too, resemble more to that of the *A. niger* NRRL 3135 phytase than to the consensus phytase-1.

30 The temperature optimum of consensus phytase-1 (71 °C) was 16-26 °C higher than the temperature optimum of the wild-type phytases (45-55 °C, Table 7) which were used to calculate the consensus sequence. The improved consensus phytase-10 showed a further

increase of its temperature optimum to 80 °C (Figure 11). The temperature optimum of the consensus phytase-1-thermo[8] was found in the same range (78 °C) using the supernatant of an overproducing *S. cerevisiae* strain. The highest temperature optimum reached of 82 °C was determined for consensus phytase-10-thermo-Q50T-K91A.

5 **Table 7:** Temperature optimum and T_m -value of consensus phytase and of the phytases from *A. fumigatus*, *A. niger*, *E. nidulans*, and *M. thermophila*. The determination of the temperature optimum was performed as described in Example 9. The T_m -values were determined by differential scanning calorimetry as described in Example 10.

phytase	temperature optimum [°C]	T_m [°C]
Consensus phytase-10-thermo-Q50T-K91A	82	89.3
Consensus phytase-10-thermo-Q50T	82	88.6
Consensus phytase-10	80	85.4
Consensus phytase-1-thermo[8]-Q50T	78	84.7
Consensus phytase-1-thermo[8]-Q50T-K91A	78	85.7
Consensus phytase-1	71	78.1
<i>A. niger</i> NRRL3135	55	63.3
<i>A. fumigatus</i> 13073	55	62.5
<i>A. fumigatus</i> 13073 α -mutant	60	67.0
<i>A. fumigatus</i> 13073 α -mutant (optimized)	63	-
<i>A. terreus</i> 9A-1	49	57.5
<i>A. terreus</i> cbs.116.46	45	58.5
<i>E. nidulans</i>	45	55.7
<i>M. thermophila</i>	55	n. d.
<i>T. thermophilus</i>	45	n. d.

Example 10

Determination of the melting point by differential scanning calorimetry (DSC)

In order to determine the unfolding temperature of the phytases, differential scanning
5 calorimetry was applied as previously published by Brugger et al (1997). Solutions of 50-
60 mg/ml homogeneous phytase were used for the tests. A constant heating rate of 10 °
C/min was applied up to 90-95 °C.

The determined melting points reflect the results obtained for the temperature
optimums (Table 7). The most stable consensus phytase designed is consensus phytase-10-
10 thermo-Q50T-K91A showing a melting temperature under the chosen condition of 89.3 °
C. This is 26 to 33.6 °C higher than the melting point of the wild-type phytases used.

Example 11

Transfer of basidiomycete phytase active site into consensus phytase-10-thermo-Q50T-K91A

15 As described previously (Example 3), mutations derived from the basidiomycete
phytase active site were introduced into the consensus phytase 10. The following five
constructs a) to e) were prepared:

- a) This construct is called consensus phytase 12, and it comprises a selected number of
active site residues of the basidio consensus sequence, its amino acid sequence
20 (consphy12) is shown in Fig. 21 (the first 26 amino acids forms the signal peptide,
amended positions are underlined);
- b) a cluster of mutations (Cluster II) was transferred to the consensus 10 sequence, viz.:
S80Q, Y86F, S90G, K91A, S92A, K93T, A94R, Y95I;
- c) analogously, another cluster of mutations (Cluster III) was transferred, viz.: T129V,
25 E133A, Q143N, M136S, V137S, N138Q, S139A;
- d) analogously, a further cluster of mutations (Cluster IV) was transferred, viz.: A168D,
E171T, K172N, F173W;

e) and finally, a further cluster of mutations (Cluster V) was transferred, viz.: Q297G, S298D, G300D, Y305T.

These constructs were expressed as described in Examples 6 to 8.

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Claims

1. A process for the preparation of a consensus protein, whereby such process is characterized by the following steps:

- 5 a) at least three, preferably four amino acid sequences are aligned by any standard alignment program known in the art,
- 10 b) amino acids at the same position according to such alignment are compared regarding their evolutionary similarity by any standard program known in the art, whereas the degree of similarity provided by such a program which defines the least similarity of the amino acids that is used for the determination of an amino acid of corresponding positions is set to a less stringent number and the parameters are set in such a way that it is possible for the program to determine from only 2 identical amino acids at a corresponding position an amino acid for the consensus protein; however, if among the compared amino acid sequences are sequences that show a much higher degree of similarity to each other than to the residual sequences, these sequences are represented by their consensus sequence determined as defined in the same way as in the present process for the consensus sequence of the consensus protein or a vote weight of 1 divided by the number of such sequences is assigned to every of -those sequences,
- 15 c) in case no common amino acid at a defined position is identified by the program, any of the amino acids, preferably the most frequent amino acid of all such sequences is selected,
- 20 d) once the consensus sequence has been defined, such sequence is back-translated into a DNA sequence, preferably by using a codon frequency table of the organism in which expression should take place,
- 25 e) the DNA sequence is synthesized by methods known in the art and used either integrated into a suitable expression vector or by itself to transform an appropriate host cell,
- 30 f) the transformed host cell is grown under suitable culture conditions and the consensus protein is isolated from the host cell or its culture medium by methods known in the art.

2. A process as claimed in claim 1 wherein the program used for the comparison of amino acids at a defined position regarding their evolutionary similarity is the program "PRETTY".

3. A process as claimed in claims 1 or 2, wherein

in a first step a consensus sequence is determined from a number of highly homologous sequences according to steps a), b) and c) of claim 1,

5 in a second step the amino acid sequence of another protein which is homologous to the consensus sequence is compared with the consensus sequence and

in a third step only those amino acid residues are replaced in the amino acid sequence of the other protein which clearly differ from the consensus sequence of this protein family calculated under moderately stringent conditions whereas at all positions of the alignment where no preferred single amino acid can be determined under moderately stringent

10 conditions the amino acids of the other protein remain unchanged.

4. A process as claimed in any one of claims 1-3, wherein

in a first step a consensus sequence is determined from homologous sequences according to steps a), b) and c) of claim 1,

15 in a second step the active center of the protein comprising all amino acid residues that are involved in forming the active center is determined in the consensus sequence and in the sequence of a homologous protein as well and

in a third step some or all of the amino acids that form the active center of the homologous protein are inserted in the backbone of the consensus sequence.

5. A process as claimed in claim 4, wherein the active center of the protein is determined by using an analysis of the three-dimensional structure of the protein.

20 6. A process as claimed in claims 4 and 5, wherein the homologous protein is an enzyme.

7. A process as claimed in claims 1 to 6, wherein the defined protein family is the family of phytases.

25 8. A process as claimed in claim 7, wherein the phytases are of fungal origin.

9. A process as claimed in claims 7 or 8, wherein the amino acid sequence of the phytase is changed by the introduction of at least one mutation selected from the group consisting of

E58A	F54Y
D69K	I73V
D197N	K94A
T214L	R101A
E222T	N153K
E267D	V158I
R291I	A203G
R329H	S205G
S364T	V217A
A379K	A227V
G404A	V234L
	P238A
	Q277E
	A287H
	A292Q
	V366I
	A396S
	E415Q
	G437A
	E451R

whereby the number represents the position in the consensus phytase sequence or a corresponding residue according to an alignment as shown in Fig. 1 when 26 amino acids (signal sequence) are added to the sequences shown in Fig. 1 and the letter before the number represents the amino acid in the phytase which is replaced by the amino acid
5 behind the number.

10. A process as claimed in any one of claims 1 to 9, wherein the host cell is of eukaryotic origin.

11. A process as claimed in claim 10, wherein eukaryotic means fungal, preferably *Aspergillus* or yeast, preferably *Saccharomyces* or *Hansenula*.
12. A consensus protein obtainable preferably obtained by a process as claimed in any one of claims 1 to 11.
- 5 13. A consensus protein which comprises the amino acid sequence shown in Figure 2 or any variants or muteins thereof (consensus phytase-1).
14. A mutein of the consensus protein of claim 13 characterized therein that in the amino acid sequence of Figure 2 the following replacements have been effected Q50L, Q50T, Q50G, Q50T-Y51N, Q50L-Y51N or Q50T-K91A.
- 10 15. A consensus protein which comprises the amino acid sequence shown in Figure 4 having the designation consensus phytase 10 (Fcp10) and any variants or muteins thereof.
16. A consensus protein which comprises the amino acid sequence shown in Figure 6 having the designation Consensus seq. 11 and any variant or mutein thereof.
- 15 17. A consensus protein which comprises the amino acid sequence shown in Figure 10 (consensus phytase 7) and any variant or mutein thereof.
18. A consensus protein which comprises the amino acid sequence shown in Figure 21 (consensus phytase 12) and any variant or mutein thereof.
19. A consensus protein which comprises the amino acid sequence shown in Figure 3 (basidio consensus) and any variant or mutein thereof.
- 20 20. A phytase being selected from amongst: *A. fumigatus* ATCC 13073 alpha-mutant; *A. fumigatus* ATCC 13073 alpha-mutant-(E59A-S126N-R329H, S364T-G404A)-Q27T; *A. fumigatus* ATCC 13073 alpha-mutant-(E59A, S126N-R329H-S364T-G404A)-Q27T-K68A, preferably the latter.
21. A food, feed or pharmaceutical composition comprising a consensus protein as claimed in any of the claims 12 to 17.

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Figure 1

1

50
 5 *A. terreus* 9A-1 KhsDCNSVDh GYQCFPELSH kWG1YAPYFS
 LQDESPFP1D VPEDChITFV
A. terreus cbs NhsDCTSVDr GYQCFPELSH kWG1YAPYFS
 LQDESPFP1D VPDDChITFV
A. niger var. *awamori* NqsTCDTVdq GYQCFSETSH LWGQYAPFFS
 10 LANESAISPD VPAGCrVTFA
A. niger T213 NqsSCDTVDQ GYQCFSETSH LWGQYAPFFS
 LANESVISPD VPAGCrVTFA
A. niger NRRL3135 NqbSCDTVDQ GYQCFSETSH LWGQYAPFFS
 LANESVISPE VPAGCrVTFA
 15 *A. fumigatus* 13073 GSkSCDTVD1 GYQCSPATSH LWGQYSPFFS
 LEDELSVSSK LPKDCrITLV
A. fumigatus 32722 GSkSCDTVD1 GYQCSPATSH LWGQYSPFFS
 LEDELSVSSK LPKDCrITLV
A. fumigatus 58128 GSkSCDTVD1 GYQCSPATSH LWGQYSPFFS
 20 LEDELSVSSK LPKDCrITLV
A. fumigatus 26906 GSkSCDTVD1 GYQCSPATSH LWGQYSPFFS
 LEDELSVSSK LPKDCrITLV
A. fumigatus 32239 GSkACDTVE1 GYQCSPGTSH LWGQYSPFFS
 LEDELSVSSD LPKDCrVTFF
 25 *E. nidulans* QNHSCNTADG GYQCFPNVSH VWGQYSPYFS
 IEQESAISED VPHGCeVTFV
T. thermophilus DSHSCNTVEG GYQCPEISH sWGQYSPFFS
 LADQSEISPD VPQNCKITFV
M. thermophila ESRPCDTPD1 GFQCgTAISH FWGQYSPYFS
 30 VpSE1DaS.. IPDDCeVTFA

 Consensus NSHSCDTVDG GYQCFPEISH LWGQYSPYFS
 LEDESAISPD VPDDC-VTFV
 Consensus phytase NSHSCDTVDG GYQCFPEISH LWGQYSPYFS
 35 LEDESAISPD VPDDCRVTFV

51

100
 40 *A. terreus* 9A-1 QVLARHGARS PThSKtKAYA AtIAAIQKSA
 TaFpGKYAFL QSYNSLDSE
A. terreus cbs QVLARHGARS PTDSKtKAYA AtIAAIQKNA
 TaLpGKYAFL KSYNSMGSE
A. niger var. *awamori* QVLSRHGARY PTESKgKKYS ALIEEIQQNV
 45 TtFDGKYAFL KTNYSLGAD
A. niger T213 QVLSRHGARY PTESKgKKYS ALIEEIQQNV
 TtFDGKYAFL KTNYSLGAD
A. niger NRRL3135 QVLSRHGARY PTDSKgKKYS ALIEEIQQNA
 TtFDGKYAFL KTNYSLGAD
 50 *A. fumigatus* 13073 QVLSRHGARY PTSSKsKKYK kLVTAlQaNA
 TdFKGKFAFL KTNYTLGAD
A. fumigatus 32722 QVLSRHGARY PTSSKsKKYK kLVTAlQaNA
 TdFKGKFAFL KTNYTLGAD
A. fumigatus 58128 QVLSRHGARY PTSSKsKKYK kLVTAlQaNA
 55 TdFKGKFAFL KTNYTLGAD

Modtaget PD

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	<i>A. fumigatus</i> 26906	QVLSRHGARY PTSSKsKkYK kLVTAlQaNA
	TdFKGKFAFL KTYNYTLGAD	
	<i>A. fumigatus</i> 32239	QVLSRHGARY PTASKsKkYK kLVTAlQKNA
	TeFKGKFAFL ETYNYTLGAD	
5	<i>E. nidulans</i>	QVLSRHGARY PTESKsKAYS GLIEAIQKNA
	TsFwGQY AFL ESYNYTLGAD	
	<i>T. thermophilus</i>	QLLSRHGARY PTSSKtE1YS QLISriQKTA
	TaYKGyY AFL KDYrYqLGAN	
	<i>M. thermophila</i>	QVLSRHGARa PT1KRaaSYv DLIDrIHHGA
10	IsYgPgYEFL RTYDYTLGAD	
	Consensus	QVLSRHGARY PTSSK-KAYS ALIEAIQKNA T-
	FKGKY AFL KTYNYTLGAD	
	Consensus phytase	QVLSRHGARY PTSSKSKAYS ALIEAIQKNA
15	TAFKGKY AFL KTYNYTLGAD	

101

	150	
20	<i>A. terreus</i> 9A-1	ELTPFGrNQL rD1GaQFYeR YNALTRhInP
	FVRATDASRV hESAEEKFVEG	
	<i>A. terreus</i> cbs	NLTPFGrNQL qD1GaQFYRR YDTLTrhInP
	FVRAADSSRV hESAEEKFVEG	
	<i>A. niger</i> var. <i>awamori</i>	DLTPFGEQEL VNSGIKFYQR YESLTRNIIP
25	FIRSSGSSRV IASGEKFIEG	
	<i>A. niger</i> T213	DLTPFGEQEL VNSGIKFYQR YESLTRNIIP
	FIRSSGSSRV IASGEKFIEG	
	<i>A. niger</i> NRRL3135	DLTPFGEQEL VNSGIKFYQR YESLTRNIVP
	FIRSSGSSRV IASGKKFIEG	
30	<i>A. fumigatus</i> 13073	DLTPFGEQQL VNSGIKFYQR YKALARSVVP
	FIRASGSDRV IASGEKFIEG	
	<i>A. fumigatus</i> 32722	DLTPFGEQQL VNSGIKFYQR YKALARSVVP
	FIRASGSDRV IASGEKFIEG	
	<i>A. fumigatus</i> 58128	DLTPFGEQQL VNSGIKFYQR YKALARSVVP
35	FIRASGSDRV IASGEKFIEG	
	<i>A. fumigatus</i> 26906	DLTAFGEQQL VNSGIKFYQR YKALARSVVP
	FIRASGSDRV IASGEKFIEG	
	<i>A. fumigatus</i> 32239	DLTPFGEQQM VNSGIKFYQK YKALAgSVVP
	FIRSSGSDRV IASGEKFIEG	
40	<i>E. nidulans</i>	DLTifGENQM VDSGaKFYRR YKNLARKnTP
	FIRASGSDRV VASAEEKFING	
	<i>T. thermophilus</i>	DLTPFGENQM IQLGIKFYnH YKSLARNaVP
	FVRCSGSDRV IASGr1FIEG	
	<i>M. thermophila</i>	ELTRtGQQQM VNSGIKFYRR YRALARKsIP
45	FVRTAGqDRV VhSAENFTQG	
	Consensus	DLTPFGENQM VNSGIKFYRR YKALARK-VP
	FVRASGSDRV IASAEKFIEG	
	Consensus phytase	DLTPFGENQM VNSGIKFYRR YKALARKIVP
50	FIRASGSDRV IASAEKFIEG	

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200

A. *terreus* 9A-1 FQTARqDDHh ANpHQPSPrV DVaIPEGSAY
 NNTLEHS1CT AFES...STV

5 A. *terreus* cbs FQNARqGDPH ANpHQPSPrV DVVIPEGTAY
 NNTLEHS1CT AFEA...STV

A. *niger* var. *awamori* FQSTKLkDPr AqpgQSSPKI DVVISEASSs
 NNTLDPGTCT VFED...SEL

A. *niger* T213 FQSTKLkDPr AqpgQSSPKI DVVISEASSs

10 NNTLDPGTCT VFED...SEL

A. *niger* NRRL3135 FQSTKLkDPr AqpgQSSPKI DVVISEASSs
 NNTLDPGTCT VFED...SEL

A. *fumigatus* 13073 FQqAKLADPG A.TNRAAPAI SVIYPESETF
 NNTLDHGVCT kFEA...SQL

15 A. *fumigatus* 32722 FQqAKLADPG A.TNRAAPAI SVIYPESETF
 NNTLDHGVCT kFEA...SQL

A. *fumigatus* 58128 FQqAKLADPG A.TNRAAPAI SVIYPESETF
 NNTLDHGVCT kFEA...SQL

A. *fumigatus* 26906 FQqAKLADPG A.TNRAAPAI SVIYPESETF

20 NNTLDHGVCT kFEA...SQL

A. *fumigatus* 32239 FQqANVADPG A.TNRAAPVI SVIYPESETY
 NNTLDHSVCT NFEA...SEL

E. *nidulans* FRKAQLhDHG S..gQATPVV NVIIPEiDGF
 NNTLDHSTCV SFEN...DER

25 T. *thermophilus* FQSAKVLDPH SDKHDAPPTI NVIIeEGPSY
 NNTLDtGSCP VFED...SSg

M. *thermophila* FHSALLADRG STvRPTlPyd mVVIPETAGa
 NNTLHND1CT AFEEgpySTI

30 Consensus FQSAKLADEPG S-PHQASPVI NVIIPEGSGY
 NNTLDHGTCT AFED---SEL

Consensus phytase FQSAKLADEPG SQPHQASPVI DVIIPEGSGY
 NNTLDHGTCT AFED...SEL

35

201

250

A. *terreus* 9A-1 GDDAvANFTA VFAPAIaQRL EADLPGVqLS
 TDDVVnLMAM CPFETVS1TD

40 A. *terreus* cbs GDAAAADNFTA VFAPAIakRL EADLPGVqLS
 ADDVVnLMAM CPFETVS1TD

A. *niger* var. *awamori* ADTVEANFTA TFAPSIRQL ENDLSGVTLT
 DTEVTVyLMDM CSFDTISTST

A. *niger* T213 ADTVEANFTA TFAPSIRQL ENDLSGVTLT

45 DTEVTVyLMDM CSFDTISTST

A. *niger* NRRL3135 ADTVEANFTA TFVPSIRQL ENDLSGVTLT

DTEVTVyLMDM CSFDTISTST

A. *fumigatus* 13073 GDEVAANFTA 1FAPDIRARa EkHLPGVTLT
 DEDVVVsLMDM CSFDTVARTS

50 A. *fumigatus* 32722 GDEVAANFTA 1FAPDIRARa EkHLPGVTLT
 DEDVVVsLMDM CSFDTVARTS

A. *fumigatus* 58128 GDEVAANFTA 1FAPDIRARa EkHLPGVTLT
 DEDVVVsLMDM CSFDTVARTS

A. *fumigatus* 26906 GDEVAANFTA 1FAPDIRARa KkHLPGVTLT

55 DEDVVVsLMDM CSFDTVARTS

A. *fumigatus* 32239 GDEVEANFTA 1FAPPAIRARI EkHLPGVqLT
 DDDVVVsLMDM CSFDTVARTA

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	<i>E. nidulans</i>	ADEiEANFTA IMGPIRKRL ENDLPGIKLT
	NENVIyLMDM CSFDTMARTA	
	<i>T. thermophilus</i>	GHDAQEKFak qFAPAIKEKI KDHLPGVDLA
	vSDVpyLMDL CPFETLARNh	
5	<i>M. thermophila</i>	GDDAQDTY1S TFAGPitARV NANLPGANLT
	DADTVaLMDL CPFETVASSS	
	Consensus	GDDAEANFTA TFAPAIRARL EADLPGVTLT DEDVV-
	LMDM CPFETVARTS	
10	Consensus phytase	GDDVEANFTA LFAPAIRARL EADLPGVTLT
	DEDVVYLMMDM CPFETVARTS	

		251
15	300	
	<i>A. terreus</i> 9A-1DAhTLSPFC DLFTAtEWtq
	YNYL1SLDKY YGYGGGNPLG	
	<i>A. terreus</i> cbsDAhTLSPFC DLFTAAEWtq
	YNYL1SLDKY YGYGGGNPLG	
20	<i>A. niger</i> var. <i>awamori</i>vDTKLSPFC DLFTHdEWih
	YDYLQSLkKY YGHGAGNPLG	
	<i>A. niger</i> T213vDTKLSPFC DLFTHdEWih
	YDYLRSLSkKY YGHGAGNPLG	
	<i>A. niger</i> NRRL3135vDTKLSPFC DLFTHdEWin
25	YDYLQSLkKY YGHGAGNPLG	
	<i>A. fumigatus</i> 13073DASQLSPFC QLFTHnEWkk
	YNYLQSLGKY YGYGAGNPLG	
	<i>A. fumigatus</i> 32722DASQLSPFC QLFTHnEWkk
	YNYLQSLGKY YGYGAGNPLG	
30	<i>A. fumigatus</i> 58128DASQLSPFC QLFTHnEWkk
	YNYLQSLGKY YGYGAGNPLG	
	<i>A. fumigatus</i> 26906DASQLSPFC QLFTHnEWkk
	YNYLQSLGKY YGYGAGNPLG	
	<i>A. fumigatus</i> 32239DASELSPFC AIFTHnEWkk
35	YDYLQSLGKY YGYGAGNPLG	
	<i>E. nidulans</i>HGTELSPFC AIFTEkEWlq
	YDYLQSLSKY YGYGAGSPLG	
	<i>T. thermophilus</i>TDT.LSPFC ALstTQeEWqa
	YDYYQSLGKY YGnGGGNPLG	
40	<i>M. thermophila</i>	sdpatadaggyNGrpLSPFC rLFSEsEWra
	YDYLQSVGKw YGYGPGNPLG	
	Consensus	----- -DATELSPFC ALFTE-EW--
	YDYLQSLGKY YGYGAGNPLG	
45	Consensus phytaseDATELSPFC ALFTHDEWRQ
	YDYLQSLGKY YGYGAGNPLG	

301

350

A. terreus 9A-1	PVQGVGVWaNE LMARLTRAPV HDHTCVNNTL
DASPATFPLN ATLYADFSHD	
5 A. terreus cbs	PVQGVGVWaNE LIARLTRSPV HDHTCVNNTL
DANPATFPLN ATLYADFSHD	
A. niger var. awamori	PTQGVGYaNE LIARLTHSPV HDDTSSNHTL
DSNPATFPLN STLYADFSHD	
A. niger T213	PTQGVGYaNE LIARLTHSPV HDDTSSNHTL
10 DSNPATFPLN STLYADFSHD	
A. niger NRRL3135	PTQGVGYaNE LIARLTHSPV HDDTSSNHTL
DSSPATFPLN STLYADFSHD	
A. fumigatus 13073	PAQGIGfNE LIARLTRSPV QDHTSTNstL
vSNPATFPLN ATMYVDFSHD	
15 A. fumigatus 32722	PAQGIGfNE LIARLTRSPV QDHTSTNstL
vSNPATFPLN ATMYVDFSHD	
A. fumigatus 58128	PAQGIGfNE LIARLTRSPV QDHTSTNstL
vSNPATFPLN ATMYVDFSHD	
20 A. fumigatus 26906	PAQGIGfNE LIARLTRSPV QDHTSTNstL
vSNPATFPLN ATMYVDFSHD	
A. fumigatus 32239	PAQGIGfNE LIARLTNSPV QDHTSTNstL
DSDPATFPLN ATIYVDFSHD	
E. nidulans	PAQGIGfNE LIARLTQSPV QDNTSTNHTL
25 DSNPATFPLD rKLYADFSHD	
T. thermophilus	PAQGVGFvNE LIARMTHSPV QDYTTVNHTL
DSNPATFPLN ATLYADFSHD	
M. thermophila	PTQGVGFvNE LLARLAGvPV RDgtSTNRTL
DGDPrTFPLG rPLYADFSHD	
30 Consensus	PAQGVGF-NE LIARLTHSPV QDHTSTNHTL
DSNPATFPLN ATLYADFSHD	
Consensus phytase	PAQGVGFANE LIARLTRSPV QDHTSTNHTL
DSNPATFPLN ATLYADFSHD	

35

351

400	
A. terreus 9A-1	SNLVSIFWAL GLYNGTAPLS qTSVESVSQT
DGYAAAWTVP FAARAYVEMM	
40 A. terreus cbs	SNLVSIFWAL GLYNGTkPLS qTTVEDITrT
DGYAAAWTVP FAARAYIEMM	
A. niger var. awamori	NGIISILFAL GLYNGTkPLS TTTVENITQT
DGFSSAWTVP FASR1YVEMM	
A. niger T213	NGIISILFAL GLYNGTkPLS TTTVENITQT
45 DGFSSAWTVP FASR1YVEMM	
A. niger NRRL3135	NGIISILFAL GLYNGTkPLS TTTVENITQT
DGFSSAWTVP FASR1YVEMM	
A. fumigatus 13073	NSMVSIFFAL GLYNGTEPLS rTSVESAkEL
DGYSASWVVP FGARAYFETM	
50 A. fumigatus 32722	NSMVSIFFAL GLYNGTGPLS rTSVESAkEL
DGYSASWVVP FGARAYFETM	
A. fumigatus 58128	NSMVSIFFAL GLYNGTEPLS rTSVESAkEL
DGYSASWVVP FGARAYFETM	
A. fumigatus 26906	NSMVSIFFAL GLYNGTEPLS rTSVESAkEL
55 DGYSASWVVP FGARAYFETM	
A. fumigatus 32239	NGMIPIFFAM GLYNGTEPLS qTSeESTKES
NGYSASWAVP FGARAYFETM	

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	<i>E. nidulans</i>	NSMISIFFAM GLYNGTQPLS mDSVESIQEm
	DGYAASWTVP FGARAYFELM	
	<i>T. thermophilus</i>	NTMTSIFaAL GLYNGTAKLS TTEIKSIEET
	DGYSAAWTVP FGGRAYIEMM	
5	<i>M. thermophila</i>	NDMMGVlgaAL GaYDGVPPLD KTArrDpEE1
	GGYAASAWVP FAARIYVEKM	
	Consensus	NSMISIFFAL GLYNGTAPLS TTSVESIEET
	DGYAASWTVP FGARAYVEMM	
10	Consensus phytase	NSMISIFFAL GLYNGTAPLS TTSVESIEET
	DGYSASWTVP FGARAYVEMM	

401

15	450	
	<i>A. terreus</i> 9A-1	QC..... RAEKE PLVRVLVNDR
	VMPLHGCTD KLGRCKrDAD	
	<i>A. terreus</i> cbs	QC..... RAEKQ PLVRVLVNDR
	VMPLHGCAVD NLGRCKrDDF	
20	<i>A. niger</i> var. <i>awamori</i>	QC..... QAEQE PLVRVLVNDR
	VVPLHGCPID aLGRCTRDSF	
	<i>A. niger</i> T213	QC..... QAEQE PLVRVLVNDR
	VVPLHGCPID aLGRCTRDSF	
	<i>A. niger</i> NRRL3135	QC..... QAEQE PLVRVLVNDR
25	VVPLHGCPVD aLGRCTRDSF	
	<i>A. fumigatus</i> 13073	QC..... KSEKE PLVRALINDR
	VVPLHGCDVD KLGRCKLNDF	
	<i>A. fumigatus</i> 32722	QC..... KSEKE PLVRALINDR
	VVPLHGCDVD KLGRCKLNDF	
30	<i>A. fumigatus</i> 58128	QC..... KSEKE SLVRALINDR
	VVPLHGCDVD KLGRCKLNDF	
	<i>A. fumigatus</i> 26906	QC..... KSEKE PLVRALINDR
	VVPLHGCDVD KLGRCKLNDF	
	<i>A. fumigatus</i> 32239	QC..... KSEKE PLVRALINDR
35	VVPLHGCAVD KLGRCKLKDF	
	<i>E. nidulans</i>	QC..... E.KKE PLVRVLVNDR
	VVPLHGCAVD KFGRCTLDDW	
	<i>T. thermophilus</i>	QC..... DDSDE PVVRVLVNDR
	VVPLHGCEVD SLGRCKrDDF	
40	<i>M. thermophila</i>	RCsggggggg ggegrQEKE eMVRVLVNDR
	VMTLkGCGAD ErGMCTLERF	
	Consensus	QC----- QAEKE PLVRVLVNDR
	VVPLHGCAVD KLGRCKLDDF	
45	Consensus phytase	QC..... QAEKE PLVRVLVNDR
	VVPLHGCAVD KLGRCKrDDF	

451

	471	
50	<i>A. terreus</i> 9A-1	VAGLSFAQAG GNWADCF--- ~
	<i>A. terreus</i> cbs	VEGLSFARAG
	GNWAECF--- ~	
	<i>A. niger</i> var. <i>awamori</i>	VrGLSFARSG GDWAECsA--- ~
	<i>A. niger</i> T213	VrGLSFARSG GDWAECFA--- ~
55	<i>A. niger</i> NRRL3135	VrGLSFARSG
	GDWAECFA--- ~	
	<i>A. fumigatus</i> 13073	VKGGLSWARSG GNWGECFS--- ~
	<i>A. fumigatus</i> 32722	VKGGLSWARSG GNWGECFS--- ~

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	<i>A. fumigatus</i> 58128	VKGLSWARSG GNWGECKS--
	<i>A. fumigatus</i> 26906	VKGLSWARSG GNWGECKS--
	<i>A. fumigatus</i> 32239	VKGLSWARSG
	GNSEQSFS--	
5	<i>E. nidulans</i>	VEGLNFARSG GNWKTCTF1--
	<i>T. thermophilus</i>	VrGLSFARqG GNWEGCYAas e
	<i>M. thermophila</i>	IESMAFARGN GKWD1CFA--
	Consensus	VEGLSFARSG GNWAECFAD--
10	Consensus phytase	VEGLSFARSG GNWAECFAD..

Figure 2

CP-1

15 ECO RI M G V F V V L L S I A T L F G S T
TATATGAATTCTATGGGCGTGTCTCGTCGTGCTACTGTCCATTGCCACCTTGGTTCGGTTCCA

1 -----+-----+-----+-----+-----+-----+-----+ 60

ATATACTTAAGTACCCGACAAAGCAGCACGATGACAGGTAACGGTGGAAACAAGCCAAGGT

W G T A L G P R G N S H S C D T V D G G

20 CATCCGGTACCGCCTGGGTCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGTG

-----+-----+-----+-----+-----+-----+-----+-----+-----+-----+

CP-2

CP-3

25 Y Q C F P E I S H D W G Q Y S P Y F S L
GTTACCAATGTTCCCAGAAATTCTCACTTGTGGGGTCAATACTCTCCATACTTCTTT

121 -----+-----+-----+-----+-----+-----+-----+ 180

CAATGGTTACAAAGGGTCTTAAAGAGTGAACACCCAGTTATGAGAGGTATGAAGAGAA

30 E D E S A I S P D V P D D C R V T F V Q

TGGAAAGACGAATCTGCTATTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTCGTTCT

181 - - - - + - - - - + - - - - + - - - - + - - - - + - - - - + 240

ACCTTCTGCTTAGACGATAAAGAGGCTGCAAGGCTGCTGACATCTCAATGAAAGCAAG

CP-4

Modtaget PD

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241 -----+-----+-----+-----+-----+-----+ 300

TTCAAAACAGATCTGTGCCACGATCTATGGTTGAAGAAGATTAGATTCCGAATGAGAC

L I E A I Q K N A T A F K G K Y A F L K

5 CTTTGATTGAAGCTATTCAAAAGAACGCTACTGCTTCAAGGGTAAGTACCGCTTCTTGA

301 -----+-----+-----+-----+-----+-----+ 360

GAAACTAACCGATAAGTTCTTGCATGACGAAAGTTCCCATTCAATGCCAAAGAAACT

CP-6

CP-7

10 T Y N Y T L G A D D L T P F G E N Q M V

AGACTTACAACACTTGGGTGCTGACGACTTGACTCCATTGGTAAAACCAATGG

361 -----+-----+-----+-----+-----+-----+ 420

TCTGAATGTTGATGTGAAACCCACGACTGCTGAAGTGGTAAGCCACTTTGGTTTACC

15 N S G I K F Y R R Y K A L A R K I V P F

TTAACTCTGGTATTAAGTTCTACAGAACGATACAAGGTTGGCTAGAAAGATTGTTCCAT

421 -----+-----+-----+-----+-----+-----+ 480

AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCAAACGATCTTCAACAAGGTA

CP-8

20 CP-9

I R A S G S D R V I A S A E K F I E G F

481 -----+-----+-----+-----+-----+-----+ 540
TCATTAGAGCTTCTGGTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCAATTGAAGGTT
AGTAATCTCGAAGACCAAGACTGTCTCAATAACGAAGACGACTTTCAAGTAACCTCCAA

25

Q S A K L A D P G S Q P H Q A S P V I D
TCCAATCTGCTAAGTTGGCTGACCGAGGTTCTCAACCACACCAAGCTTCTCCAGTTATTG

541 -----+-----+-----+-----+-----+-----+ 600

AGGTTAGACGATTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTCAAGAGGGTCAATAAC

30

CP-10

CP-11

V I I P E G S G Y N N T L D H G T C T A
ACGTTATTATTCCAGAAGGATCCGGTTACAACACACTTGGACCACGGTACTTGTACTG

601 -----+-----+-----+-----+-----+-----+ 660

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TGCAATAATAAGGTCTTCCtAGgCCAATGTTGTTGAAACCTGGTGCCATGAACATGAC

F E D S E L G D D V E A N F T A L F A P

CTTCGAAGACTCTGAATTGGGTGACGACGTTGAAGCTAACCTCACTGCTTGGCGCTC

5 661 -----+-----+-----+-----+-----+-----+ 720

GAAAGCTTCTGAGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGAAACAAGCGAG

CP-12

A I R A R L E A D L P G V T L T D E D V

10 CAGCTATTAGAGCTAGATTGGAAGCTGACTGCCAGGTGTTACTTGACTGACGAAAGACG

721 -----+-----+-----+-----+-----+-----+ 780

GTCGATAATCTCGATCTAACCTCGACTGAACGGTCCACAATGAAACTGACTGCTTCTGC

CP-13

15 V Y L M D M C P F E T V A R T S D A T E

TTGTTTACTTGATGGACATGTGTCCTCGAAACTGTTGCTAGAACTTCTGACGCTACTG

781 -----+-----+-----+-----+-----+-----+ 840

AACAAATGAACTACCTGTACACAGGTAAGCTTGACAACGATCTGAAAGACTGCGATGAC

20 L S P F C A L F T H D E W R Q Y D Y L Q

AATTGTCCTCATTCTGTGCTTGTACTCACGACGAATGGAGACAATACGACTACTTGC

841 -----+-----+-----+-----+-----+-----+ 900

TTAACAGAGGTAAGAACGAAACAAGTGAGTGCTGCTTACCTCTGTTATGCTGATGAAACG

CP-14

25 CP-15

S L G K Y Y G Y G A G N P L G P A Q G V

AATCTTGGTAAGTACTACGGTTACGGTGTGGTAACCCATTGGTCCAGCTCAAGGTG

901 -----+-----+-----+-----+-----+-----+ 960

TTAGAAACCCATTGATGCCATGCCAACGACCATTGGTAACCCAGGTCGAGTTCCAC

30 G F A N E L I A R L T R S P V Q D H T S

TTGGTTTGTGCTAACGAAATTGATTGCTAGATTGACTAGATCTCCAGTTCAAGACCACACTT

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961 -----+-----+-----+-----+-----+
 1020

AACCAAAGCGATTGCTTAACTAACGATCTAACTGATCTAGAGGTCAAGTTCTGGTGTGAA

CP-16
 5 CP-17

T N H T L D S N P A T F P L N A T L Y A
 CTACTAACACACTTGGACTCTAACCCAGCTACTTCCCATTGAACGCTACTTGTACG

1021 -----+-----+-----+-----+-----+
 1080

GATGATTGGTGTGAAACCTGAGATTGGGTGATGAAAGGGTAACTTGCGATGAAACATGC

D F S H D N S M I S I F F A L G L Y N G
 CTGACTTCTCTCACGACAACTCTATGATTCTATTCTCGCTTGGTTGTACAACG

1081 -----+-----+-----+-----+-----+
 15 1140

GACTGAAGAGAGTGCTGTTGAGATACTAAAGATAAAAGAAGCGAAACCAAACATGTTGC

CP-18
 CP-19

T A P L S T T S V E S I E E T D G Y S A
 GTACTGCTCCATTGCTACTACTTCTGTTGAATCTATTGAAGAAACTGACGGTTACTCTG

1141 -----+-----+-----+-----+-----+
 1200

CATGACGAGGTAACAGATGATGAAGACAACTTAGATAACCTCTTGAUTGCCAATGAGAC

S W T V P F G A R A Y V E M M Q C Q A E
 CTTCTGGACTGTTCCATTGGTGTAGAGCTTACGTTGAATGCAATGTCAAGCTG

1201 -----+-----+-----+-----+-----+
 1260

GAAGAACCTGACAAGGTAAAGCCACGATCTGAATGCAACTTACTACGTTACAGTTGAC

CP-20
 CP-21

K E P L V R V L V N D R V V P L H G C A
 AAAAGGAACCATTGGTTAGAGTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG

1261 -----+-----+-----+-----+-----+
 35 1320

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TTTCCTGGTAACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACAC

V D K L G R C K R D D F V E G L S F A R

CTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTGTCTTCGCTA

5 1321 -----+-----+-----+-----+-----+-----+
1380

GACAACTGTTCAACCCATCTACATTCTCTGCTGAAGCAACTTCAAACAGAAAGCGAT

CP-22

S G G N W A E C F A * Eco RI

10 GATCTGGTGGTAACTGGGCTGAATGTTCGCTTAAGAATTCAATA

1381 -----+-----+-----+-----+-----+-----+ 1426

CTAGACCACCATTGACCCGACTTACAAAGCGAATTCTTAAGTATA

15

Figure 3

1

50

P. involutus (phyA1) SvP.KnTAp^t FPIPeseQrn WSPYSPYFPL AeYkAPPAGC
20 QInQVNIIQRP. involutus (phyA2) SvP.RniAPK FSIPeseQrn WSPYSPYFPL AeYkAPPAGC
EInQVNIIQRT. pubescens hiPlRdTSAc LdVTrDvQqs WSmYSPYFPa AtYvAPPASC
QInQVHIIQR25 A. pediades GgvvQaTfvQ pfFPpQiQds WAAYTPYYPV qaYtPPPkDC
KITQVNIIQRP. lycii StQfsfvAAQ LPIP_aQnts_n WGPYdPFFPV EpYaAPPEGC
tvtQVNLIQR30 Basidio S-P-R-TAAQ LPIP-Q-Q-- WSPYSPYFPV A-Y-APPAGC QI-
QVNIIQR

Modtaget PD

- 60 -

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51

100

P. involutus (phyA1) HGARFPTSGA TTRIKAGLTK LQGvqnfTDA KFNFIksfkY
dLGnsDLVPF

5 *P. involutus* (phyA2) HGARFPTSGA ATRIKAGLSK LQSvqnfTDP KFDFIKsFTY
dLGtsDLVPF

T. pubescens HGARFPTSGA AkRIQTAVAK LKAAsnyTDP 1LAFVtNyTY
sLGqDsLVeL

10 *A. pediades* HGARFPTSGA GTRIQAAVkk LQSAktYtDP RLDFLtNyTY
tLGhDDLVPF

P. lycii HGARWPTSGA rSRqvAAVAK IQmArpftDP KYEFLnDfvY
kFGvADLLPF

15 **Basidio** HGARFPTSGA ATRIQAAVAK LQSA---TDP KLDFL-N-TY -LG-
15 **DDLVPF**

101

150

20 *P. involutus* (phyA1) GAaQSfDAGQ EAFArYskLV SkNNLPFIRA dGSDRVVDSA
TNWTAGFAsA

P. involutus (phyA2) GAaQSfDAG1 EvFARYSkLV SsDNLPFIRS dGSDRVVDTA
TNWTAGFAsA

25 *T. pubescens* GAtQSSEAGQ EAFTrySsLV SaDELPFVRA SGSDRVVATA
nNWTAGFAlA

A. pediades GALQSSQAGE ETFqRYSfLV SkENLPFVRA SSSNRVVDSA
TNWTEGFSAA

P. lycii GAQShQTGt DmYTRYStLf egGDVPFVRA AGdQRVVVDSS
TNWTAGFGdA

30 **Basidio** GA-QSSQAGQ EAFTryS-LV S-DNLPFVRA SGSDRVVDSA
TNWTAGFA-A

151

200

P. involutus (phyA1) ShNTvqPkLn LILPQtGNDT LEDNMCPaAG DSDPQvNaWL
AVafPSITAR

40 *P. involutus* (phyA2) SrNAiqPkLd LILPQtGNDT LEDNMCPaAG ESDPQvDaWL
AsafPSVTAQ

	<i>T. pubescens</i> AqFAPPMTAR	SsNSitPvLs VIISeaGNDT LDDNMCPaAG DSDPQvNqWL
	<i>A. pediades</i> SIYGTPIAAnR	ShHvlnPiLf VILSEs1NDT LDDaMCPnAG sSDPQtGiWt
5	<i>P. lycii</i> GVFAPnITAR	SgETvlPtLq VVLqEeGNcT LcNNMCPnEv DGDest.tWL
	Basidio AVFAPPITAR	S-NT--P-L- VILSE-GNDT LDDNMCP-AG DSDPQ-N-WL
10		
		201
	250	
15	<i>P. involutus</i> (phyA1) giPGsFeAFa	LNAAAPSvNL TDtDAfNLvs LCAF1TVSkE kkSdFCtLFE
	<i>P. involutus</i> (phyA2) giPGsFeAFa	LNAAAPGANL TDaDAfNLvs LCPFmTVSkE qkSdFCtLFE
	<i>T. pubescens</i> elQAE.dAFa	LNAGAPGANL TDtDTyNLlt LCPFETVAtE rrSeFCDIYE
20	<i>A. pediades</i> .tPEEFaqFe	LNqqAPGANI TAaDvsNLip LCAFETIVkE tpSpFCNLF.
	<i>P. lycii</i> .tAEEYvSYe	LNAAAPSANL SDsDAltLmd MCPFDLSSg naSpFCDLF.
25	Basidio AF-	LNAAAPGANL TD-DA-NL-- LCPFETVS-E --S-FCDLF --PEEF-
		251
30	300	
	<i>P. involutus</i> (phyA1) NTQTNRNRTLDA	YgGDLDKFYG TGyGQeLGPV QGVGYVNELI ARLTnsAVRD
	<i>P. involutus</i> (phyA2) NTQTNRNRTLDA	YaGDLDKFYG TGyGQALGPV QGVGYINELL ARLTnsAVnD
35	<i>T. pubescens</i> HTQTNstLDS	YnADLDKFYG TGyGQPLGPV QGVGYINELI ARLTaQnVSD
	<i>A. pediades</i> NTQTNRNRTLDS	YfGDLDKFYG TGyGQPLGPV QGVGYINELL ARLTemPVRD
40	<i>P. lycii</i> ETQTNRNRTLDS	YyyDLDKYYG TGpGNALGPV QGVGYVNELL ARLTgQAVRD

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Basidio
NTQTNRTLDS

Y-GDLDKFYG TGYGQPLGPV QGVGYINELL ARLT-QAVRD

5

301

350

P. involutus (phyA1) SPvTFPLNKT FYADFSHDN1 MVAVFSAMGL FrQPAPLsTS
vPNPwRTWrT

10 *P. involutus* (phyA2) APdTFPLNKT MYADFSHDN1 MVAVFSAMGL FrQSAPLsTS
tPDPNRTWLT

T. pubescens SPETFPLNRT LYADFSHDNQ MVAIFSAMGL FNQSAPLDPT
tPDPaRTFLv

15 *A. pediades* SP1TFPLDRS IYADLSHDNQ MIAIFSAMGL FNQSSPLDPS
fPNPKRTWVT

P. lycii dPaTFPLNRT FYADFSHDNT MVPIFAALGL FNaTA.LDPL
kPDeNR1Wd

20 **Basidio** SP-TFPLNRT FYADFSHDNQ MVAIFSAMGL FNQSAPLDPS -
PDPNRTWVT

351

400

25 *P. involutus* (phyA1) SsLVPPFSGRM VVERLsC..f GT..... tkv
RVLVQDqVQP

P. involutus (phyA2) SsVVPFSARM aVERLsC..a GT..... tkv
RVLVQDqVQP

30 *T. pubescens* kKIVPFSARM VVERLdC..g GA..... qsv
RLLVNDAVQP

A. pediades SRLtPFSARM VtERLLCqrd GTgsgggpsri mrngnvqtfv
RILVNDALQP

P. lycii SKLVPFSGHM tVEKLaC... sgkeav
RVLVNDAVQP

35 **Basidio** SKLVPFSSARM VVERL-C--- GT----- v
RVLVNDAVQP

40

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P. involutus (phyA1) LEFCGGDrNG 1CTLAKFVES QtFARsDGaG DFEKCFATsA ~
P. involutus (phyA2) LEFCGGDqDG 1CALDkFVES QaYARsGGaG DFEKCLATTv ~
T. pubescens LAFCGADtsG vCTLDAFVES QaYARNDGEG DFEKCFAT~~ ~
A. pediades LKFCGGDmDS 1CTLEAFVES QkYAREDGQG DFEKCFD~~~ ~
5 *P. lycii* LEFCGG.vDG vCeLsAFVES QtYARENGQG DFAKCgfvpS e

Basidio LEFCGGD-DG -CTLDAFVES Q-YAREDGQG DFEKCFATP- -

Figure 4

		1
50		
5	<i>A. terreus</i> 9a1 VPeDCHITFV	KhsdCNSVDh GYQCFPELSH KWGLYAPYFS LqDESPFP1D
	<i>A. terreus</i> cbs VPdDCHITFV	NhsdCtSVDr GYQCFPELSH KWGLYAPYFS LqDESPFP1D
10	<i>A. niger</i> var. <i>awamori</i> VPaGCRVTFa	NqsTCDTVdQ GYQCFSEtSH LWGQYAPFFS LANESAISPD
	<i>A. niger</i> NRRL3135 VPaGCRVTFa	NqsSCDTVdQ GYQCFSEtSH LWGQYAPFFS LANESvISPE
	<i>A. fumigatus</i> 13073 LPkDCRITLV	GSkSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK
15	<i>A. fumigatus</i> 32722 LPkDCRITLV	GSkSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK
	<i>A. fumigatus</i> 58128 LPkDCRITLV	GSkSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK
20	<i>A. fumigatus</i> 26906 LPkDCRITLV	GSkSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK
	<i>A. fumigatus</i> 32239 LPkDCRVTfv	GSkACDTVE1 GYQCSPGtSH LWGQYSPFFS LEDE1SVSSD
	<i>E. nidulans</i> VPhGCeVTfv	QNHSCNTaDG GYQCFPNVSH VWGQYSPYFS IEQESAISeD
25	<i>T. thermophilus</i> VPqNCKITFV	DSHSCNTVEG GYQCPEISH sWGQYSPFFS LADQSEISPD
	<i>T. lanuginosa</i> VPkGCRVtFv	----- ~~~~nvDIAR hWGQYSPFFS LAEvSEISPA
30	<i>M. thermophila</i> IPdDCeVTFa	ESRPCDTpD1 GFQCGTAISH FWGQYSPYFS VPsElDaS..
	Basidio pPaGCQIxqV	xSxPxrxxtAA qLPipxQxqx xWSPYSPYFP VAXyxA....
35	Consensus GCRVTfv	NSHSCDTVdG GYQC-PEISH LWGQYSPFFS LADESAISPD VP-
	Fcp10 VPKGCRVTfv	NSHSCDTVdG GYQCFPEISH LWGQYSPFFS LADESAISPD

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100		
5	<i>A. terreus</i> 9a1 QSYNYSLDSE	QVLARHGARS PThSKTKaYA AtIaAIQKSA TaFpGKYAFL
	<i>A. terreus</i> cbs KSYNYSMGSE	QVLARHGARS PTdSKTKaYA AtIaAIQKNA TaLpGKYAFL
	<i>A. niger</i> var. <i>awamori</i> KTYNYSLGAD	QVLSRHGARY PTeSKGKKYS ALIeEIQQNv TtFDGKYAFL
10	<i>A. niger</i> NRRL3135 KTYNYSLGAD	QVLSRHGARY PTdSKGKKYS ALIeEIQQNA TtFDGKYAFL
	<i>A. fumigatus</i> 13073 KTNYNLTGAD	QVLSRHGARY PTSSKSKKYk kLvtAIQaNA TdFKGKFAFL
15	<i>A. fumigatus</i> 32722 KTNYNLTGAD	QVLSRHGARY PTSSKSKKYk kLvtAIQaNA TdFKGKFAFL
	<i>A. fumigatus</i> 58128 KTNYNLTGAD	QVLSRHGARY PTSSKSKKYk kLvtAIQaNA TdFKGKFAFL
	<i>A. fumigatus</i> 26906 KTNYNLTGAD	QVLSRHGARY PTSSKSKKYk kLvtAIQaNA TdFKGKFAFL
20	<i>A. fumigatus</i> 32239 ETNYNLTGAD	QVLSRHGARY PTASKSKKYk kLvtAIQKNA TeFKGKFAFL
	<i>E. nidulans</i> ESNYNLTGAD	QVLSRHGARY PTeSKSKaYS GLIeAIQKNA TsFwGQYAFL
25	<i>T. thermophilus</i> KdYrYqLGAN	QLLSRHGARY PTSSKTELYS qLIsriQkTA TaYKGyYAFL
	<i>T. lanuginosa</i> RdYaYhLGAD	QVLSRHGARY PTAhKSEvYA ELLqrIQDtA TeFKGDFAFL
	<i>M. thermophila</i> RTYDYTLGAD	QVLSRHGARA PTlkRAasYv DLIdrIHhGA isYgPgYEFL
30	<i>Basidio</i> xnxtYxLGxD	NIIqRHGARF PTSGaAtRiq AaVakLQsax xxtDPKLDL
	Consensus KTNYNLTGAD	QVLSRHGARY PTSSKSKKYS ALI-AIQKNA T-FKGKYAFL
35	Fcp10 KTNYNLTGAD	QVLSRHGARY PTSSKSKKYS ALIEAIQKNA TAFKGKYAFL

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5 150

		ELTPFGrNQL rD1GaQFYeR YNAL.TRhIn PFVRATDAsR
	A. terreus 9a1 VhESAEKFVE	
	A. terreus cbs VhESAEKFVE	NLTPFGrNQL qD1GaQFYRR YDTL.TRhIn PFVRAADSSR
10	A. niger var. awamori VIASGEKFIE	DLTPFGEQEL VNSGIKFYQR YESL.TRnII PFIRSSGSSR
	A. niger NRRL3135 VIASGKKFIE	DLTPFGEQEL VNSGIKFYQR YESL.TRnIV PFIRSSGSSR
15	A. fumigatus 13073 VIASGEKFIE	DLTPFGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR
	A. fumigatus 32722 VIASGEKFIE	DLTPFGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR
	A. fumigatus 58128 VIASGEKFIE	DLTPFGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR
20	A. fumigatus 26906 VIASGEKFIE	DLTAFGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR
	A. fumigatus 32239 VIASGEKFIE	DLTPFGEQQM VNSGIKFYQK YKAL.AgsVV PFIRSSGSDR
25	E. nidulans VVASAEKFIN	DLTiFGENQM VDSGaKFYRR YKnL.ARknt PFIRASGSDR
	T. thermophilus VIASGr1FIE	DLTPFGENQM IQLGIKFYnH YKSL.ARnaV PFVRCGSDR
	T. lanuginosa VIASAEffFnr	NLTRFGEQQM MESGrQFYHR YReq.AReIV PFVRAAGSAR
30	M. thermophila VVhSAENFtQ	ELTRtGQQQM VNSGIKFYRR YRAL.ARksI PFVRTAGqDR
	Basidio VVDSAtNwtA	DLvPFGAxQs sQAGqEaFtR YsxEvSxdnL PFVRASGSDR
35	Consensus VIASAEKFIE	DLTPFGEQQM VNSGIKFYRR YKAL-AR-IV PFVRASGSDR
	Fcp10 VIASAEKFIE	DLTPFGEQQM VNSGIKFYRR YKAL.ARkIV PFVRASGSDR

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200

5	<i>A. terreus</i> 9a1 TAFEs...St	GFQTARqDDh hAnphQPSPr VDVaIPEGsA YNNTLEHSLC
	<i>A. terreus</i> cbs TAFEa...St	GFQNARqGDP hAnphQPSPr VDVVIPEGtA YNNTLEHSIC
	<i>A. niger</i> var. <i>awamori</i> TvFEd...SE	GFQSTKLnDP rAqpgQSSPk IDVVISEAss sNNTLDpGtC
10	<i>A. niger</i> NRRL3135 TvFEd...SE	GFQSTKLnDP rAqpgQSSPk IDVVISEAss sNNTLDpGtC
	<i>A. fumigatus</i> 13073 TkFEa...SQ	GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC
15	<i>A. fumigatus</i> 32722 TkFEa...SQ	GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC
	<i>A. fumigatus</i> 58128 TkFEa...SQ	GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC
	<i>A. fumigatus</i> 26906 TkFEa...SQ	GFQqAKLADP gAt.nRAAPa ISVIIPESeT FNNTLDHGVC
20	<i>A. fumigatus</i> 32239 TnFEa...SE	GFQqANVADP gAt.nRAAPV ISVIIPESeT YNNTLDHSVC
	<i>E. nidulans</i> vSFEn...dE	GFRkAQLhDh g.s.gQATPV VNVIIPEidG FNNTLDHStC
25	<i>T. thermophilus</i> PvFEd...Ss	GFQSAKVLDP hSdkhDAPPt INVIIeEGpS YNNTLDtGSC
	<i>T. lanuginosa</i> PAaEe...Ap	GFQdAKdrDP rSnkdQAePV INVIISEEtG sNNTLDgltC
	<i>M. thermophila</i> TAFEegPySt	GFHSALLADR gStvrPTlPy dmVVIPETaG aNNTLHNDLC
30	<i>Basidio</i>PxAG	GFaxA..... .sxntxxPx LxVILSExg. .NDTLDDNMC
	Consensus TAFE--P-SE	GFQSAKLADP -A---QASPV INVIIPEG-G YNNTLDHGLC
35	Fcp10 TAFE...SE	GFQSAKLADP GANPHQASPV INVIIPEGAG YNNTLDHGLC

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	<i>A. terreus</i> 9a1 MCPFETVS1T	VGDDaVANFT AVFAPAIaQR LEAdLPGVQL StDDVVNLMA
	<i>A. terreus</i> cbs MCPFETVS1T	VGDAaADNFT AVFAPAIakR LEAdLPGVQL SADDVVNLMA
5	<i>A. niger</i> var. <i>awamori</i> LADtVEANFT AtFAPSIrqr LEEndLSGVtL TDtEVtyLMD MCSFDTISTS	
	<i>A. niger</i> NRRL3135 MCSFDTISTS	LADtVEANFT AtFvPSIqr LEEndLSGVtL TDtEVtyLMD
10	<i>A. fumigatus</i> 13073 MCSFDTVarT	LGDEVAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD
	<i>A. fumigatus</i> 32722 MCSFDTVarT	LGDEVAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD
	<i>A. fumigatus</i> 58128 MCSFDTVarT	LGDEVAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD
15	<i>A. fumigatus</i> 26906 MCSFDTVarT	LGDEVAANFT ALFAPdIRAR aKkhLPGVtL TDEDVVSLMD
	<i>A. fumigatus</i> 32239 MCSFDTVarT	LGDEVEANFT ALFAPAIRAR IEkhLPGVQL TDDVVSLMD
20	<i>E. nidulans</i> MCSFDTMART	rADEIEANFT AIMGPPIRkR LEEndLPGIKL TNENVIyLMD
	<i>T. thermophilus</i> LCPFETLArn	gGHDaQEKFKA kqFAPAI1EK IKDhLPGVDL AvsDVpyLMD
	<i>T. lanuginosa</i> LCPFDTVGsd	.DptqpAEFl qVFGPRV1kK ItkhMPGVNL T1EDVplFMD
25	<i>M. thermophila</i> LCPFETVass	IGDDaQDtY1 StFAGPItAR VNAaLPGaNL TDADtVaLMD
	<i>Basidio</i> LCPFETVS..	dSDpqxnW1 AVFAPPItAR LNAAaPGaNL TDxDaxNLxx
30	Consensus MCPFDTVA-T	LGDDVEANFT AVFAPPIRAR LEA-LPGVNL TDEDVVNLMD
	Fcp10 MCPFDTVART	LGDDVEANFT AVFAPPIRAR LEAHLPGVNL TDEDVVNLMD
35	300	251
	<i>A. terreus</i> 9a1 dKYYGYGGGN	dD..Aht...LSPF CDLFTa..tE WtQNYL1SL

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	A. terreus cbs	dD..Aht...LSPF CDLFTa..aE WtQYNYL1SL
	dKYYGYGGGN	
	A. niger var. awamori	Tv..DTK...LSPF CDLFTH..dE WiHYDYLQSL
	kKYYGHGAGN	
5	A. niger NRRL3135	Tv..DTK...LSPF CDLFTH..dE WiNYDYLQSL
	kKYYGHGAGN	
	A. fumigatus 13073	SD..ASQ...LSPF CQLFTH..nE WkkYNYLQSL
	gKYYGYGAGN	
10	A. fumigatus 32722	SD..ASQ...LSPF CQLFTH..nE WkkYNYLQSL
	gKYYGYGAGN	
	A. fumigatus 58128	SD..ASQ...LSPF CQLFTH..nE WkkYNYLQSL
	gKYYGYGAGN	
	A. fumigatus 26906	SD..ASQ...LSPF CQLFTH..nE WkkYNYLQSL
	gKYYGYGAGN	
15	A. fumigatus 32239	AD..ASE...LSPF CAIFTH..nE WkKYDYLQSL
	gKYYGYGAGN	
	E. nidulans	AH..GTE...LSPF CAIFTE..KE WlQYDYLQSL
	sKYYGYGAGS	
20	T. thermophilus	ht..DT...LSPF CALsTQ..eE WqaYDYYQSL
	gKYYGnGGGN	
	T. lanuginosa	PvlfPrQ...LSPF CHLFTa..dD WmaYDYYyTL
	dKYYSHGGGS	
	M. thermophila	SsdpATadag ggngrpLSPF CrLFSE..sE WraYDYLQSV
	gKWyGYGPGN	
25	BasidioxexxSxF CDLFexxpeE FxaFxYxgdL
	dKFYGtGyGQ	
	Consensus	SD--ATQ--- -----LSPF CDLFTH---E W-QYDYLQSL -
	KYYGYGAGN	
30	Fcp10	SD..ATQ...LSPF CDLFTH..dE WIQYDYLQSL
	GKYYGYGAGN	
	350	301
	A. terreus 9a1	PLGPvQGVGW aNELMARLTR A.PVHDHTCV NNTLDASPAT
	FPLNATLYAD	
	A. terreus cbs	PLGPvQGVGW aNELIARLTR S.PVHDHTCV NNTLDANPAT
	FPLNATLYAD	

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A. niger var. awamori PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSNPAT
FPLNSTLYAD

A. niger NRRL3135 PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSSPAT
FPLNSTLYAD

5 A. fumigatus 13073 PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT
FPLNATMYvD

A. fumigatus 32722 PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT
FPLNATMYvD

10 A. fumigatus 58128 PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT
FPLNATMYvD

A. fumigatus 26906 PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT
FPLNATMYvD

A. fumigatus 32239 PLGPAQGIGF tNELIARLTN S.PVQDHTST NsTLDSDPAT
FPLNATIYvD

15 E. nidulans PLGPAQGIGF tNELIARLTQ S.PVQDNTST NHTLDSNPAT
FPLDrkLYAD

T. thermophilus PLGPAQGVGF vNELIARMTH S.PVQDYTTv NHTLDSNPAT
FPLNATLYAD

20 T. lanuginosa AFGPSRGVGF vNELIARMTg N1PVKDHTTv NHTLddNPET
FPLDAvLYAD

M. thermophila PLGPTQGVGF vNELLARLA. GvPVRDgTST NRTLDGDPrT
FPLGrPLYAD

Basidio PLGPvQGVGY iNELLARLTx qa.VRDNTqT NRTLDSSPxT
FPLNrTFYAD

25 Consensus PLGPAQGVGF -NELIARLTH S.PVQDHTST NHTLDSNPAT
FPLNATLYAD

Fcp10 PLGPAQGVGF vNELIARLTH S.PVQDHTST NHTLDSNPAT
FPLNATLYAD

30 351

400

A. terreus 9a1 FSHDSnLVSI FWALGLYNGT aPLSqTSVE. .SvsQTDGYA
AAWTVPFAAR

35 A. terreus cbs FSHDSnLVSI FWALGLYNGT kPLSqTTVE. .ditrTDGYA
AAWTVPFAAR

A. niger var. awamori FSHDNGIISI LFALGLYNGT kPLSTTTVE. .NitQTDGFS
SAWTVPFASR

	A. niger NRRL3135 SAWTVPFASR	FSHDNGIISI LFALGLYNGT KPLSTTTVE. .NitQTDGFS
	A. fumigatus 13073 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKE1DGYS
5	A. fumigatus 32722 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT gPLSrTSVE. .SaKE1DGYS
	A. fumigatus 58128 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKE1DGYS
10	A. fumigatus 26906 ASWvVPFGAR	FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKE1DGYS
	A. fumigatus 32239 ASWAVPFGAR	FSHDNGMIPF FFAMGLYNGT ePLSqtSeE. .StKESNGYS
	E. nidulans ASWTVPFGR	FSHDNSMISI FFAMGLYNGT qPLSmDSVE. .SiQEmDGYS
15	T. thermophilus AAWTVPGGR	FSHDNTMtsI FaALGLYNGT aKLSTTeIK. .SIEETDGYS
	T. lanuginosa ASWTVPFAAR	FSHDNTMtGI FsAMGLYNGT kPLSTSkiQP pTgAAADGYA
20	M. thermophila ASWAVPFAAR	FSHDNdMMGV LgALGaYDGv pPLdkTA..R rdpEE1GGYA
	Basidio TSklVPPFSAR	FSHDNqMVAI FsAMGLFNqS aPLdPSxpDP nrt.....Wv
25	ASWTVPFAAR	Consensus FSHDNTMVS1 FFALGLYNGT -PLSTTSVEP -S-EETDGYS
	ASWTVPFAAR	Fcp10 FSHDNTMVS1 FFALGLYNGT KPLSTTSVE. .SIEETDGYS
30	450	401
	A. terreus 9al PLHGCPTDKL	AYVEMMQC.. ra..... EKEPL VRVLVNDRVM
	A. terreus cbs PLHGCADVNL	AYIEMMQC.. ra..... EKQPL VRVLVNDRVM
35	A. niger var. awamori 1YVEMMQC.. Qa..... EQEPL VRVLVNDRVM PLHGCPIDaL	1YVEMMQC.. Qa..... EQEPL VRVLVNDRVM
	A. niger NRRL3135 PLHGCPVDA	1YVEMMQC.. Qa..... EQEPL VRVLVNDRVM

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	<i>A. fumigatus</i> 13073 PLHGCDVDKL	AYfEtMQC.. Ks..... EKEPL VRaLINDRVV
	<i>A. fumigatus</i> 32722 PLHGCDVDKL	AYfEtMQC.. Ks..... EKEPL VRaLINDRVV
5	<i>A. fumigatus</i> 58128 PLHGCDVDKL	AYfEtMQC.. Ks..... EKESL VRaLINDRVV
	<i>A. fumigatus</i> 26906 PLHGCDVDKL	AYfEtMQC.. Ks..... EKEPL VRaLINDRVV
10	<i>A. fumigatus</i> 32239 PLHGCADVDKL	AYfEtMQC.. Ks..... EKEPL VRaLINDRVV
	<i>E. nidulans</i> PLHGCADVDF	AYfELMQC.. E..... KKEPL VRVLVNDRVV
	<i>T. thermophilus</i> PLHGCEVDSL	AYIEMMQC.. Dd..... SDEPV VRVLVNDRVV
15	<i>T. lanuginosa</i> PLHGCrVDRW	AYVELLRC.. Etetsseeee EG... EDEPF VRVLVNDRVV
	<i>M. thermophila</i> TLkGCGaDER	iYVEkMRC.. sgggggggggg EGrqeKDEeM VRVLVNDRVM
20	Basidio PLEfCGgDxd	mvVERLxCxx xgtxxxxxx xxxxooooxx VRVLVNDaVq
	Consensus PLHGCGVDKL	AYVEMMQC-- E----- EG--- EKEPL VRVLVNDRVV
25	Fcp10 PLRGCGVDKL	AYVEMMQC.. EA..... EKEPL VRVLVNDRVV

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		451	482
	<i>A. terreus</i> 9a1	GRCKrDAFVA GLSFAQAG.. GNWADCF--- --	
	<i>A. terreus</i> cbs	GRCKrDDFVE GLSFARAG.. GNWAECF--- --	
	<i>A. niger</i> var. <i>awamori</i>	GRCtrDsFVr GLSFARSG.. GDWAECsA--- --	
5	<i>A. niger</i> NRRL3135	GRCtrDsFVr GLSFARSG.. GDWAECFA--- --	
	<i>A. fumigatus</i> 13073	GRCK1NDFVK GLSWARSG.. GNWGECFS--- --	
	<i>A. fumigatus</i> 32722	GRCK1NDFVK GLSWARSG.. GNWGECFS--- --	
	<i>A. fumigatus</i> 58128	GRCK1NDFVK GLSWARSG.. GNWGECFS--- --	
	<i>A. fumigatus</i> 26906	GRCK1NDFVK GLSWARSG.. GNWGECFS--- --	
10	<i>A. fumigatus</i> 32239	GRCK1KDFVK GLSWARSG.. GNSEQSFS--- --	
	<i>E. nidulans</i>	GRCtlDDWVE GLNFARSG.. GNWKTCT1- --	
	<i>T. thermophilus</i>	GRCKrDDFVr GLSFARqG.. GNWEGCYAas e-	
	<i>T. lanuginosa</i>	GRCRrDEWIK GLTFARqG.. GHWDrCF--- --	
	<i>M. thermophila</i>	GmCtlErFIE SMAFARGN.. GKWDlCFA--- --	
15	Basidio	GxCtlDAFVE SqxYAReDgq GDFEKCFAtp xx	
	Consensus	GRCK-DDFVE GLSFARSG-- GNWEECFAs --	
	Fcp10	GRCKRDDFVE GLSFARSG.. GNWEECFAs ..	

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Figure 5

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5 ECO RI M G V F V V L L S I A T L F G S T 17
TATATGAATTCATGGGCGTGGTCTGCTGCTACTGTCCATTGCCACCTGTTCTGGTTCCA

1 -----+-----+-----+-----+-----+-----+-----+ 60

ATATACCTTAAGTACCCCCACAAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGT

S G T A L G P R G N S H S C D T V D G G 37
10 CATCCGGTACCGCCTGGTCCCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGCTG

GTAGGCCATGGCGGAACCCAGGAGCACCATTAGAGTGAGAACACTGTGACAACTGCCAC

CP-2

CP-3.10

15 Y Q C F P E I S H L W G Q Y S P F F S L 57
 GTTACCAATGTTCCAGAAATTCTCACTTGTGGGTCAATACTCTCCATTCTTCTCTT

121 -----+-----+-----+-----+-----+-----+-----+-----+ 180

CAATGGTTACAAAGGGTCTTAAAGAGTGAACACCCCCAGTTATGAGAGGTAAGAAGAGAA

20 A D E S A I S P D V P K G C R V T E V C 77

TGGCTGACGAATCTGCTATTTCTCCAGACGGTCCAAAGGGTTGTAGACTRACTTCGTTG

ACCGACTGCTTAGACGATAAGAGGTCTGCAAGGTTCCGGACATCTGAACTGAAAGGAGC

CP-4.10 CP-5.10

V L S R H G A R Y P T S S K S K X Y C P

AAGTTTGTCTAGACACGGTGCTAGATAACCGAACCTCTTCTAAGCTGAGCTGCTG

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301 -----+-----+-----+-----+-----+-----+-----+ 360

GAAACTAACCGATAAGTTCTTGCATGACGAAAGTCCCATTCACTGCAGAAAGAACT

CP-6

CP-7.10

5 T Y N Y T L G A D D D L T P F G E Q Q M V 137

AGACTTACAACACTACACTTGGGTGCTGACGACTTGACTCCATTGGTGAACAACAAATGG

361 -----+-----+-----+-----+-----+-----+-----+ 420

TCTGAATGTTGATGTGAAACCCACGACTGCTGAACGTGAGGTAAGCCACTTGTTGTTACC

10 N S G I K F Y R R Y K A L A R K I V P F 157

TTAACCTCTGGTATTAAGTTCTACAGAAGATAACAAGGCTTGGCTAGAAAGATTGTTCCAT

421 -----+-----+-----+-----+-----+-----+-----+ 480

AATTGAGACCATAATTCAAGATGTCTCTATGTTCCGAAACCGATCTTCTAACAGGTA

CP-8.1015 CP-9.10

Y R A S G S D R V I A S A E K F I E G F 177

TCGTTAGAGCTTCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCAATTGAAGGTT 481 -----+-----+-----+-----+-----+-----+-----+ 540

AGCAATCTCGAAGACCAAGACTGTCTCAATAACGAAGACGACTTTCAAGTAACCTCCAA

20

Q S A K L A D P G A N P H Q A S P V I N 197
TCCAATCTGCTAACGTTGGCTGACCCAGGTGCTAACCCACACCAAGCTTCTCCAGTTATTA

541 -----+-----+-----+-----+-----+-----+-----+ 600

AGGTTAGACGATTCAACCGACTGGTCCACGATTGGGTGTTGAGAGGTCAATAAT

25 CP-10.10CP-11.10V I I P E G A G Y N N T L D H G L C T A 217
ACGTTATTATTCCAGAAGGTGCTGGTTACAACACACTTGGACCACGGTTGTGTACTG

601 -----+-----+-----+-----+-----+-----+-----+ 660

30 TGCAATAATAAGGTCTTCCACGACCAATGTTGTTGAAACCTGGTGCACACATGAC

F E E S E L G D D V E A N F T A V F A P 237

CTTTCGAAGAACGTTGAATTGGGTGACGACGTTGAAGCTAACCTCACTGCTGTTTCGCTC

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661 -----+-----+-----+-----+-----+-----+ 720

GAAAGCTTCTTAGACTTAACCCACTGCTGCAACTCGATTGAAGTGACGACAAAAGCGAG

CP-12.10

5 P I R A R L E A H L P G V N L T D E D V 257

CACCTATTAGAGCTAGATTGAAAGCTCACTGCCAGGTGTTAACTTGACTGACGAAGACG

721 -----+-----+-----+-----+-----+-----+ 780

GTGGATAATCTCGATCTAACCTTCGAGTGAACGGTCCACAATTGAACTGACTGCTTCTGC

10 CP-13.10

V N L M D M C P F D T V A R T S D A T Q 277

TTGTTAACCTGATGGACATGTGTCATTGACACTGTTGCTAGAACTTCTGACGCTACTC

781 -----+-----+-----+-----+-----+-----+ 840

AACAAATTGAACTACCTGTACACAGGTAAGCTGTGACAACGATCTGAAGACTGCGATGAG

15 L S P F C D L F T H D E W I Q Y D Y L Q 297

AATTGTCTCCATTCTGTGACTTGTTCACTCACGACGAATGGATTCAATAACGACTACTTC

841 -----+-----+-----+-----+-----+-----+ 900

TTAACAGAGGTAAGACACTGAACAAAGTGAGTGCTGCTTACCTAAGTTATGCTGATGAACG

20 CP-14.10
CP-15.10

S L G K Y Y G Y G A G N P L G P A Q G V 317

AATCTTGGTAAGTACTACGGTTACGGTGTGGTAACCCATTGGTCCAGCTCAAGGTG

901 -----+-----+-----+-----+-----+-----+ 960

25 TTAGAAACCCATTGATGCCATGCCAAATGCCACGACCATTGGTAACCCAGGTCGAGTTCCAC

G F V N E L I A R L T H S P V Q D H T S 337

TTGGTTTGTAAAGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTT

961 -----+-----+-----+-----+-----+-----+
30 1020 AACCAAAGCAATTGCTTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAACP-16.10CP-17.10

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T N H T L D S N P A T F P L N A T L Y A 357

CTACTAACACACTTGGACTCTAACCCAGCTACTTCCCATTGAACGCTACTTTGTACG

1021 -----+-----+-----+-----+-----+-----+
1080

5 GATGATTGGTGTGAAACCTGAGATTGGTGTGATGAAAGGGTAACCTGCGATGAAACATGC

D F S H D N T M V S I F F A L G L Y N G 377

CTGACTTCTCTCACGACAACACTATGGTTCTATTTCCTCGCTTGGTTGTACAACG

1081 -----+-----+-----+-----+-----+-----+
10 1140

GACTGAAGAGAGTGCTGTTGTGATACCAAAGATAAAAGAAGCGAAACCAAACATGTTGC

CP-18.10

CP-19.10

T K P L S T T S V E S I E E T D G Y A A 397

15 GTACTAACGCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAAACTGACGGTTACGCTG

1141 -----+-----+-----+-----+-----+-----+
1200

CATGATTGGTAACAGATGATGAAGACAACCTAGATAACCTCTTGACTGCCAATGCGAC

20 S W T V P F A A R A Y V E M M Q C E A E 417

CTTCTTGGACTGTTCCATTGCGCTGCTAGAGCTTACGTTGAATGATGCAATGTGAAGCTG

1201 -----+-----+-----+-----+-----+-----+
1260

GAAGAACCTGACAAGGTAAAGCCACGATCTGAATGCAACTTACTACGTTACACTTCGAC

25

CP-20.10

CP-21.10

K E P L V R V L V N D R V V P L H G C G 437

AAAAGGAACCATTGGTTAGAGTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG

1261 -----+-----+-----+-----+-----+-----+
30 1320

TTTCCTTGGTAACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAAACGTGCCAACAC

V D K L G R C K R D D F V E G L S F A R 457

GTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTGTCTTCGCTA

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1321 -----+-----+-----+-----+-----+-----+-----+
 1380
 CACAACTGTTCAACCCATCTACATTCTCTGCTGAAGCAACTCCAAACAGAAAGCGAT
 S G G N W E E C F A * Eco RI 467
CP-22.10
 GATCTGGTGGTAACTGGGAAGAACATGTTCGCTTAAGAATTCAATA
 1381 -----+-----+-----+-----+-----+-----+-----+ 1426
 CTAGACCACCATGACCCCTCTTACAAAGCGAATTCTTAAGTATAT

10

Figure 6

		1
50		
15	<i>P. involutus</i> (phyA1) pPaGCQInqV	----- -FPipeseqR nWSPYSPYFP LAEyKA....
	<i>P. involutus</i> (phyA2) pPaGCEInqV	----- -FsipeseqR nWSPYSPYFP LAEyKA....
20	<i>T. pubescens</i> pPaSCQInqV	----- -LDvtRDVqQ sWSmYSPYFP aAtyvA....
	<i>A. pediades</i> pPKDCKITqV	----- -pffpPQIqD sWAaYTPYYP VqAyTP....
	<i>P. lycii</i> pPEGCtVTqV	----- -LPipAQnTs nWGPYdPFFP VEpyAA....
25	<i>A. terreus</i> 9a1 VPEDCHITFV	KhsdCNSVDh GYQCfPELSh kWG1YAPYFS LqDESPFPlD
	<i>A. terreus</i> cbs VPDDCHITFV	NhsdCtSVDr GYQCfPELSh kWG1YAPYFS LqDESPFPlD
30	<i>A. niger</i> var. <i>awamori</i> VPaGCRVTFa	NqsSCDTVDq GYQCfSEtSH LWGQYAPFFS LANESAISPD
	<i>A. niger</i> T213 VPaGCRVTFa	NqsSCDTVDq GYQCfSEtSH LWGQYAPFFS LANESvISPD
	<i>A. niger</i> NRRL3135 VPaGCRVTFa	NqsSCDTVDq GYQCfSEtSH LWGQYAPFFS LANESvISPE
35	<i>A. fumigatus</i> ATCC13073 LPKDCRITLV	GSkSCDTVD1 GYQC8PAtSH LWGQYSPFFS LEDE1SVSSK

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	<i>A. fumigatus</i> ATCC32722	GSkSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK LPKDCRITLV
	<i>A. fumigatus</i> ATCC58128	GSkSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK LPKDCRITLV
5	<i>A. fumigatus</i> ATCC26906	GSkSCDTVd1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK LPKDCRITLV
	<i>A. fumigatus</i> ATCC32239	GSkACDTVd1 GYQCSPGtSH LWGQYSPFFS LEDE1SVSSD LPKDCRVTfV
10	<i>E. nidulans</i> VPhGCeVTfV	QNHSCNTaDg GYQCfPNVSH VWGQYSPYFS IEQESAISeD
	<i>T. thermophilus</i> VPQNCKITfV	DSHSCNTVEg GYQCPEISH sWGQYSPFFS LADQSEISPD
	<i>T. lanuginosa</i> VPKGCRVeFV	----- ----nvDIAR hWGQYSPFFS LAEvSEISPA
15	<i>M. thermophila</i> IPDDCeVTfA	ESRPCDTpD1 GFQCgTAISH FWGQYSPYFS VPSe1DaS..
 Consensus Seq. 11 VPKGCRVTfV		NSHSCDTVd- GYQC-PEISH LWGQYSPFFS LADESAISPD
20		
100		51
	<i>P. involutus</i> (phyA1) KSFKYdLGns	NIIqRHGARF PTSGaTtRik AgLtKLQgvq nftDAKFnFI
25	<i>P. involutus</i> (phyA2) KSFTYdLGTs	NIIqRHGARF PTSGaAtRik AgLsKLQsvq nftDPKFDFI
	<i>T. pubescens</i> tnYtYSLGqD	HIIqRHGARF PTSGaAKRiq TaVAKLkaaS nytdp1LAFV
30	<i>A. pediades</i> tnYtYTLGhD	NIIqRHGARF PTSGaGtRiq AaVKKLQsak TytdPRLDFL
	<i>P. lycii</i> NdFvYkFGvA	NLIqRHGARW PTSGarsRqv AaVAKIQmar PftDPKYEFL
	<i>A. terreus</i> 9a1 QSYNYSLDSE	QVLARHGARs PThSKTKaYA AtIAaIQKSA TaFpGKYAFL
35	<i>A. terreus</i> cbs KSYNYSMGSE	QVLARHGARs PTdSKTKaYA AtIAaIQKNA TaLpGKYAFL
	<i>A. niger</i> var. <i>awamori</i> KTYNYSLGAD	QVLSRHGARY PTeSKGKKYS ALIEeIQQNv TtFDGKYAFL

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	A. niger T213 KTYNYSLGAD	QVLSRHGARY PTeSKGKKYS ALIEeIQQNv TtFDGKYAFL
	A. niger NRRL3135 KTYNYSLGAD	QVLSRHGARY PTdSKGKKYS ALIEeIQQNA TtFDGKYAFL
5	A. fumigatus ATCC13073 KTNYNLTGAD	QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL
	A. fumigatus ATCC32722 KTNYNLTGAD	QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL
10	A. fumigatus ATCC58128 KTNYNLTGAD	QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL
	A. fumigatus ATCC26906 KTNYNLTGAD	QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL
	A. fumigatus ATCC32239 ETNYNLTGAD	QVLSRHGARY PTASKSKKYk kLVtaIQKNA TeFKGKFAFL
15	E. nidulans ESNYNLTGAD	QVLSRHGARY PTeSKSKaYS GLIEaIQKNA TsFwgQYAF
	T. thermophilus KdYrYqLGAN	QLLSRHGARY PTSSKTELYS qLIIsRIQKtA TaYKGyYAF
20	T. lanuginosa RdYaYhLGAD	QVLSRHGARY PTAhKSEvYA ELLQRIQDtA TeFKGDFAF
	M. thermophila RTYDYTLGAD	QVLSRHGARa PT1kRAasYv DLIDRIHhGA isYgPgYEFL
25	Consensus Seq. 11 KTNYNLTGAD	QVLSRHGARY PTSSKSKKYS ALIERIQKNA T-FKGKYAFL

101

150

30	P. involutus (phyA1) VVDSAtNWtA	DLvPFGAAQs fDAGqEaFaR YskLvSKNnL PFIRAdGSDR
	P. involutus (phyA2) VVDTAtNWtA	DLvPFGAAQs fDAGLEvFaR YskLvSsDnL PFIRSDGSDR
	T. pubescens VVATANNWtA	sLveLGAtQs sEAGqEaFtR YsSLvSaDeL PFVRASGSDR
35	A. pediades VVDSAtNWtE	DLvPFGAlQs sQAGeEtFQR YsfLvSKEnL PFVRASSSNR
	P. lycii VVDSStNWtA	DL1PFGANQs hQTGtDMYtR YsTlFEGGdV PFVRAAGdQR

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	<i>A. terreus</i> 9a1 VhESAEKFVE	ELTPFGrNQL rDlGaQFYeR YNAL.TRHIn PFVRATDAsR
	<i>A. terreus</i> cbs VhESAEKFVE	NLTPFGrNQL qDlGaQFYRR YDTL.TRHIn PFVRAADSSR
5	<i>A. niger</i> var. <i>awamori</i> VIASGEKFIE	DLTPFGEQEL VNSGIKFYQR YESL.TRNII PFIRSSGSSR
	<i>A. niger</i> T213 VIASGEKFIE	DLTPFGEQEL VNSGIKFYQR YESL.TRNII PFIRSSGSSR
10	<i>A. niger</i> NRRL3135 VIASGKKFIE	DLTPFGEQEL VNSGIKFYQR YESL.TRNIV PFIRSSGSSR
	<i>A. fumigatus</i> ATCC13073 VIASGEKFIE	DLTPFGEQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR
	<i>A. fumigatus</i> ATCC32722 VIASGEKFIE	DLTPFGEQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR
15	<i>A. fumigatus</i> ATCC58128 VIASGEKFIE	DLTPFGEQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR
	<i>A. fumigatus</i> ATCC26906 VIASGEKFIE	DLTAFGEQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR
20	<i>A. fumigatus</i> ATCC32239 VIASGEKFIE	DLTPFGEQQM VNSGIKFYQK YKAL.AgSVV PFIRSSGSDR
	<i>E. nidulans</i> VVASAEKFIN	DLTiFGENQM VDSGaKFYRR YKnL.ARKnT PFIRASGSDR
	<i>T. thermophilus</i> VIASGr1FIE	DLTPFGENQM IQLGIKFYnH YKSL.ARNaV PFVRCSGSDR
25	<i>T. lanuginosa</i> VIASAEfFnr	NLTRFGEEQM MESGrQFYHR YREq.AREIV PFVRAAGSAR
	<i>M. thermophila</i> VVhSAENFTQ	ELTRtGQQQM VNSGIKFYRR YRAL.ARksI PFVRTAGqDR
30	<i>Consensus Seq. 11</i> VIASAEKFIE	DLTPFGENQM VNSGIKFYRR YKAL-ARNIV PFVRASGSDR

151

200

35	<i>P. involutus</i> (phyA1) PAaGD.....	GFaSA..... .shNtvqPk LNLILPQ..T gNDTLEDNMC
	<i>P. involutus</i> (phyA2) PAaGE.....	GFaSA..... .srNaiqPk LDLILPQ..T gNDTLEDNMC

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	<i>T. pubescens</i>	GFaIa..... .ssNsItpV LSVIISE..A gNDTLDDNMC PAaGD.....
	<i>A. pediades</i>	GFsAA..... .shHvlNPI LfVILSE..S LNDTLDDAMC PnaGs.....
5	<i>P. lycii</i>	GFgdA..... .sgEtvlPt LQVVLQE..E gNcTLCNNMC PnevD.....
	<i>A. terreus</i> 9a1	GFQTARqDDh hAnpHQPSPr VDVaIPEGSA YNNTLEHSLC TAFEs...ST
10	<i>A. terreus</i> cbs	GFQNARqGDP hAnpHQPSPr VDVVIPEGTA YNNTLEHSIC TAFEA...ST
	<i>A. niger</i> var. <i>awamori</i>	GFQSTKLkDP rAqpgQSSPk IDVVISEASS sNNTLDpGtC TvFED...Se
	<i>A. niger</i> T213	GFQSTKLkDP rAqpgQSSPk IDVVISEASS sNNTLDpGtC TvFED...Se
15	<i>A. niger</i> NRRL3135	GFQSTKLkDP rAqpgQSSPk IDVVISEASS sNNTLDpGtC TvFED...Se
	<i>A. fumigatus</i> ATCC13073	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC TkFEA...Sq
20	<i>A. fumigatus</i> ATCC32722	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC TkFEA...Sq
	<i>A. fumigatus</i> ATCC58128	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC TkFEA...Sq
	<i>A. fumigatus</i> ATCC26906	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC TkFEA...Sq
25	<i>A. fumigatus</i> ATCC32239	GFQqANVADP gAt.NRAAPV ISVIIPESeT YNNTLDHSVC TnFEA...Se
	<i>E. nidulans</i>	GFRKAQLhDh g.s.gQATPV VNVIPISEt FNNTLDHStC vSFEN...de
30	<i>T. thermophilus</i>	GFQSAKVLDP hSdKHDAPPt INVIIeEGPS YNNTLDtGSC PvFED...SS
	<i>T. lanuginosa</i>	GFQdAKdrDP rSnkDQAEPV INVIISEETG sNNTLDgltC PAaEE...AP
	<i>M. thermophila</i>	GFHSALLADR gStvRPTlPy dmVVIPETAG aNNTLHNDLC TAFEEgpyST
35	<i>Consensus Seq. 11</i>	GFQSAKLDAP -A--HQASPV INVIIPEGSG YNNTLDHGLC TAFED---ST

250

	<i>P. involutus</i> (phyA1)	.SDpqvnaW1 AVafPSItAR LNAAaPSVNL TDtDafNLVs LCAF1TVSK.
5	<i>P. involutus</i> (phyA2)	.SDpqvDaW1 AsafPSVtAQ LNAAaPGaNL TDADafNLVs LCPFmTVSK.
	<i>T. pubescens</i>	.SDpqvnQW1 AqFAPPMtAR LNAGaPGaNL TDtDtyNLLt LCPFETVAt.
10	<i>A. pediades</i>	.SDpqtGiWT SIYGTPiAnR LNqqaPGaNI TAADVsNLIP LCAFETIVK.
	<i>P. lycii</i>	.GDEST.tW1 GVFAPnItAR LNAAaPSaNL SDsDaLtLMD MCPFDLSS.
	<i>A. terreus</i> 9a1	VGDDAvANFT AVFAPAIaqR LEAdLPGVQL StDDVVNLMA MCPFETVS1T
15	<i>A. terreus</i> cbs	VGDAADNFT AVFAPAIakR LEAdLPGVQL SADDVVNLMA MCPFETVS1T
	<i>A. niger</i> var. <i>awamori</i>	LADtvEANFT AtFAPSIRqR LEndLSGVtL TDtEVtyLMD MCSFDTIStS
20	<i>A. niger</i> T213	LADtvEANFT AtFAPSIRqR LEndLSGVtL TDtEVtyLMD MCSFDTIStS
	<i>A. niger</i> NRRL3135	LADtvEANFT AtFvPSIRqR LEndLSGVtL TDtEVtyLMD MCSFDTIStS
	<i>A. fumigatus</i> ATCC13073	LGDEvAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD MCSFDTVART
25	<i>A. fumigatus</i> ATCC32722	LGDEvAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD MCSFDTVART
	<i>A. fumigatus</i> ATCC58128	LGDEvAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD MCSFDTVART
30	<i>A. fumigatus</i> ATCC26906	LGDEvAANFT ALFAPdIRAR aKkhLPGVtL TDEDVVSLMD MCSFDTVART
	<i>A. fumigatus</i> ATCC32239	LGDEvEANFT ALFAPAIRAR IEkhLPGVQL TDDVVSLMD MCSFDTVART
	<i>E. nidulans</i>	rADEiEANFT AIMGPPIRKR LEndLPGIKL TNENVIyLMD MCSFDTMART
35	<i>T. thermophilus</i>	gGHDAQEKFKA kqFAPAIKEK IKDhLPGVDL AvsDVpyLMD LCPFETLARn
	<i>T. lanuginosa</i>	.DptqpAEF1 qVFGPRVlkK ItkhMPGVNL TLEDVp1FMD LCPFDTVGsd

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M. thermophila
LCPFETVASS

IGDDAQDTYL StFAGPItAR VNAnLPGaNL TDADtVaLMD

5 *Consensus Seq. 11* LGDDAEANFT AVFAPPiRAR LEA-LPGVNL TDEDVVNLMD
MCPFDtvART

	251
	300
10	<i>P. involutus (phyA1)</i>ekkSdF CtLFegiPGs FeaFAYggdL dKFYGtGyGQ
	<i>P. involutus (phyA2)</i>eqkSdF CtLFegiPGs FeaFAYagdL dKFYGtGyGQ
	<i>T. pubescens</i>errSeF CDIYeelqAE .daFAYnadL dKFYGtGyGQ
15	<i>A. pediades</i>etpSPF CNLF..TPEE FaQFEYFgdL dKFYGtGyGQ
	<i>P. lycii</i>gnaSPF CDLF..TAAE YvsYEYYydl dKYYGtGPGN
20	<i>A. terreus</i> 9a1dD..Aht... LSPF CDLF..TAtE WtQNYLLSL dKYYGYGGGN
	<i>A. terreus</i> cbsdD..Aht... LSPF CDLF..TAAE WtQNYLLSL dKYYGYGGGN
	<i>A. niger</i> var. <i>awamori</i> Tv..DTK... LSPF CDLF..ThDE WiHYDYLQSL KKYYGHGAGN
25	<i>A. niger</i> T213Tv..DTK... LSPF CDLF..ThDE WiHYDYLRSL KKYYGHGAGN
	<i>A. niger</i> NRRL3135Tv..DTK... LSPF CDLF..ThDE WiNYDYLQSL KKYYGHGAGN
30	<i>A. fumigatus</i> ATCC13073 SD..ASQ... LSPF CQLF..ThNE WkKYNYLQSL gKYYGYGAGN
	<i>A. fumigatus</i> ATCC32722 SD..ASQ... LSPF CQLF..ThNE WkKYNYLQSL gKYYGYGAGN
	<i>A. fumigatus</i> ATCC58128 SD..ASQ... LSPF CQLF..ThNE WkKYNYLQSL gKYYGYGAGN
35	<i>A. fumigatus</i> ATCC26906 SD..ASQ... LSPF CQLF..ThNE WkKYNYLQSL gKYYGYGAGN
	<i>A. fumigatus</i> ATCC32239 AD..ASE... LSPF CAIF..ThNE WkKYDYLQSL gKYYGYGAGN

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	<i>E. nidulans</i>	AH..GTE... LSPF CAIF..TEKE W1QYDYLQSL SKYYGYGAGS
	<i>T. thermophilus</i>	ht..DT... LSPF CALs..TqEE WqaYDYYQSL gKYYGnGGGN
5	<i>T. lanuginosa</i>	PvlfPrQ... LSPF CHLF..TADD WmaYDYYyTL dKYYSHGGGS
	<i>M. thermophila</i>	SsdpATadag ggnggrpLSPF CrLF..SEsE WraYDYLQSV gKWyGYGPGN
10	Consensus Seq. 11	SD--ATQ--- -----LSPF CDLF--TADE W-QYDYLQSL - KYYGYGAGN

301

350

15	<i>P. involutus</i> (phyA1)	eLGPvQGVGY vNELIARLTN S.AVRDNTqT NRTLDASPvT FPLNkTFYAD
	<i>P. involutus</i> (phyA2)	ALGPvQGVGY iNELLARLTN S.AVNDNTqT NRTLDAApDT FPLNkTMYAD
20	<i>T. pubescens</i>	PLGPvQGVGY iNELIARLTa q.nVsDHTqT NsTLDSSPET FPLNrTLYAD
	<i>A. pediades</i>	PLGPvQGVGY iNELLARLTm .PVRDNTqT NRTLDSSPlT FPLDrSIYAD
	<i>P. lycii</i>	ALGPvQGVGY vNELLARLTg q.AVRDETqT NRTLDSDPAT FPLNrTFYAD
25	<i>A. terreus</i> 9a1	PLGPvQGVGW aNELMARLTR A.PVHDHTCv NNTLDASPAT FPLNATLYAD
	<i>A. terreus</i> cbs	PLGPvQGVGW aNELIARLTR S.PVHDHTCv NNTLDANPAT FPLNATLYAD
30	<i>A. niger</i> var. <i>awamori</i>	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSNPAT FPLNSTLYAD
	<i>A. niger</i> T213	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSNPAT FPLNSTLYAD
	<i>A. niger</i> NRRL3135	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSSPAT FPLNSTLYAD
35	<i>A. fumigatus</i> ATCC13073	PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTlvSNPAT FPLNATMYvD
	<i>A. fumigatus</i> ATCC32722	PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTlvSNPAT FPLNATMYvD

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	<i>A. fumigatus</i> ATCC58128	PLGPAQQGIGF tNELIARLTR S.PVQDHST NsTLvSNPAT FPLNATMYvD
	<i>A. fumigatus</i> ATCC26906	PLGPAQQGIGF tNELIARLTR S.PVQDHST NsTLvSNPAT FPLNATMYvD
5	<i>A. fumigatus</i> ATCC32239	PLGPAQQGIGF tNELIARLTN S.PVQDHST NsTLDSDPAT FPLNATIYvD
	<i>E. nidulans</i>	PLGPAQQGIGF tNELIARLTQ S.PVQDNTST NHTLDSNPAT FPLDrkLYAD
10	<i>T. thermophilus</i>	PLGPAQGVGF vNELIARMTH S.PVQDYTTv NHTLDSNPAT FPLNATLYAD
	<i>T. lanuginosa</i>	AFGPSRGVGF vNELIARMTg N1PVKDHTTv NHTLDDNPET FPLDAvLYAD
	<i>M. thermophila</i>	PLGPTQGVGF vNELLARLA. GvPVRDgtST NRTLDGDPrT FPLGrPLYAD
15		
	Consensus Seq. 11	PLGPAQGVGF -NELIARLT S-PVQDHST NHTLDSNPAT FPLNATLYAD
		351
20	400	
	<i>P. involutus</i> (phyA1)	FSHDN1MVAV FsAMGLFrqP aPLSTSvpNP wrt.....Wr TSSLVPPFSGR
	<i>P. involutus</i> (phyA2)	FSHDN1MVAV FsAMGLFrqS aPLSTSTpDP nrt.....wl TSSvVPPFSAR
25	<i>T. pubescens</i>	FSHDNqMVAI FsAMGLFNqS aPLdPTTpDP art.....Fl vKKiVPPFSAR
	<i>A. pediades</i>	LSHDNqMIAI FsAMGLFNqS sPLdPSfpNP krt.....Wv TSRltPFSAR
30	<i>P. lycii</i>	FSHDNTMVPI FaALGLFNAT a.LdPlkpDe nrl.....Wv DSk1VPPFSGH
	<i>A. terreus</i> 9a1	FSHDSnLVSI FWALGLYNGT aPLSqtSVES Vs..QTDGYA AAWTVPFAAR
	<i>A. terreus</i> cbs	FSHDSnLVSI FWALGLYNGT KPLSqtTTVED It..rTDGYA AAWTVPFAAR
35	<i>A. niger</i> var. <i>awamori</i>	FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS SAWTVPFASR
	<i>A. niger</i> T213	FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS SAWTVPFASR

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	<i>A. niger</i> NRRL3135	FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS SAWTVPFASR
	<i>A. fumigatus</i> ATCC13073	FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..E1DGYS ASWvVPFGAR
5	<i>A. fumigatus</i> ATCC32722	FSHDNSMVSI FFALGLYNGT gPLSrTSVES ak..E1DGYS ASWvVPFGAR
	<i>A. fumigatus</i> ATCC58128	FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..E1DGYS ASWvVPFGAR
10	<i>A. fumigatus</i> ATCC26906	FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..E1DGYS ASWvVPFGAR
	<i>A. fumigatus</i> ATCC32239	FSHDNGMIPF FFAMGLYNGT EPLSqtSeES tk..ESNGYS ASWAVPFGAR
	<i>E. nidulans</i>	FSHDNSMISI FFAMGLYNGT QPLSmdSVES Iq..EmDGYA ASWTVPFAR
15	<i>T. thermophilus</i>	FSHDNTMtsI FaALGLYNGT akLSTTeIKS Ie..ETDGYS AAWTVPFGR
	<i>T. lanuginosa</i>	FSHDNTMtsGI FsAMGLYNGT KPLSTSkiQP ptgaAADGYA ASWTVPFAR
20	<i>M. thermophila</i>	FSHDNDMMGV LgALGaYDGv pPLdkTArrd ..peElGGYA ASWAVPFAAR
	Consensus Seq. 11	FSHDNTMVSI FFALGLYNGT KPLSTTSVES I---ETDGYS ASWTVPFAR
25		401
	450	
	<i>P. involutus</i> (phyA1)	mvVERLsC.. fGt..... Tk VRVLVQDQVq PLEfCGgDRn
	<i>P. involutus</i> (phyA2)	maVERLsC.. AGt..... Tk VRVLVQDQVq PLEfCGgDQd
30	<i>T. pubescens</i>	mvVERLDC.. GGa..... Qs VRLLVNDaVq PLafCGaDts
	<i>A. pediades</i>	mvTERLlCQr DGtGSGGpsr imrNgnvQTF VRILVNDaLq PLkfCGgDmd
35	<i>P. lycii</i>	mtVEkLaC.. sgKea VRVLVNDaVq PLEfCGg.vd
	<i>A. terreus</i> 9al	AYVEMMQCrA EK...EPL VRVLVNDRVM PLHGCptDKL

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	<i>A. terreus</i> cbs	AYIEMMQCrAEK...QPL VRVLVNDRVM PLHCAVDNL
	<i>A. niger</i> var. <i>awamori</i>	LYVEMMQCQAEQ...EPL VRVLVNDRVM PLHGCPIDaL
5	<i>A. niger</i> T213	LYVEMMQCQAEQ...EPL VRVLVNDRVM PLHGCPIDaL
	<i>A. niger</i> NRRL3135	LYVEMMQCQAEQ...EPL VRVLVNDRVM PLHGCPVDAaL
10	<i>A. fumigatus</i> ATCC13073	AYfEtMQCKSEK...EPL VRaLINDRVM PLHGCDVDKL
	<i>A. fumigatus</i> ATCC32722	AYfEtMQCKSEK...EPL VRaLINDRVM PLHGCDVDKL
	<i>A. fumigatus</i> ATCC58128	AYfEtMQCKSEK...ESL VRaLINDRVM PLHGCDVDKL
15	<i>A. fumigatus</i> ATCC26906	AYfEtMQCKSEK...EPL VRaLINDRVM PLHGCDVDKL
	<i>A. fumigatus</i> ATCC32239	AYfEtMQCKSEK...EPL VRaLINDRVM PLHCAVDKL
20	<i>E. nidulans</i>	AYfELMQCE.KK...EPL VRVLVNDRVM PLHGCAVDF
	<i>T. thermophilus</i>	AYIEMMQCDDsD...EPV VRVLVNDRVM PLHGCEVDSL
	<i>T. lanuginosa</i>	AYVELLRCET ETsSeEEeEG ..ED...EPF VRVLVNDRVM PLHGCrVDRW
25	<i>M. thermophila</i>	iYVEkMRCsG GGgGgGGgEG ..rQekdEeM VRVLVNDRVM TLkGCGaDER
	Consensus Seq. 11	AYVEMMQCEA GG-G-GG-EG --EK---EPL VRVLVNDRVM PLHGCGVDKL
30		
		451
	<i>P. involutus</i> (phyA1)	G1CtLAKFVE SqTFARSDga GDFEKCFAts a-
	<i>P. involutus</i> (phyA2)	G1CaLDKFVE SqAYARSGga GDFEKCLAtt v-
	<i>T. pubescens</i>	GvCtLDAFVE SqAYARNDge GDFEKCFAt- --
35	<i>A. pediades</i>	S1CtLEAFVE SqkYAReDgq GDFEKCFD-- --
	<i>P. lycii</i>	GvCELsAFVE SqTYAReNgq GDFAKCgfvp se

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	<i>A. terreus</i> 9a1	GRCKrDAFVA GLSFAQAG.. GNWADCF--- --
	<i>A. terreus</i> cbs	GRCKrDDFVE GLSFARAG.. GNWAECF--- --
	<i>A. niger</i> var. <i>awamori</i>	GRCtrDsFVr GLSFARSG.. GDWAECsA-- --
	<i>A. niger</i> T213	GRCtrDsFVr GLSFARSG.. GDWAECFA-- --
5	<i>A. niger</i> NRRL3135	GRCtrDsFVr GLSFARSG.. GDWAECFA-- --
	<i>A. fumigatus</i> ATCC13073	GRCKLNDFK GLSWARSG.. GNWGECFS-- --
	<i>A. fumigatus</i> ATCC32722	GRCKLNDFK GLSWARSG.. GNWGECFS-- --
	<i>A. fumigatus</i> ATCC58128	GRCKLNDFK GLSWARSG.. GNWGECFS-- --
	<i>A. fumigatus</i> ATCC26906	GRCKLNDFK GLSWARSG.. GNWGECFS-- --
10	<i>A. fumigatus</i> ATCC32239	GRCKLKDFVK GLSWARSG.. GNSEQSFS-- --
	<i>E. nidulans</i>	GRCtlLDDWVE GLNFARSG.. GNWktCFT1- --
	<i>T. thermophilus</i>	GRCKrDDFVr GLSFARqG.. GNWEGCYAas e~
	<i>T. lanuginosa</i>	GRCRrDEWIK GLTFARqG.. GHWDrcF~~~ --
	<i>M. thermophila</i>	GmCtLErFIE SMAFARGN.. GKWD1CFA-- --
15		
	Consensus Seq. 11	GRCKLDDFVE GLSFARSG-- GNWAECFA-- --

20

25

Figure 7

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- 90 -

20 M G V F V V L L S I A T L F G S T S G T
 ATGGGCGTGGTCTCGTCGTGCTACTGTCCATTGCCACCTGGTTCCACATCCGGTACC
 5 1 -----+-----+-----+-----+-----+-----+-----+-----+
 60 TACCCGCACAAGCAGCAGATGACAGGTAACGGTGGAACAGCAAGGTGTAGGCCATGG
 10 40 A L G P R G N S H S C D T V D G G Y Q C
 GCCTTGGGTCTCGTGGTAATTCTCACTCTTGACACTGTTGACGGTGGTACCAATGT
 120 61 -----+-----+-----+-----+-----+-----+-----+-----+
 CGGAACCCAGGAGCACCATTAAAGAGTGAGAACACTGTGACAACTGCCACCAATGGTTACA
 15 15 F P E I S H L W G T Y S P Y F S L A D E
 60 TTCCCAGAAATTCTCACTTGTTGGGTACCTACTCTCCATACTTCTCTTGGCAGACGAA
 20 180 121 -----+-----+-----+-----+-----+-----+-----+
 AAGGGTCTTAAAGAGTGAAACACCCATGGATGAGAGGTATGAAGAGAAACCGTCTGCTT
 80 S A I S P D V P D D C R V T F V Q V L S
 25 TCTGCTATTCAGACGTTCCAGACGACTGTAGAGTTACTTCGTTCAAGTTTGCT
 240 187 -----+-----+-----+-----+-----+-----+-----+
 AGACGATAAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAGTTCAAAACAGA
 30 100 R H G A R Y P T S S A S K A Y S A L I E
 AGACACGGTGCTAGATAACCAACTTCTCTCGGTCTAAGGCTTACTCTGCTTTGATTGAA
 241 241 -----+-----+-----+-----+-----+-----+-----+
 300 35 TCTGTGCCACGATCTATGGGTTGAAGAAGACGCAGATTCCGAATGAGACGAAACTAATT
 120 A I Q K N A T A F K G K Y A F L K T Y N

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GCTATTCAAAAGAACGCTACTGCTTCAGGGTAAGTACGCTTCTGAAGACTTACAAC
 301 -----+-----+-----+-----+-----+-----+-----+-----+
 360

CGATAAGTTTCTTGCATGACGAAAGTCCCATTGCGAAAGAACTTCTGAATGTTG
 5

Y T L G A D D L T P F G E N Q M V N S G
 140

TACACTTTGGGTGCTGACGACTTGACTCCATTGGTGAACCAATGGTTAATCTGGT
 10 361 -----+-----+-----+-----+-----+-----+-----+-----+
 420

ATGTGAAACCCACGACTGCTGAAC TGAGGTAAAGCCACTTTGGTTACCAATTGAGACCA
 160

I K F Y R R Y K A L A R K I V P F I R A
 15

ATTAAGTTCTACAGAAGATAACAAGGCTTGGCTAGAAAGATTGTTCCATTAGAGCT
 421 -----+-----+-----+-----+-----+-----+-----+-----+
 480

TAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTCTAACAGGTAAAGTAATCTCGA
 20

S G S D R V I A S A E K F I E G F Q S A
 180

TCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAGTTCAATTGAAGGTTCCAATCTGCT
 481 -----+-----+-----+-----+-----+-----+-----+-----+
 540

AGACCAAGACTGTCTCAATAACGAAGACGACTTTCAAGTAACCTCCAAAGGTTAGACGA
 25

K L A D P G S Q P H Q A S P V I N V I I
 200

AAGTTGGCTGACCCAGGTTCTAACCAACACACCAAGCTTCTCCAGTTATTACGTGATCATT
 30 541 -----+-----+-----+-----+-----+-----+-----+-----+
 600

TTCAACCGACTGGTCCAAGAGTTGGTGTGGTCAAGAGGTCAATAATTGCACTAGTAA
 35 220

P E G S G Y N N T L D H G T C T A F E D
 CCAGAAGGATCCGGTTACAACACACTTGGACCACGGTACTGTACTGCTTCGAAGAC

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601 -----+-----+-----+-----+-----+-----+
660
GGTCTTCCTAGGCCAATGTTGTTGAAACCTGGTGCATGAACATGACGAAAGCTCTG
5 S E L G D D V E A N F T A L F A P A I R
240
TCTGAATTAGGTGACGACGTTGAAGCTAACCTCACTGCTTGTTGCTCCAGCTATTAGA
661 -----+-----+-----+-----+-----+-----+
720
10 AGACTTAATCCACTGCTGCAACTTCGATTGAAGTGACGAAACAAGCGAGGTCGATAATCT
A R L E A D L P G V T L T D E D V V Y L
260
GCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTGACTGACGAAGACGTTGTTACTTG
15 721 -----+-----+-----+-----+-----+-----+
780
CGATCTAACCTCGACTGAACGGTCCACAATGAAACTGACTGCTTCTGCAACAAATGAAC
M D M C P F D T V A R T S D A T E L S P
20 280
ATGGACATGTGTCCATTGACACTGTCGCTAGAACTTCTGACGCTACTGAATTGTCTCCA
781 -----+-----+-----+-----+-----+-----+
840
TACCTGTACACAGGTAAGCTGTGACAGCGATCTGAAGACTGCGATGACTTAACAGAGGT
25 F C A L F T H D E W I Q Y D Y L Q S L G
300
TTCTGTGCTTGTTCACTCACGACGAATGGATCCAATACGACTACTGCAAAGCTTGGGT
30 900
AAGACACGAAACAAGTGAGTGCTGCTTACCTAGGTTATGCTGATGAACGTTCGAACCCA
K Y Y G Y G A G N P L G P A Q G V G F A
320
35 AAGTACTACGGTTACGGTGTGGTAACCCATTGGGTCCAGCTCAAGGTGTTGGTTCGCT
901 -----+-----+-----+-----+-----+-----+
960

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TTCATGATGCCAATGCCACGACCATTGGTAACCCAGGTCGAGTCCACAACCAAAGCGA

N E L I A R L T H S P V Q D H T S T N H
340

5 AACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTTCTACTAACAC
961 ---+-----+-----+-----+-----+-----+
1020

TTGCTTAACGATCTAACGACTGAGTGAGAGGTCAAGTTCTGGTGTGAAGATGATTGGTG

10 T L D S N P A T F P L N A T L Y A D F S
360

ACTTTGGACTCTAACCCAGCTACTTTCCCATTGAACGCTACTTGTACGCTGACTTCTCT
1021 ---+-----+-----+-----+-----+-----+
1080

15 TGAAACCTGAGATTGGTCGATGAAAGGTAACTTGCATGAAACATGCGACTGAAGAGA
380 H D N T M I S I F F A L G L Y N G T K P
CACGACAAACACTATGATATCTATTTCTTCGCTTGGTTGTACAACGGTACCAAGCCA
20 1081 ---+-----+-----+-----+-----+
1140

GTGCTGTTGTGATACTATAGATAAAAGAAGCGAAACCAAACATGTTGCCATGGTTCGGT

25 L S T T S V E S I E E T D G Y S A S W T
400

TTGTCTACTACTCTGTTGAATCTATTGAAGAAACTGACGGTTACTCTGCTTCTGGACT
1141 ---+-----+-----+-----+-----+
1200

AACAGATGATGAAGACAACCTAGATAACTCTTGACTGCCAATGAGACGAAGAACCTGA
30 V P F A A R A Y V E M M Q C Q A E K E P
420

GTTCCATTGCTGCTAGAGCTACGTTGAAATGATGCAATGTCAAGCTGAAAAGGAACCA
1201 ---+-----+-----+-----+-----+
35 1260

CAAGGTAAGCGACGATCTGAATGCAACTTACTACGTTACAGTTGACTTTCCCTGGT

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440 L V R V L V N D R V V P L H G C A V D K

TTGGTTAGAGTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTGCTGTTGACAAG

5 1261 ---+-----+-----+-----+-----+-----+
1320

AACCAATCTAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACACGACAACGTTC

10 460 L G R C K R D D F V E G L S F A R S G G

TTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTGTCTTCGCTAGATCTGGTGGT

1321 ---+-----+-----+-----+-----+-----+
1380

AACCCATCTACATTCTCTCTGCTGAAGCAACTTCAAACAGAAAGCGATCTAGACCACCA

15 N W A E C F A * 467

AACTGGGCTGAATGTTCGCTTAA

1381 ---+-----+-----+ 1410

TTGACCCGACTTACAAAGCGAATT

20

25

30

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5

Figure 8

10 M G V F V V L L S I A T L F G S T S G T
20 ATGGGCGTGGTCTCGTGTGCTACTGTCCATTGCCACCTGTTGGTCCACATCCGGTACC
15 60 1 -----+-----+-----+-----+-----+-----+
TACCCGCACAAGCAGCAGGATGACAGGTAACGGTGGAACAGCCAAGGTGTAGGCCATGG
40 A L G P R G N S H S C D T V D G G Y Q C
20 GCCTTGGGTCTCGGTAACCTCTCACTCTGTGACACTGTTGACGGTGGTACCAATGT
61 120 61 -----+-----+-----+-----+-----+-----+
CGGAACCCAGGAGCACCATTGAGAGTGAGAACACTGTGACAACGTGCCACCAATGGTTACA
25 A F P E I S H L W G T Y S P F F S L A D E
60 TTCCCAGAAATTCTCACTTGTGGGTACATACTCTCCATTCTCTCTGGCTGACGAA
121 180 121 -----+-----+-----+-----+-----+-----+
AAGGGTCTTAAAGAGTGAACACCCATGTATGAGAGGTAAGAAGAGAAACCGACTGCTT

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80 S A I S P D V P K G C R V T F V Q V L S
 80
 TCTGCTATTCAGACGTTCAAAGGGTTGAGAGTTACTTCGTTCAAGTTGTCT
 5 181 -----+-----+-----+-----+-----+-----+
 240
 AGACGATAAAAGAGGTCTGCAAGGTTCCAACATCTCAATGAAAGCAAGTTCAAAACAGA
 10 100 R H G A R Y P T S S A S K A Y S A L I E
 10 100
 AGACACGGTGCTAGATAACCAACTTCTCGGTCTAAGCGTACTCTGCTTGATTGAA
 241 -----+-----+-----+-----+-----+-----+
 300
 TCTGTGCCACGATCTATGGTTGAAGAAGACGCAGATTCCGATGAGACGAAACTAACCT
 15 A I Q K N A T A F K G K Y A F L K T Y N
 120
 GCTATTCAAAAGAACGCTACTGCTTCAGGGTAAGTACGCTTCTTGAAGACTTACAAC
 20 360 301 -----+-----+-----+-----+-----+
 20 360
 CGATAAGTTTCTTGCATGACGAAAGTCCCATTGCGAAAGAACTTCTGAATGTTG
 A Y T L G A D D L T P F G E Q Q M V N S G
 140
 25 TACACTTGGGTGCTGACGACTTGACTCCATTGGTGAACAACAAATGGTTAACTCTGGT
 361 -----+-----+-----+-----+-----+
 420
 420
 ATGTGAAACCCACGACTGCTGAACGTGAGGTAAGCCACTTGTGTTACCAATTGAGACCA
 30 I K F Y R R Y K A L A R K I V P F I R A
 160
 ATTAAGTTCTACAGAAGATAACAGGCTTGGCTAGAAAGATTGTTCCATTAGAGCT
 421 -----+-----+-----+-----+-----+
 480
 480
 35 TAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTCTAACAGGTAAAGTAATCTCGA
 180 S G S D R V I A S A E K F I E G F Q S A

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TCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCAATTGAAGGTTCCAATCTGCT
 481 -----+-----+-----+-----+-----+-----+-----+
 540
 AGACCAAGACTGTCTCAATAACGAAGACGACTTTCAAGTAACCTCCAAAGGTTAGACGA
 5
 K L A D P G A N P H Q A S P V I N V I I
 200
 AAGTTGGCTGACCCAGGTGCTAACCCACACCAAGCTTCTCCAGTTATTAAACGTTATTATT
 541 -----+-----+-----+-----+-----+-----+-----+
 10 600
 TTCAACCGACTGGGTCCACGATTGGGTGTGGTTCGAAGAGGTCAATAATTGCAATAATAA
 220
 P E G A G Y N N T L D H G L C T A F E E E
 15
 CCAGAAGGTGCTGGTTACAACAAACACTTGGACCACGGTTGTGTACTGCTTCGAAGAA
 601 -----+-----+-----+-----+-----+-----+
 660
 GGTCTTCCACGACCAATGTTGTTGAAACCTGGTGCACACATGACGAAAGCTTCTT
 20
 S E L G D D V E A N F T A V F A P P I R
 240
 TCTGAATTGGGTGACGACGTTGAAGCTAACCTCACTGCTGTTTCGCTCCACCAATTAGA
 661 -----+-----+-----+-----+-----+-----+
 720
 25
 AGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGACAAAAGCGAGGTGGTTAATCT
 260
 A R L E A H L P G V N L T D E D V V N L
 721 -----+-----+-----+-----+-----+-----+
 780
 GCTAGATTGGAAGCTCACCGCCAGGTGTTAACCTGACTGACGAAGACGTTGTTAACCTG
 30
 CGATCTAACCTTCGAGTGAAACGGTCCACAATTGAACTGACTGCTTCTGCAACAAATTGAAAC
 280
 M D M C P F D T V A R T S D A T Q L S P
 35
 ATGGACATGTGTCCATTGACACTGTTGCTAGAACCTCTGACGCTACTCAATTGTCTCCA

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781 -----+-----+-----+-----+-----+
 840

TACCTGTACACAGGTAAGCTGTGACAACGATCTGAAGACTGCGATGAGTTAACAGAGGT

5 F C D L F T H D E W I Q Y D Y L Q S L G
 300

TTCTGTGACTTGTTCACTCACGACGAATGGATTCAATACGACTACTTGCAATCTTGGGT

841 -----+-----+-----+-----+-----+
 900

AAGACACTGAACAAGTGAGTGCTGCTTACCTAACGTTATGCTGATGAACGTTAGAAACCCA

320 K Y Y G Y G A G N P L G P A Q G V G F V

AAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTGTTGGTTCGTT

15 901 -----+-----+-----+-----+-----+
 960

TTCATGATGCCAATGCCACGACCATTGGTAACCCAGGTCGAGTTCCACAACCAAAGCAA

20 340 N E L I A R L T H S P V Q D H T S T N H

AACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTTCTACTAACAC

961 -----+-----+-----+-----+-----+
 1020

TTGCTTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAAGATGATTGGTG

25 T L D S N P A T F P L N A T L Y A D F S
 360

ACTTTGGACTCTAACCCAGCTACTTCCCATTGAACGCTACTTGTACGCTGACTTCTCT

1021 -----+-----+-----+-----+-----+
 30 1080

TGAAACCTGAGATTGGTCGATGAAAGGGTAACTTGCGATGAAACATGCGACTGAAGAGA

380 H D N T M V S I F F A L G L Y N G T K P

35 CACGACAAACACTATGGTTCTATTTCTTCGCTTGGTTGTACAACGGTACTAACCCA

1081 -----+-----+-----+-----+-----+
 1140

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GTGCTGTTGTGATACCAAAGATAAAAAGAAGCGAAACCCAAACATGTTGCCATGATTCGGT

400 L S T T S V E S I E E T D G Y S A S W T

5 TTGTCTACTACTTCTGTTGAATCTATTGAAGAAACTGACGGTTACTCTGCTCTTGGACT

1141 -----+-----+-----+-----+-----+-----+
1200

AACAGATGATGAAGACAACTTAGATAACTCTTTGACTGCCAATGAGACGAAGAACCTGA

10 V P F A A R A Y V E M M Q C E A E K E P
420

GTTCCATTGCTGCTAGAGCTTACGTTGAAATGATGCAATGTGAAGCTGAAAAGGAACCA

1201 -----+-----+-----+-----+-----+-----+
1260

15 CAAGGTAAGCGACGATCTGAATGCAACTTACTACGTTACACTTCGACTTTCCCTGGT

440 L V R V L V N D R V V P L H G C G V D K

TTGGTTAGAGTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTGGTGGTACAAG

20 1261 -----+-----+-----+-----+-----+
1320

AACCAATCTCAAAACCAATTGCTGTCTAACAAAGGTAACGTGCCAACACCACAACTGTT

25 460 L G R C K R D D F V E G L S F A R S G G

TTGGGTAGATGTAAGAGAGACCGACTCGTTGAAGGTTGTCTTCGCTAGATCTGGTGGT

1321 -----+-----+-----+-----+-----+
1380

AACCCATCTACATTCTCTGCTGAAGCAACTCCAAACAGAAAGCGATCTAGACCACCA

30 N W E E C F A * 467

AACTGGGAAGAATGTTCGCTTAA

1381 -----+-----+----- 1404

TTGACCCCTTCTTACAAAGCGAATT

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10

15

Figure 9

20 M G V F V V L L S I A T L F G S T S G T 20
ATGGGGGTTTCGTCGTTCTATTATCTATCGCGACTCTGTTGGCAGCACATGGGCACT
1 -----+-----+-----+-----+-----+-----+-----+ 60
TACCCCCAAAGCAGCAAGATAATAGATAGCGCTGAGACAAGCCGTCGTAGCCGTGA

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A L G P R G N H S K S C D T V D L G Y Q 40
 GCGCTGGGCCCGTGGAAATCACTCCAAGTCCTGCGATACGGTAGACCTAGGGTACCAAG
 61 -----+-----+-----+-----+-----+ 120
 5 CGCGACCCGGGGGACCTTAGTGAGGTCAGGACGCTATGCCATCTGGATCCATGGTC

 C S P A T S H L W G T Y S P X F S L E D 60
 TGCTCCCTGCGACTTCTCATCTATGGGCACGTACTGCCATaCTTTGCTCGAGGAC
 121 -----+-----+-----+-----+-----+ 180
 10 ACGAGGGGACGCTGAAGAGTAGATAACCCCGtgCATGAGGGTAtGAAAAGCGAGCTCCTG

 E L S V S S K L P K D C R I T L V Q V L 80
 GAGCTGTCCGTGTCGAGTAAGCTTCCAAGGATTGCCGGATCACCTTGGTACAGGTGCTA
 181 -----+-----+-----+-----+-----+ 240
 15 CTCGACAGGCACAGCTCATTGAAAGGTTCTAACGGCTAGTGGAACCATGTCCACGAT

 S R H G A R Y P T S S K S K K Y K K L I 100
 TCGGCCATGGAGCGCGGTACCCAACCAGCTCCAAGAGCAAAAGTATAAGAAGCTTaTt
 241 -----+-----+-----+-----+-----+ 300
 20 AGCGCGGTACCTCGGCCATGGTTGGTCGAGGTTCTCGTTTTCATATTCTCGAAAtAa

 T A I Q A N A T D F K G K Y A F L K T Y 120
 ACGGCGATCCAGGCCAATGCCACCGACTTCAAGGGCAAGTAcGCCTTTTGAAAGACGTAC
 301 -----+-----+-----+-----+-----+ 360
 25 TGCCGCTAGGTCCGGTTACGGTGGCTGAAGTTCCGTTCAAGGGAAACCCCTCGTCGACCTTGAGC

 N Y T L G A D D L T P F G E Q Q L V N S 140
 AACTATACTCTGGGTGCGGATGACCTCACTCCCTTGGGAGCAGCAGCTGGTGAACCTCG
 361 -----+-----+-----+-----+-----+ 420
 30 TTGATATGAGACCCACGCCACTGGAGTGAGGGAAACCCCTCGTCGACCTTGAGC

 G I K F Y Q R Y K A L A R S V V P F I R 160
 GGCATCAAGTTCTACCAGAGGTACAAGGCTCTGGCGCGAGTGTGGTGCCGTTATTCCG

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421 -----+-----+-----+-----+-----+-----+ 480

CCGTAGTTCAAGATGGTCTCCATGTTCCGAGACCGCGCGTCACACCACGGCAAATAAGCG

A S G S D R V I A S G E K F I E G F Q Q 180

5 GCCTCAGGCTCGGACCGGGTTATTGCTTCGGAGAGAAGTTCATCGAGGGGTTCCAGCAG

481 -----+-----+-----+-----+-----+-----+ 540

CGGAGTCCGAGCCTGGCCAATAACGAAGCCCTCTCTCAAGTAGCTAGCTCCCAAGGTCGTC

A K L A D P G A T N R A A P A I S V I I 200

10 GCAGAGCTGGCTGATCCTGGCGCGACGAACCGCGCCGCTCCGGCGATTAGTGTGATTATT

541 -----+-----+-----+-----+-----+-----+ 600

CGCTTCGACCGACTAGGACCGCGCTGCTTGGCGCGGCGAGGCCGCTAACACACTAATAAA

P E S E T F N N T L D H G V C T K F E A 220

15 CCGGAGAGCGAGACGTTCAACAATACGCTGGACCACGGTGTGTGACAGAAGTTGAGGCG

601 -----+-----+-----+-----+-----+-----+ 660

GGCCTCTCGCTCTGCAAGTTGTTATGCGACCTGGTGCCACACACGTGCTTCAAACCTCCG

S Q L G D E V A A N F T A L F A P D I R 240

20 AGTCAGCTGGGAGATGAGGTTGCGGCCATTCACTGCGCTTTGCACCCGACATCCGA

661 -----+-----+-----+-----+-----+-----+ 720

TCAGTCGACCCCTACTCCAACGCCGGTAAAGTGACGCGAGAACGTGGCTGTAGGCT

A R L E K H L P G V T L T D E D V V S L 260

25 GCTCGCctCGAGAACGATCTTCCCTGGCGTGACGCTGACAGACGAGGACGTTGTCAGTCTA

721 -----+-----+-----+-----+-----+-----+ 780

CGAGCGGgaGCTCTCGTAGAAGGACCGCACTGCGACTGTCTGCTCCTGCAACAGTCAGAT

M D M C P F D T V A R T S D A S Q L S P 280

30 ATGGACATGTGTcCGTTGATACGGTAGCGCGACCAGCGACGCAAGTCAGCTGTACCG

781 -----+-----+-----+-----+-----+-----+ 840

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TACCTGTACACAgGCAAACATGCCATCGCGCTGGTCGCTGCGTTAGTCGACAGTGGC

F C Q L F T H N E W K K Y D Y L Q S L G 300

TTCTGTCAACTCTCACTCACAATGAGTGGAAAGAAGTACgACTACCTTCAGTCCTGGC

5 841 -----+-----+-----+-----+-----+-----+ 900

AAGACAGTTGAGAAGTGAGTGTACTCACCTTCTTCATGcTGATGGAAGTCAGGAACCCG

K Y Y G Y G A G N P L G P A Q G I G F T 320

AAGTACTACGGCTACGGCGCAGGCAACCCCTGGGACCGGCTCAGGGGATAGGGTTCA

10 901 -----+-----+-----+-----+-----+-----+ 960

TTCATGATGCCGATGCCGCTCCGTTGGGAGACCCCTGGCGAGTCCCCTATCCCAAGTGG

N E L I A R L T R S P V Q D H T S T N S

340

15 AACGAGCTGATTGCCCGGTTGACgCGTTGCCAGTGCAGGACCACACCAGCACTAACTCG

961 -----+-----+-----+-----+-----+-----+ 1020

TTGCTCGACTAACGGCCAATGcGCAAGCGGTACGTCTGGTGTGGTCGTGATTGAGC

20 360 T L V S N P A T F P L N A T M Y V D F S

ACTCTAGTCTCCAACCCGGCACCTCCGTTGAACGCTACCATGTACGTGACTTTCA

1021 -----+-----+-----+-----+-----+ 1080

TGAGATCAGAGGTTGGCCGGTGGAAAGGCAACTTGCATGGTACATGCAGCTGAAAAGT

380 H D N S M V S I F F A L G L Y N G T E P

CACGACAAACAGCATGGTTCCATCTCTTGCAATTGGCCTGTACAACGGCACTGAACCC

30 1081 -----+-----+-----+-----+-----+ 1140

GTGCTGTTGTCGTACCAAAAGGTAGAAGAACGTAACCCGGACATGTTGCCGTGACTTGGG

35 400 L S R T S V E S A K E L D G Y S A S W V

- 104 -

TTGTCCCGGACCTCGGTGGAAAGGCCAAGGAATTGGATGGGTATTCTGCATCCTGGGTG

1141 -----+-----+-----+-----+-----+
1200

AACAGGGCCTGGAGCCACCTTCGCGGTTCTTAACCTACCCATAAGACGTAGGACCCAC

5
V P F G A R A Y F E T M Q C K S E K E P
420GTGCCTTCGGCGCGAGCCTACTT^{CG}AGACGATGCAATGCAAGTCGGAAAAGGAGCCT1201 -----+-----+-----+-----+-----+
10 1260

CACGGAAAGCCGCGCTCGGATGAAGCTCTGCTACGTTACGTTAGCCTTTCTCGGA

L V R A L I N D R V V P L H G C D V D K
440

15 CTTGTTCGCGCTTGATTAATGACCGGGTTGTGCCACTGCATGGCTGCATGTGGACAAG

1261 -----+-----+-----+-----+-----+
1320

GAACAAGCGCGAAACTAATTACTGGCCAACACGGTGACGTACCGACGCTACACCTGTC

20 L G R C K L N D F V K G L S W A R S G G
460

CTGGGGCGATGCAAGCTGAATGACTTTGTCAAGGGATTGAGTTGGCCAGATCTGGGGC

1321 -----+-----+-----+-----+-----+
1380

25 GACCCCGCTACGTTGACTTACTGAAACAGTTCCCTAACTCAACCCGGTCTAGACCCCCG

N W G E C F S * 467

AACTGGGGAGAGTGCTTAGTTGA

1381 -----+-----+----- 1404

30 TTGACCCCTCTCACGAAATCAACT

Figure 10

5 CP-1

Eco RI M G V F V V L L S I A T L F G S T

TATATGAATTCATGGCGTGTCGTGTACTGTCCATTGCCACCTTGTTCGGTCCA

1 1-----+-----+-----+-----+-----+-----+-----+-----+ 60

ATATACTTAAGTACCCGACAAGCAGCACGATGACAGGTAACGGTGGAACAGCCAAGGT

10

S G T A L G P R G N S H S C D T V D G G

CATCCGGTACCGCCTTGGGTCTCGTGGTAATTCTCACTCTGTGACACTGTTGACGGTG

61 61-----+-----+-----+-----+-----+-----+-----+-----+ 120

GTAGGCCATGGCGAACCCAGGAGCACCATTAAGAGTGAGAACACTGTGACAACGTGCCAC

15 CP-2 CP-3

Y Q C F P E I S H L W G Q Y S P Y F S L

GTTACCAATGTTCCCAGAAATTCTCACTTGTGGGTCAATACTCTCCATACTCTCTT

121 121-----+-----+-----+-----+-----+-----+-----+ 180

CAATGGTTACAAAGGGTCTTAAAGAGTGAAACACCCAGTTATGAGAGGTATGAAGAGAA

20

E D E S A I S P D V P D D C R V T F V Q

TGGAAGACGAATCTGCTATTCAGACGTTCCAGACGACTGTAGAGTTACTTCGTT

181 181-----+-----+-----+-----+-----+-----+-----+ 240

ACCTTCTGCTTAGACGATAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAG

25 CP-4.7

CP-5.7

V L S R H G A R Y P T D S K G K K Y S A

AAGTTTGTCTAGACACGGTGTAGATAACCAACTgactCTAAGggtaAGaagTACTCTG

241 241-----+-----+-----+-----+-----+-----+-----+ 300

TTCAAAACAGATCTGTGCCACGATCTATGGGTTGActgAGATTCCcaATTCTtcATGAGAC

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L I E A I Q K N A T A F K G K Y A F L K
CTTTGATTGAAGCTATTCAAAAAGAACGCTACTGCTTCAGGGTAAGTACGCTTCTTGA
301 -----+-----+-----+-----+-----+-----+ 360

5 GAAACTAACCTCGATAAGTTCTTGCATGACGAAAGTTCCATTGCGAAAGAACT
CP-6
CP-7

T Y N Y T L G A D D L T P F G E N Q M V
AGACTTACAACACTACACTGGGTGCTGACGACTTGACTCCATTGGTAAAACCAAATGG
10 361 -----+-----+-----+-----+-----+-----+ 420
TCTGAATGTTGATGTGAAACCCACGACTGCTGAAC TGAGGTAAGCCACTTTGGTTACC

N S G I K F Y R R Y K A L A R K I V P F
TTAACCTGGTATTAAGTTCTACAGAAGATAAAGGCTTGCTAGAAAGATTGTTCCAT
15 421 -----+-----+-----+-----+-----+-----+ 480
AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCAAACCGATCTTCTAACAGGT
CP-8.7

CP-9

I R A S G S S R V I A S A E K F I E G F
TCATTAGAGCTCTGGTTCTtctAGAGTTATTGCTTCTGCTGAAAAGTTCAATTGAAGGTT
20 481 -----+-----+-----+-----+-----+-----+ 540
AGTAATCTCGAAGACCAAGAagaTCTCAATAACGAAGACGACTTTCAAGTAACCTCCAA

Q S A K L A D P G S Q P H Q A S P V I D
TCCAATCTGCTAAGTTGGCTGACCCAGGGTCTCAACCCACACCAAGCTTCTCCAGTTATTG
25 541 -----+-----+-----+-----+-----+-----+ 600
AGGTTAGACGATTCAACCGACTGGGTCCAAGAGTTGGTGTGGTCAAGAGGGTCAATAAC
CP-10.7
CP-11.7

V I I S E A S S Y N N T L D P G T C T A
ACGTTATTATTtctGAcgctTCTtctTACAACAAACACTTGGACccAGGTACTTGTACTG
30 601 -----+-----+-----+-----+-----+-----+ 660

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TGCAATAATAAagaCTgcgaAGGagaATGTTGTTGTGAAACCTGggtCCATGAACATGAC

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F E D S E L A D T V E A N F T A L F A P

CTTTCGAAGACTCTGAATTGgctGACactGTTGAAGCTAACTTCACTGCTTGTTCGCTC

661 -----+-----+-----+-----+-----+-----+ 720

GAAAGCTTCTGAGACTTAACcgactGtgaCAACTCGATTGAAGTGACGAAACAAGCGAG

5

CP-12.7

A I R A R L E A D L P G V T L T D T E V

CAGCTATTAGAGCTAGATTGGAAGCTGACTTGCAGGTGTTACTTGACTGACactgaaG

721 -----+-----+-----+-----+-----+-----+ 780

10 GTCGATAATCTCGATCTAACCTCGACTGAACGGTCACAATGAAACTGACTGtgaacttc

CP-13.7

T Y L M D M C S F E T V A R T S D A T E

TTactTACCTGATGGACATGTGTTctTTCGAAACTGTTGCTAGAACTTCTGACGCTACTG

15 781 -----+-----+-----+-----+-----+-----+ 840

AAtgaATGAACCTACCTGTACACAagaAAGCTTGACAAACGATCTGAAGACTGCGATGAC

L S P F C A L F T H D E W R H Y D Y L Q

AATTGTCTCCATTCTGTGCTTGTTCACTCACGACGAATGGAGACacTACGACTACTGC

20 841 -----+-----+-----+-----+-----+-----+ 900

TTAACAGAGGTAAGACACGAAACAAAGTGAGTGCTGCTTACCTCTgtgATGCTGATGAACG

CP-14.7

CP-15.7

S L K K Y Y G H G A G N P L G P T Q G V

25 AATCTTGAAGAAGTACTACGGTcacGGTGCTGGTAACCCATTGGGTCCAactCAAGGTG

901 -----+-----+-----+-----+-----+-----+ 960

TTAGAAAActtcTTCATGATGCCAgtgCCACGACCATTGGGTAAACCCAGGTTgaGTTCCAC

G F A N E L I A R L T R S P V Q D H T S

30 TTGGTTTCGCTAACGAATTGATTGCTAGATTGACTAGATCTCCAGTTCAAGACCACATT

961 -----+-----+-----+-----+-----+-----+ 1020

AACCAAAGCGATTGCTTAACTAACGATCTAACTGATCTAGAGGTCAAGTTCTGGTGTGAA

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CP-16
CP-17.7

T N H T L D S N P A T F P L N A T L Y A
CTACTAACACACTTGGACTCTAACCCAGCTACTTCCCATTGAACGCTACTTGTACG

5 1021 -----+-----+-----+-----+-----+
1080

GATGATTGGTGTGAAACCTGAGATTGGGTCGATGAAAGGTAACTTGCGATGAAACATGC

D F S H D N G I I S I F F A L G L Y N G
10 CTGACTTCTCTCACGACAAACggtattATTCTATTTCTCGCTTGGTTGTACAACG

1081 -----+-----+-----+-----+-----+
1140

GACTGAAGAGAGTGCTGTTGccataaTAAAGATAAAAGAAGCGAAACCAAACATGTTGC

CP-18.7
CP-19.7

T A P L S T T S V E S I E E T D G Y S S
GTACTGCTCCATTGCTACTACTTCTGTTGAATCTATTGAAGAAACTGACGGTTACTCT

1141 -----+-----+-----+-----+-----+
1200

20 CATGACGAGGTAACAGATGATGAAGACAACTTAGATAACTTCTTGACTGCCAATGAGAa

A W T V P F A S R A Y V E M M Q C Q A E

ctgctTGGACTGTTCCATTGcttctAGAGCTTACGTTGAAATGATGCAATGTCAAGCTG
1201 -----+-----+-----+-----+-----+
25 1260

gacgaACCTGACAAGGTAAGcgaagaTCTCGAATGCAACTTACTACGTTACAGTTGAC

CP-20
CP-21

K E P L V R V L V N D R V V P L H G C A
30 AAAAGGAACCATTGGTTAGAGTTGGTAAACGACAGAGTTGTTCCATTGCACGGTTGTG

1261 -----+-----+-----+-----+-----+
1320

TTTCCCTGGTAACCAATCTCAAAACCAATTGCTGTCACAAAGGTAACGTGCCAACAC

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V D K L G R C K R D D F V E G L S F A R

CTGTTGACAAGTTGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTGTCTTCGCTA

1321 -----+-----+-----+-----+-----+-----+
1380

5 GACAACGTGTTCAACCCATCTACATTCTCTGCTGAAGCAACTTCAAACAGAAAGCGAT

CP-22

S G G N W A E C F A * Eco RI

GATCTGGTGGTAACTGGGCTGAATGTTCGCTTAAGAATTCAATATA

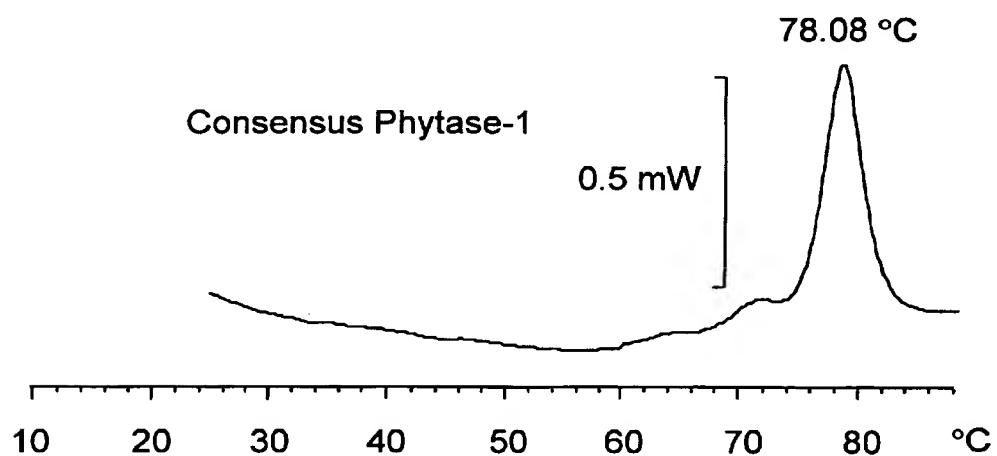
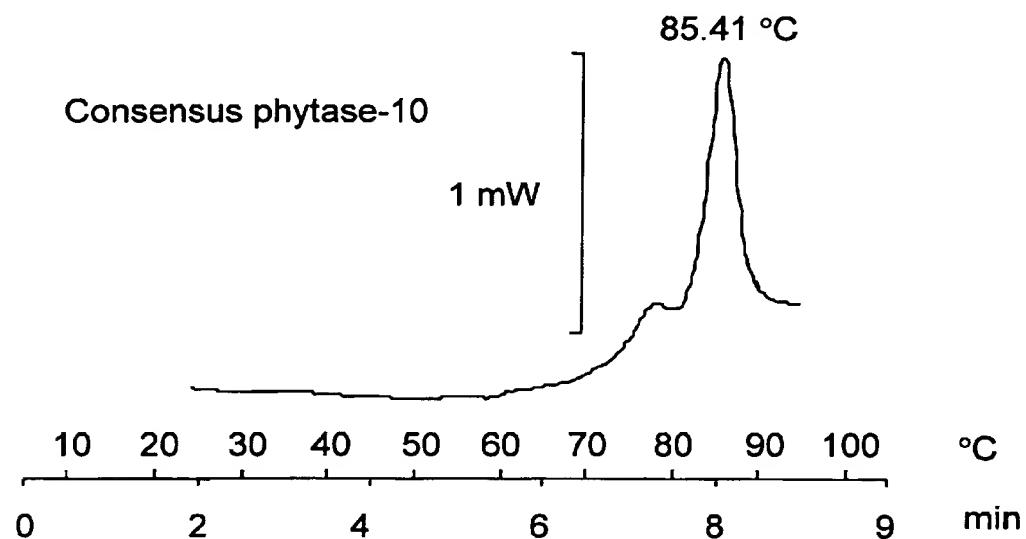
1381 -----+-----+-----+-----+-----+----- 1426

10 CTAGACCACCATTGACCCGACTTACAAAGCGAATTCTTAAGTATAT

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Figure 11



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Figure 12

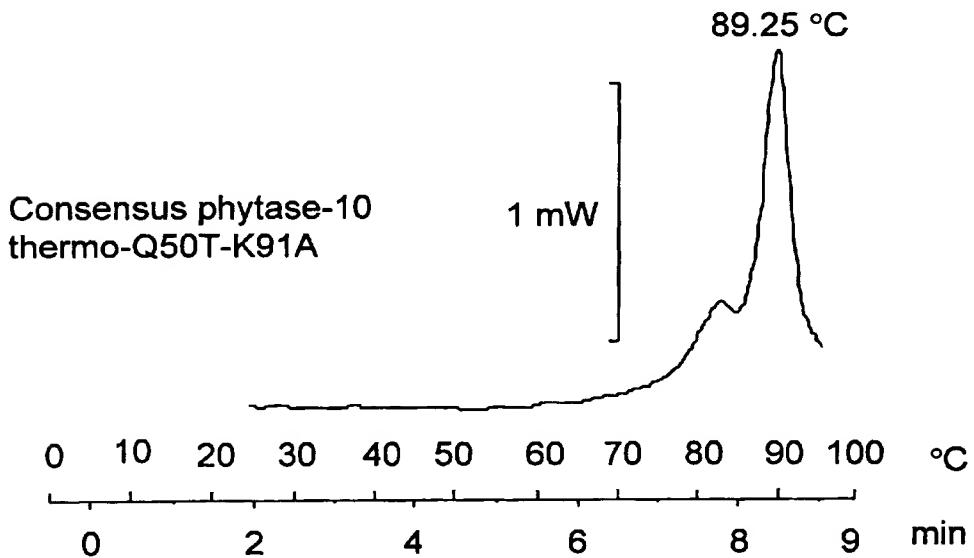
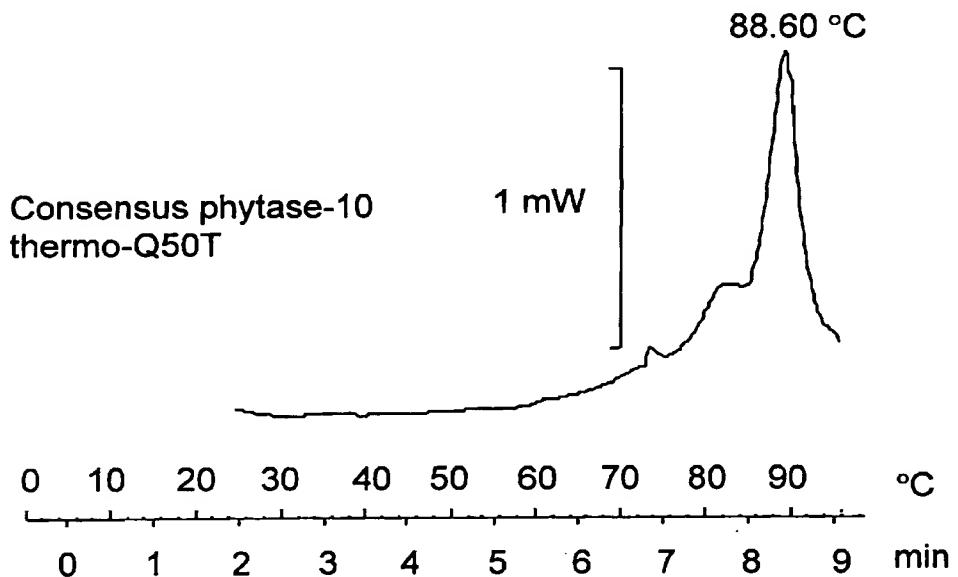


Figure 13

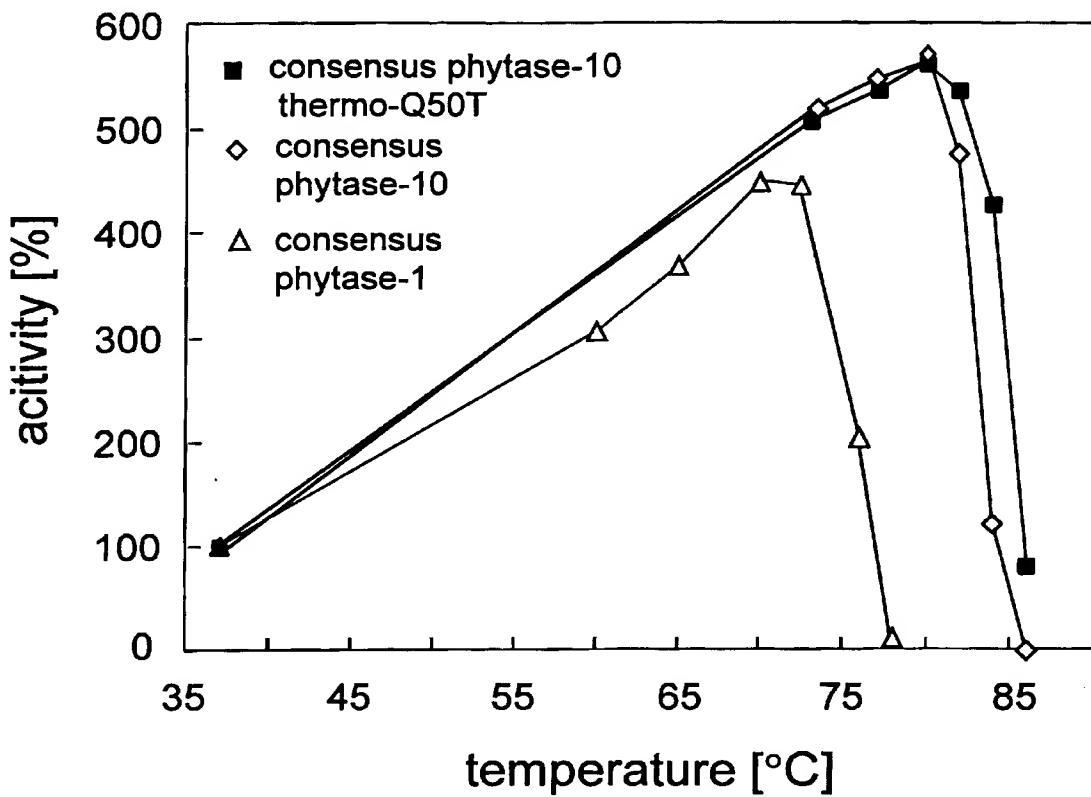


Figure 14

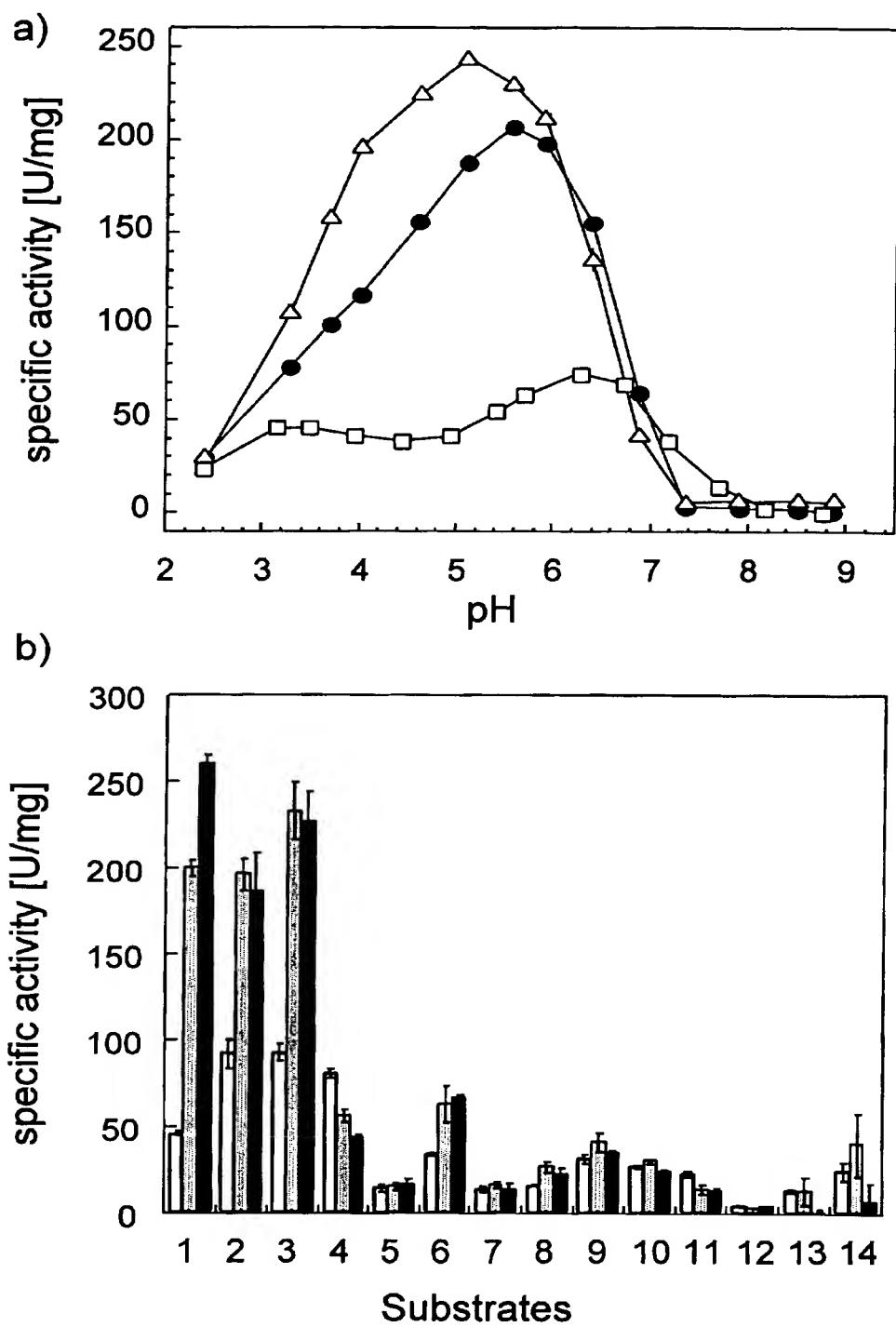


Figure 15

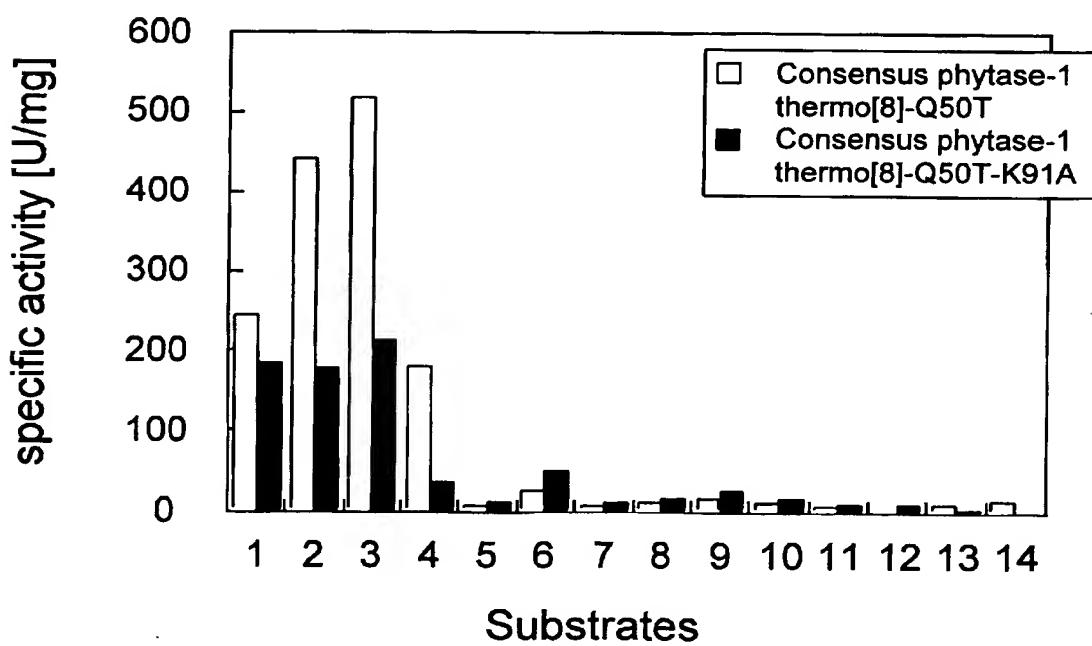
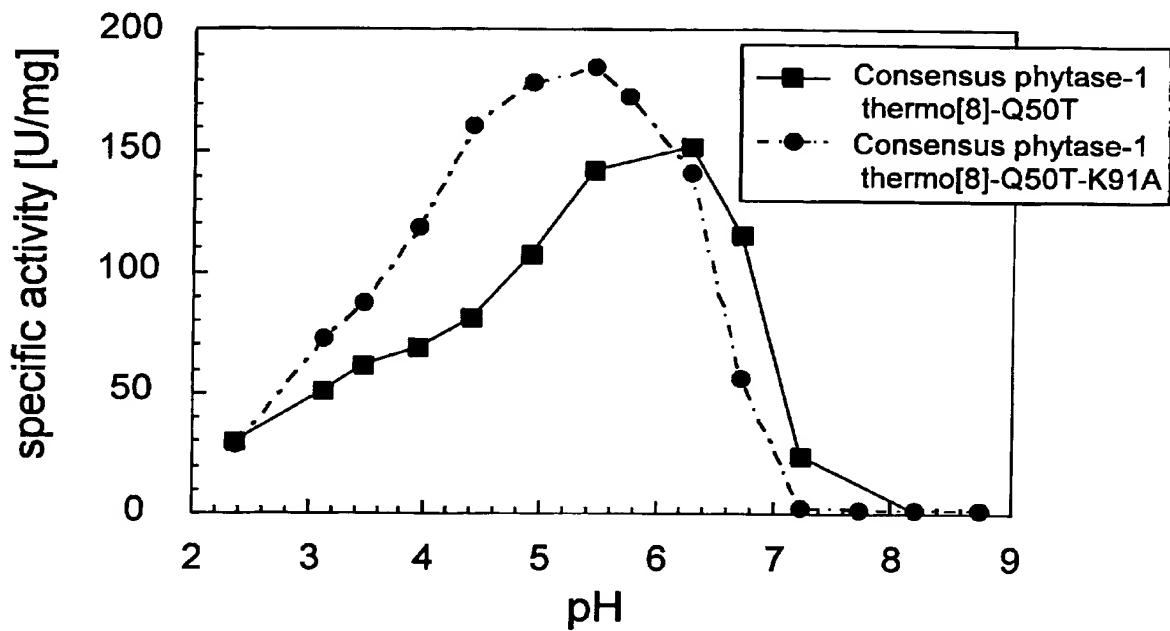


Figure 16

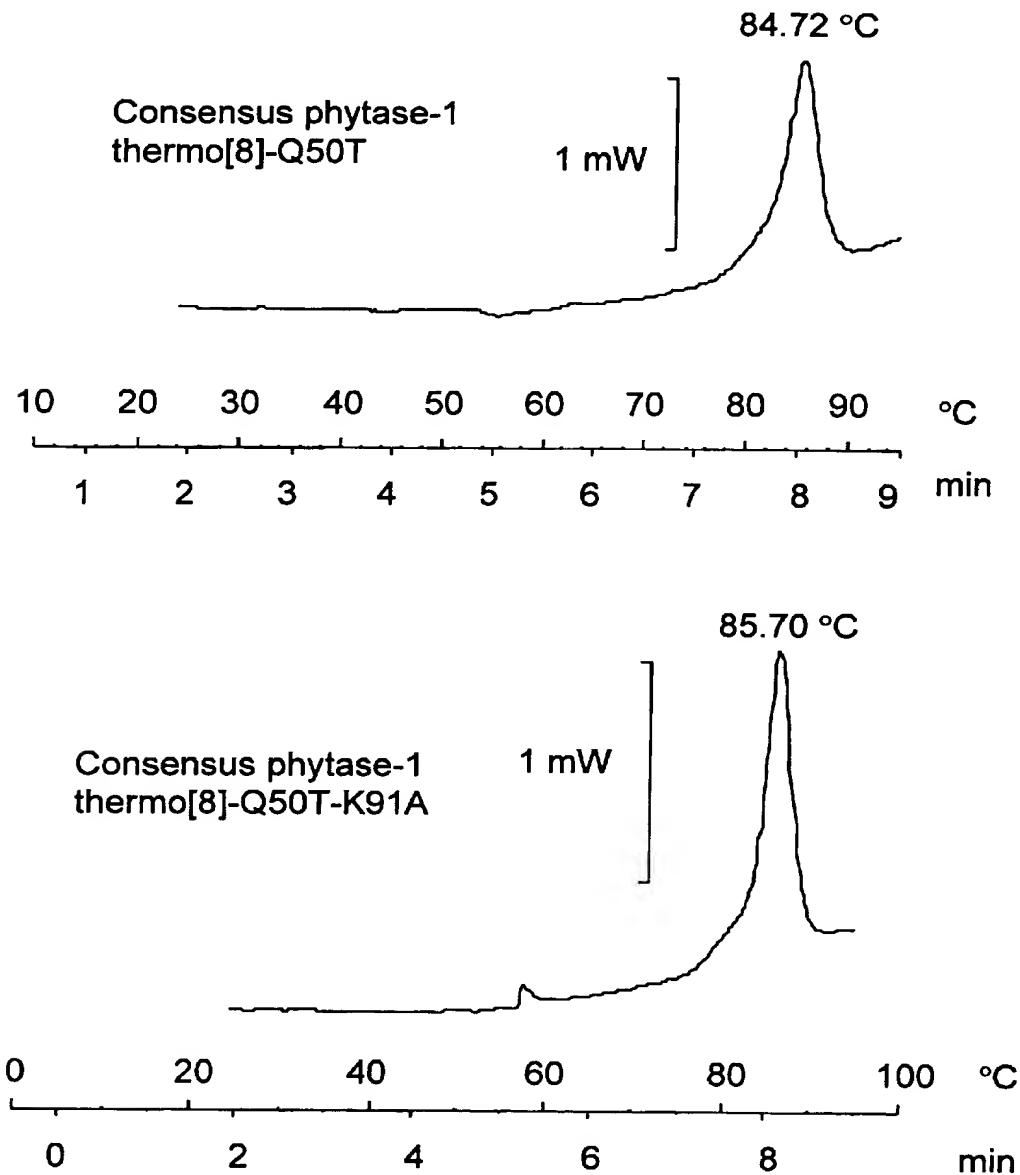


Figure 17

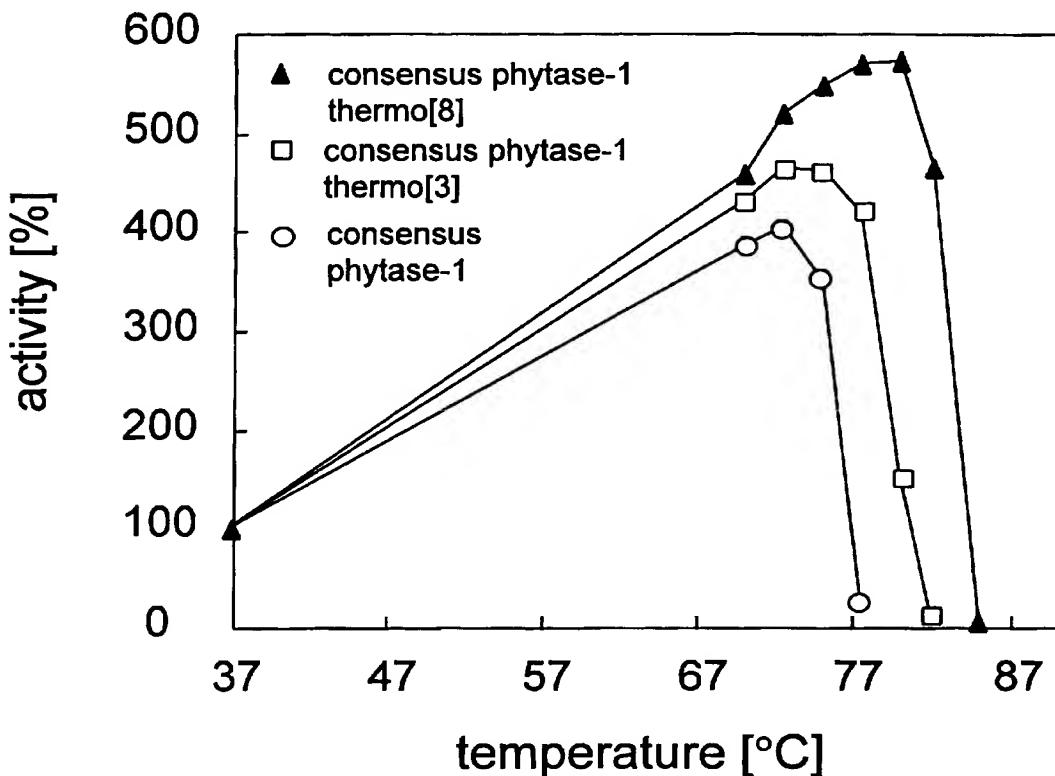
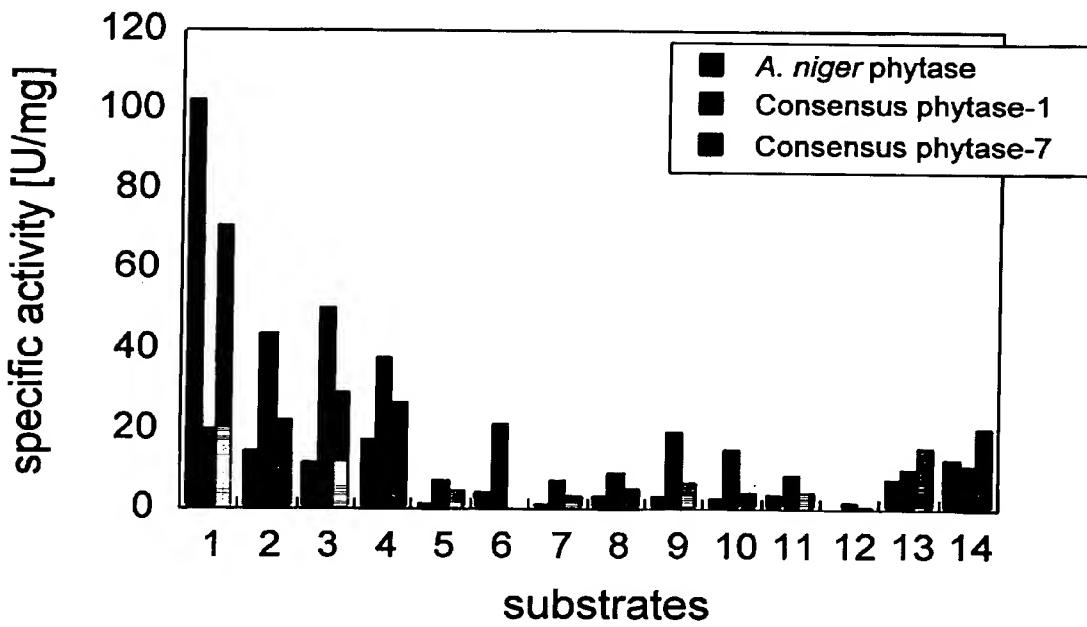
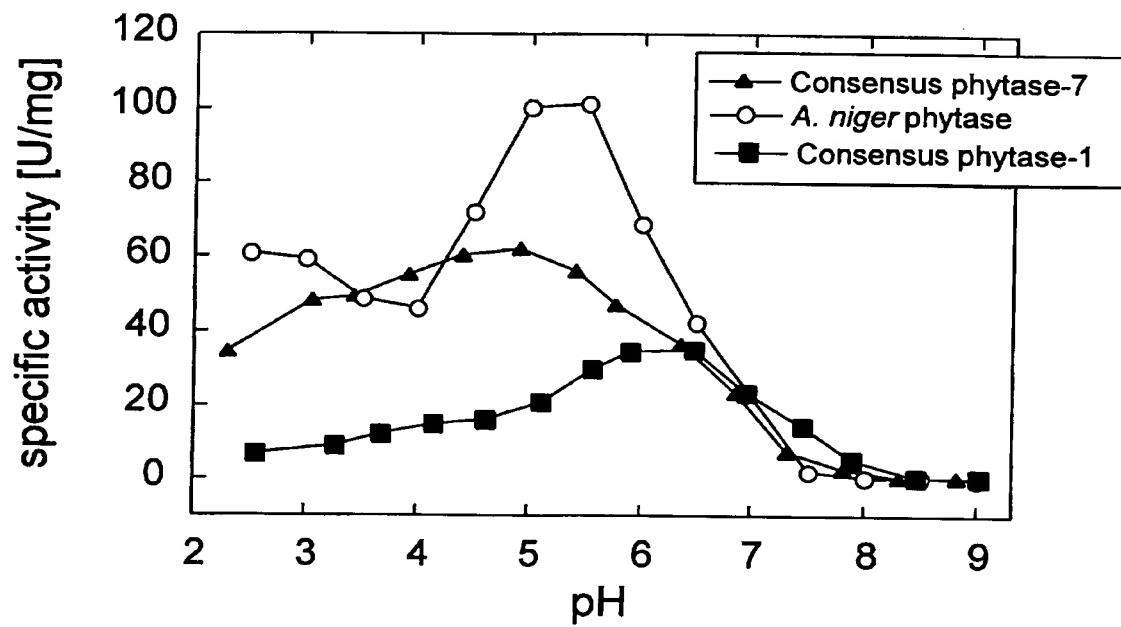


Figure 18



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Figure 19

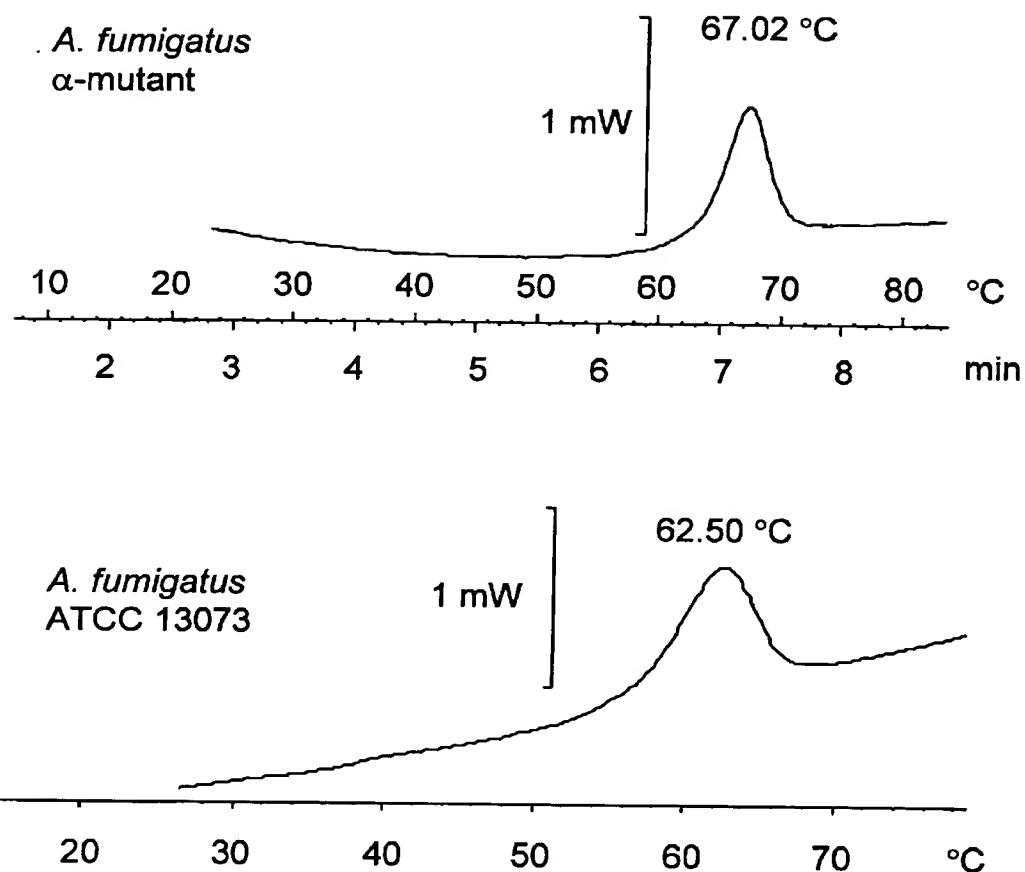
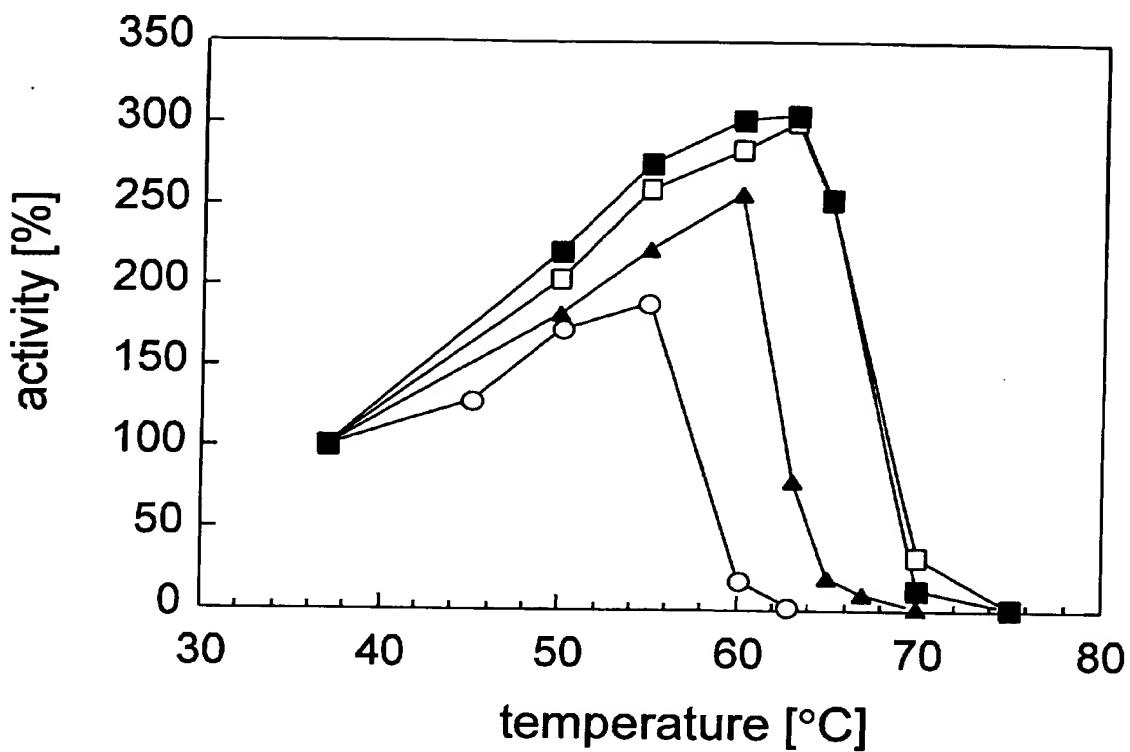


Figure 20



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Figure 21

1 MGVVVLLSI ATLFGSTSGT ALGPRGNSHS CDTVDGGYQC FPEISSNWSP
51 YSPYFSLADE SAISPDVPKG CRVTFVQVLQ RHGARFPTSG AATRISALIE
101 AIQKNATAFK GKYAFLKTYN YTLGADDLYP FGANQSSQAG IKFYRRYKAL
5 151 ARKIVPFIRA SGSDRVIDSA TNWIEGFQSA KLADPGANPH QASPVINVII
201 PEGAGYNNTL DHGLCTAEE SELGDDVEAN FTAVFAPPIR ARLEAHLPGV
251 NLTDEDVVNL MDMCPFDTVA RTSDATELSP FCDLFTHDEW IQYDYLGDLD
301 KYYGTGAGNP LGPAQGVGFV NELIARLTHS PVQDHSTSNH TLDSNPATFP
351 LNATLYADFS HDNTMVAIFF ALGLYNGTKP LSTTSVESIE ETDGYSASWL
10 401 VPFSARMYVE MMQCEAEKEP LVRVLVNDRV VPLHCGVDK LGRCKRDDFV
451 EGLSFARSGG NWEECFA

REC'D -
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Abstract

This invention relates to a new improved consensus phytase by introduction of
5 additional phytase sequences into the sequence alignment and the method of
the introduction process. Furthermore, the invention relates to the transfer of
stabilizing amino acid exchanges found by the new method into homologous
proteins. Furthermore, the invention relates to the replacement of a whole
active site of a phytase. It also relates to the corresponding DNA sequences
10 and its generation, methods to produce such phytases and the use thereof.
